

Isoflavones Made Simple – Genistein’s Agonist Activity for the Beta-Type Estrogen Receptor Mediates Their Health Benefits

Mark F. McCarty, NutriGuard Research, 1051 Hermes Ave., Encinitas, CA 92024

Abstract

Soy isoflavones, the focus of much research and controversy, are often referred to as “weak estrogens”. In fact, genistein is a relatively potent agonist for the recently characterized beta isoform of the estrogen receptor (ERbeta). The low nanomolar serum concentrations of unconjugated free genistein achieved with high-nutritional intakes of soy isoflavones are near the binding affinity of genistein for this receptor, but are about an order of magnitude lower than genistein’s affinity for the “classical” alpha isoform of the estrogen receptor (ERalpha). Moreover, these concentrations are far too low to inhibit tyrosine kinases or topoisomerase II, *in vitro* activities of genistein often cited as potential mediators of its physiological effects. The thesis that these physiological effects are in fact mediated by ERbeta activation provides a satisfying rationale for genistein’s clinical activities. Hepatocytes do not express ERbeta; this explains why soy isoflavones, unlike oral estrogen, neither modify serum lipids nor provoke the prothrombotic effects associated with increased risk for thromboembolic disorders. The lack of uterotrophic activity of soy isoflavones reflects the fact that ERalpha is the exclusive mediator of estrogen’s impact in this regard. Vascular endothelium expresses both ERalpha and ERbeta, each of which has the potential to induce and activate nitric oxide synthase; this may account for the favorable influence of soy isoflavones on endothelial function in postmenopausal women and ovariectomized rats. The ERbeta expressed in osteoblasts may mediate the reported beneficial impact of soy isoflavones on bone metabolism. Suggestive evidence that soy-rich diets decrease prostate cancer risk, accords well with the observation that ERbeta appears to play an antiproliferative role in healthy prostate. In the breast, ERalpha promotes epithelial proliferation, whereas ERbeta has a restraining influence in this regard – consistent with the emerging view that soy isoflavones do not increase breast cancer risk, and possibly may diminish it. Premenopausal women enjoy a relative protection from kidney failure; since ERbeta is an antagonist of TGF- β signaling in mesangial cells, soy isoflavones may have nephroprotective potential. Estrogen also appears to protect women from left ventricular hypertrophy, and recent evidence suggests that this effect is mediated by ERbeta. In conjunction with reports that isoflavones may have a modestly beneficial impact on menopausal symptoms – perhaps reflecting the presence of ERbeta in the hypothalamus – these considerations suggest that soy isoflavone regimens of sufficient potency may represent a safe and moderately effective alternative to HRT in postmenopausal women. Further clinical research is required to characterize the impact of optimal genistein intakes on endothelial and bone function in men. Studies with ERbeta-knockout mice could be helpful for clarifying whether ERbeta does indeed mediate the chief physiological effects of low nanomolar genistein. S-equol, a bacterial metabolite of daidzein, has an affinity for ERbeta nearly as high as that of genistein; whether this compound contributes meaningfully to the physiological efficacy of soy isoflavones in some individuals is still unclear.

Physiological Concentrations of Free Genistein Activate Estrogen Receptor-Beta

The key to understanding the health-protective potential of soy isoflavones may have been provided by Kuiper and colleagues, who first established the existence of a “novel” estrogen receptor, now known as estrogen receptor beta (ERbeta) to distinguish it from the “classical” estrogen receptor alpha (ERalpha).^{1,2} These workers assessed the affinity of these receptors for a range of xenobiotic and phytochemical estrogenic compounds, including the soy isoflavones.^{3,4} They established that genistein has agonist activity for both ERalpha and ERbeta, but that genistein’s affinity for ERbeta is considerably greater; genistein’s affinities for ERbeta and ERalpha were determined to be 8.4 nM and 145 nM, respectively. For daidzein, the corresponding values were 100 nM and 420 nM, indicative of its much lower affinity for these receptors. At saturating concentrations, both genistein and daidzein could interact with either of these receptors to activate transcription from estrogen response elements, at least as effectively as the physiological ligand 17 β -estradiol.

A number of subsequent studies have examined genistein’s comparative abilities to bind to and promote transcription from the two estrogen receptor isoforms.⁵⁻¹² Although the absolute values obtained in these studies differ, genistein’s binding affinity for ERbeta consistently emerges as 7-30-fold greater than its affinity for ERalpha; this is paralleled by genistein’s ability to activate transcription with ERbeta at a lower concentration than with ERalpha. As a rule, ERbeta-mediated transcription is approximately half-maximal at a genistein concentration of 10 nM, whereas ERalpha-mediated transcription is minimal at this concentration, only becoming substantial as genistein rises above 100 nM.

One recent study has examined the impact of isoflavones on the rate at which the ER isoforms bind to the estrogen response element in DNA; they determined the isoflavone concentration which would increase this binding rate by 50%.¹³ For genistein, this value was determined to be 30 nM for ERbeta and 15 μ M for ERalpha – once again indicative of marked selectivity for ERbeta. The corresponding values for daidzein were 350 nM for ERbeta and >300 μ M for ERalpha. For equol – which can be generated from daidzein in the GI tract by bacterial reductive activity – the values were 400 nM and 3.5 μ M.

Some of the key effects of estrogen result not from transcriptional activation at estrogen response elements, but from transcriptional repression of certain promoters that bind NF-kappaB, such as the IL-6 promoter.¹⁴ This latter effect reflects interaction of the activated estrogen receptor with NF-kappaB in a manner that does not entail binding of the estrogen receptor to DNA. Genistein has been shown to activate ERbeta such that it is capable of inhibiting the TNF response element; this effect was half-maximal at a genistein concentration of only 8.5 nM.⁷ In contrast, in cells overexpressing ERalpha, only a moderate transrepression was seen with a genistein concentration of 1 μ M.

Estrogen receptors can also influence the transcriptional activity of AP-1, Sp1, and certain c-AMP response elements, through interactions that do not entail direct binding to

DNA.¹⁵⁻¹⁷ In particular, agonist-activated ERalpha often increases the transcriptional activation mediated by AP-1, possibly by binding to coactivators that interact with fos/jun.^{15;18} In contrast, activated ERbeta has the opposite effect, suppressing AP-1-mediated transcription.^{15;19-22} Since AP-1 exerts various pro-proliferative effects, these findings may help to rationalize the opposing effects of ERalpha and ERbeta on cell proliferation in certain tissues, as cited below. In particular, ERalpha activates the cyclin D1 promoter through its AP-1 and c-AMP response elements, whereas ERbeta has a suppressive effect in this regard.²¹ These considerations suggest that low nanomolar concentrations of genistein may have the potential to exert certain effects opposite to those of the classical estrogen receptor - including anti-proliferative effects.

The relevance of these findings becomes evident when one considers the plasma genistein concentration in subjects who habitually consume a soy-rich diet. Adlercreutz et al. reported that the mean concentration of total genistein (free plus conjugated) was 276 nM in Japanese men; however, only about 4% of this was in free or sulphated form, the balance consisting of the glucuronate conjugate which presumably has limited intracellular access.^{23;24} More recent studies have specifically measured unconjugated plasma isoflavones; one of these found that unconjugated genistein constituted only about 1.1-1.5% of the total plasma pool of genistein – the higher percentage being observed transiently in the first 2 hours after soy ingestion.²⁵ The low content of unconjugated isoflavones is presumed to reflect rapid hepatic glucuronidation of these compounds. Since total serum genistein concentrations of around 1.5 µM are noted during prolonged supplementation with high-physiological doses of genistein,²⁶ this would correspond to an unconjugated genistein concentration of about 20 nM if 1.3% of total genistein were in free form. After several subjects ingested a meal providing 125 g (dry weight) of whole soybeans, free serum genistein rose to about 20-40 nM within about 2 hours, persisting at this level at 8 hours, but returning to baseline after 24 hours.²⁷ About half of the unconjugated genistein in serum is bound to serum proteins, so the effective concentration available to cells may be about 50% of the measured total concentration;²⁸ in other words, the physiological impact of 20 nM serum genistein may be comparable to that observed with 10 nM genistein *in vitro*.

These findings encourage the speculation that high physiological serum levels of free genistein – i.e. those achievable by ingesting a soy-rich diet – will achieve ample activation of ERbeta, but only minimal or modest activation of ERalpha. As we shall see, this thesis appears capable of rationalizing both the safety and the physiological benefits of dietary genistein.

Irrelevance of Other Suspected Effects

Much of the speculation regarding the physiological effects of isoflavones makes reference to *in vitro* studies in which genistein has been shown to inhibit tyrosine kinases or topoisomerase II, or to modulate activation of mitogenic signaling pathways in cultured cells. The effects of genistein on tyrosine kinase or topoisomerase activity require concentrations well into the micromolar range.^{29;30} Similarly, the great majority of studies showing that genistein is a signal modulator in cells have used micromolar

concentrations of this agent. These effects thus have no conceivable relevance to the physiological impact of genistein. To the best of my knowledge, activation of estrogen receptors is the only effect of genistein that has been documented in the low nanomolar range.

There is a recent report that genistein, as well as daidzein and biochanin A, have agonist activity for the so-called estrogen-related receptors (ERRs).³¹ These receptors are structural relatives of genuine estrogen receptors, and can activate transcription from estrogen response elements, but do not bind estrogen, and possess constitutive activity. Although isoflavones can bind to ERR and modestly enhance their transactivational activity, this effect is minimal and statistically insignificant at a genistein concentration of 1 μ M – nor are daidzein or biochanin A much more active in this regard. Thus, interaction of soy isoflavones with ERRs is unlikely to be of physiological significance.

Soy Isoflavones Have No Hepatic Effects

Studies in rats, primates, and humans demonstrate that hepatocytes express ERalpha, but not ERbeta.³²⁻³⁶ Many of the notable physiological effects of oral estrogen are mediated in the liver. Thus, oral estrogen lowers LDL cholesterol, raises serum triglycerides, boosts synthesis of angiotensinogen and sex hormone-binding globulin, and decreases hepatic production of IGF-I – effects which are thought to reflect direct estrogenic activity in hepatocytes.³⁷⁻⁴⁴ Moreover, oral estrogen up-regulates thrombotic mechanisms by modulating hepatic production of a range of plasma proteins which regulate thrombosis. Thus, oral estrogen increases plasma concentrations of clotting factors VII and IX, activated protein C, and C-reactive protein, while decreasing those of antithrombin, proteins C and S, and tissue factor pathway inhibitor.⁴⁵⁻⁴⁹ These effects are much less substantial when estrogens are administered transdermally; this is thought to explain the observation that risk for venous thromboembolism is increased far more by oral estrogen than by transdermal estrogen.^{47;50} Since physiological concentrations of genistein can be expected to have only a modest impact on ERalpha activity, it is not surprising that none of these effects are observed when soy isoflavones are ingested.⁵¹⁻⁵⁶

Gallstone risk is higher for women than men, and this has been traced to the fact that activated ERalpha boosts cholesterol output to the bile. In mice, ERalpha-selective agonists, but not ERbeta-selective agonists, have this effect.⁵⁷ It can be deduced that soy isoflavones will not increase risk for gallstones.

The modest impact of soy protein-rich diets on elevated LDL cholesterol in some studies presumably reflects replacement of “high-quality” animal protein (such as casein) with “lower-quality” plant protein.⁵⁸ Indeed, Sirtori, who first established the utility of soy protein-based diets for cholesterol reduction, pointed out that the soy protein isolate he used to first demonstrate this effect was devoid of isoflavones!⁵⁹ He maintained that the utility of his regimen was contingent on replacing animal protein with plant protein.⁶⁰ The studies which document reduction of elevated LDL cholesterol with supplemental soy protein have typically used comparable intakes of milk protein for the placebo

group;⁶¹ there is little evidence that simply adding soy protein to a diet that remains high in animal protein will lower elevated LDL.

While it may be disappointing to concede that soy isoflavones cannot lower LDL cholesterol, or diminish cancer risk by suppressing plasma IGF-I, it is nonetheless comforting to realize that isoflavones will not increase thrombotic risk in the way that oral estrogens do. Conceivably, the prothrombotic hepatic effects of oral estrogen are largely if not wholly responsible for the unanticipated increase in risk for myocardial infarction observed during recent prospective trials of oral hormone replacement therapy – despite the favorable influence of estrogen activity on vascular endothelium and LDL cholesterol.⁶²

No Uterotrophic Activity

Physiological concentrations of soy isoflavones can also be expected to be safe for the uterine endometrium, as the uterotrophic effect of estrogens appears to be mediated solely by ERalpha. This has been demonstrated elegantly in ERalpha-knockout mice, in which estrogens fail to exert a uterotrophic effect.⁶³ As would be expected, soy isoflavones have shown no impact on endometrial proliferation in clinical studies.⁶⁴⁻⁶⁸ Although endometrium expresses both alpha and beta receptors, synthetic agonists specific for ERbeta do not decrease uterine weight in rats or prevent the proliferative response to a concurrently administered ERalpha-specific agonist.³⁶ In women, soy isoflavones do not suppress the endometrial proliferative response to estrogen.⁶⁹ Based on these observations, ingestion of genistein within the nutritional range would not be expected to either increase or decrease endometrial cancer risk.

Nonetheless, when rats are fed doses of genistein that could be considered pharmacological – for example, 750 mg/kg diet, resulting in a free serum genistein level of 400 nM – uterotrophic activity is indeed seen.⁷⁰ This finding is consistent with the responsiveness of ERalpha to high nanomolar concentrations of genistein. The implication is that genistein doses which greatly exceed the nutritional range should not be presumed to be safe from the standpoint of endometrial cancer risk.

Genistein Up-Regulates eNOS Activity in Vascular Endothelium

One of the chief reasons for suspecting that hormone replacement therapy would decrease cardiovascular risk is that estrogens have a favorable impact on endothelial function, promoting the activity of the endothelial isoform of nitric oxide synthase (eNOS); this results both from increased transcription, and also from extranuclear effects of activated estrogen receptors (both ERalpha and ERbeta) exerted at the plasma membrane.⁷¹⁻⁷⁶ Estrogen can also enhance the bioactivity of nitric oxide by down-regulating NADPH oxidase expression – more specifically, that of its gp91phox subunit - in human endothelial cells.⁷⁷ Vascular endothelium expresses both ERalpha and ERbeta, and their effects on human umbilical vein endothelial cells appear to be quite comparable.⁷⁸ Studies point to a role for ERalpha in the induction of eNOS and/or NO production in the endothelial cells of diverse species;⁷⁹⁻⁸⁴ the impact of ERbeta in this regard is less clear.

Whereas estrogen increased the expression of eNOS in the coronary arteries of ovariectomized ERalpha knockout mice,⁸⁵ consistent with a role for ERbeta in eNOS induction, no such effect was seen in their cerebrovascular arteries.⁸¹ Induction of eNOS by ERbeta in rat cardiac myocytes and in human myometrium has been reported.^{86;87} In the vascular smooth muscle cells of rodents, ERbeta exerts both antihyperplastic and antihypertensive effects.^{88;89}

Clinical observations appear consistent with the possibility that ERbeta supports endothelial eNOS activity in at least some vascular beds. Adequate oral intakes of genistein have been reported to improve endothelium-dependent vasodilation – as well as other markers for endothelial NO production - in postmenopausal women.⁹⁰⁻⁹³ The most striking findings in this regard have been reported by Squadrito and colleagues, who administered either 54 mg free genistein daily, a typical oral hormone replacement regimen, or a placebo, to 90 postmenopausal women for 1 year.⁹¹ Genistein treatment increased brachial endothelium-dependent vasodilation and post-occlusive blood flow relative to placebo; the effects of the hormone replacement regimen were quite similar. Moreover, plasma levels of nitric oxide metabolites were doubled, and plasma levels of endothelin-1 virtually halved, in both the genistein and hormone replacement groups. (This suppression of endothelin-1 may reflect the ability of NO to inhibit endothelial secretion of endothelin.)⁹⁴ Other studies have likewise reported favorable effects of soy isoflavone supplementation (or administration of isoflavone-rich soy protein) on vascular endothelial function in women – albeit a few studies have failed to observe such an effect.^{95;96} One study reported an improvement in arterial compliance, but not in acetylcholine-mediated vasodilation.⁹⁷ In ovariectomized rats, dietary genistein has been shown to improve endothelium-dependent, but not endothelium-independent, vasodilation.⁹⁸⁻¹⁰⁰

Squadrito suggests that lower intakes of genistein, or shorter duration of supplementation, might account for the two negative reports. Indeed, each of these studies administered 80 mg of mixed conjugated soy isoflavones daily,^{95;96} it would take about 150 mg of such a preparation to provide the 54 mg of pure genistein used in the Squadrito study. The fact that Squadrito administered free genistein, rather than the genistein glycoside (genistin) that occurs natively in unfermented soy foods, might also have some bearing in this regard. There are conflicting reports regarding the relative bioavailabilities of free isoflavones and conjugated isoflavones; one Japanese study concluded that, during longterm administration, administration of free isoflavones yielded plasma concentrations of total isoflavones that were roughly twice as high as those achieved during administration of conjugated isoflavones.¹⁰¹ On the other hand, three recent American and Swiss studies conclude that, after single oral doses, the availabilities of the two forms of isoflavones are roughly comparable.¹⁰²⁻¹⁰⁴ Although isoflavone glycosides are not taken up by intestinal cells,¹⁰⁵ these compounds are readily converted to free glycosides by membrane-bound or bacterial β -glucosidases in the intestinal tract;¹⁰⁶ the resulting free isoflavones are absorbable.¹⁰⁷ The propensity of gut bacteria to degrade isoflavones varies from person to person and influences their bioavailability – slow metabolizers achieve higher plasma levels of genistein and daidzein.¹⁰⁸ Since free isoflavones should be absorbed more rapidly, one would suspect that they would be more effective than

conjugates in rapid metabolizers – but this has not been documented. Evidently, more research is needed to evaluate the relative impact of free vs. conjugated isoflavones on plasma isoflavone levels during long-term administration.

The favorable effects of genistein on endothelial function might be operative in men as well, in light of a report that intrabrachial administration of genistein led to a nitric oxide-mediated increase in forearm blood flow in male volunteers; infusion of 17beta-estradiol – but not daidzein – had a comparable effect.¹⁰⁹ On the other hand, one study reported a modest reduction in endothelium-dependent vasodilation after men had ingested isoflavone-rich soy protein (40 g daily, providing 118 mg isoflavones) for three months.¹¹⁰ The impact of dietary genistein on endothelial function in males should be studied further. Estrogen can boost endothelium-dependent vasodilation in males,¹¹¹⁻¹¹³ and studies in aromatase knockout male mice indicate that endogenous estrogen improves endothelial function in the males of this species.¹¹⁴

In light of the versatile role which endothelial nitric oxide production plays in preservation of vascular health, it seems likely that genistein's ability to up-regulate eNOS function could be exploited to decrease vascular risk. Epidemiological evidence suggests that estrogen may be largely responsible for the relative protection from cardiovascular disease enjoyed by premenopausal women.¹¹⁵⁻¹¹⁷ It would be a fortunate development indeed if men could use genistein to achieve at least a portion of this benefit without incurring typical estrogenic side effects.

Sadly, induction of eNOS is not always an unalloyed benefit; in endotheliopathies associated with increased superoxide production, tetrahydrobiopterin deficiency leads to an “uncoupling” of eNOS that impairs its activity while turning it into a superoxide generator.¹¹⁸⁻¹²⁰ Fortunately, there is recent evidence that high-dose folic acid can restore the proper protective function of eNOS when tetrahydrobiopterin is deficient.¹²¹⁻¹²⁵ Thus, it is reasonable to suggest that concurrent administration of high-dose folate could be a prudent adjuvant to genistein supplementation in patients at risk for endothelial dysfunction. Supplemental arginine, as well as various measures which lessen superoxide production by NADPH oxidase, may also help to optimize the bioefficacy of the eNOS expressed by dysfunctional endothelium.¹²⁵⁻¹²⁷

Favorable Effects on Bone Metabolism

Both ERalpha and ERbeta are expressed in the main cell types present in human bone: osteoblasts, osteoclasts, and osteocytes.¹²⁸⁻¹³⁰ Studies with osteoblast-derived cell lines indicate that these two receptors modulate the transcription of distinctly different sets of genes, with only a modest amount of overlap.¹³¹ The effects of ERalpha on gene expression tend to be amplified in ERbeta knockout mice, suggesting that ERbeta down-regulates some responses to ERalpha.¹³² Cortical bone density is greater, and age-related loss of trabecular density is lower, in ERbeta knockout mice as compared to wild type.¹³³ Yet in ovariectomized ERalpha knockout mice, estrogen promotes increased bone density – albeit not as effectively as it does in ERalpha knockouts; evidently, both types of estrogen receptor can act to preserve bone integrity.¹³⁴ Indeed, trabecular bone

density tends to be increased in ERalpha knockout female mice.¹³⁵ These effects may be sex-specific, as estrogen did not improve bone density in orchidectomized ERalpha knockout mice.¹³⁴ In a human osteoblast-derived cell line (MG63) in which ERalpha had been silenced with antisense plasmids, estrogen increased collagen and alkaline phosphatase secretion, demonstrating an anabolic effect of ERbeta in these cells.¹³⁶

The increase in bone osteoclastic activity and bone resorption ushered in by menopause is thought to stem primarily from an alteration of osteoblast function; soluble mediators produced by osteoblasts have a major impact on the functional status of nearby osteoclasts. Estrogen inhibits osteoblast production of IL-6, an important trophic factor for osteoclasts; it also boosts production of osteoprotegerin, a soluble “false receptor” which inhibits activity of RANKL, another important trophic factor for osteoclasts.^{14;137;138} It is thus of particular interest that, in physiological concentrations, genistein has been shown to suppress IL-6 production, and boost osteoprotegerin production, in human osteoblast-derived cell lines.^{139;140} These effects are inhibited by concurrent incubation with an estrogen receptor antagonist. These findings strongly suggest that genistein can interact with ERbeta to achieve transrepression of the IL-6 promoter – as it does with the TNF promoter.⁷

Genistein also has the potential to improve bone metabolism through its impact on vascular eNOS, in light of recent evidence that this enzyme is a mediator of the osteogenic impact of estrogen on osteoblasts.¹⁴¹⁻¹⁴³

In light of these considerations, it is not surprising that, in non-uterotrophic doses, genistein has repeatedly been shown to promote maintenance of bone density in ovariectomized rats and mice.¹⁴⁴⁻¹⁴⁹ Whether this effect would be lost in ERbeta knockout mice – thus confirming that ERbeta mediates the effect – has not been determined. Surprisingly, two drugs said to be potent and selective ERbeta agonists did not inhibit bone loss in ovariectomized rats.^{36;150} However, the fact that these agents have agonist activity for transcriptional promotion in some contexts, does not necessarily imply that they will have such activity in other contexts – nor that they will interact with ERbeta in a manner that achieves transrepression of the IL-6 promoter.

Clinical studies of supplementation with soy isoflavones or isoflavone-rich soy protein in postmenopausal women have reached divergent conclusions: some find that such supplementation has a favorable effect on markers of bone metabolism and on preservation of bone density,^{26;151-156} whereas others do not.¹⁵⁷⁻¹⁵⁹ Perhaps the most impressive study in this regard was that of Squadrito and colleagues.²⁶ These researchers recruited 90 postmenopausal women, 47-57 years of age, who were randomized to receive either genistein (54 mg daily), a standard HRT combination (17β-estradiol/norethisterone acetate), or a matching placebo; response was evaluated at 6 and 12 months. Genistein treatment was found to decrease excretion of pyridinium cross-links (a marker for collagen catabolism in bone) at both 6 and 12 months; this response was quite comparable to that achieved with HRT. In contrast to HRT, however, genistein increased serum levels of bone alkaline phosphatase and osteocalcin, markers for osteoblastic activity. At 12 months, the serum RANKL/osteoprotegerin ratio was notably

reduced in the genistein group, to a greater extent than in the HRT group.¹⁵⁶ Finally, after 12 months, bone mineral density in the femoral neck and lumbar spine had increased by 3-4% in both the genistein and HRT group, as contrasted to modest losses of density noted in the placebo group. This study is notable for its comparatively long duration – sufficient to evaluate changes in bone density – and for its use of a fairly ample dose of free genistein. The total serum genistein (conjugated plus free) in the genistein-supplemented group, measured during a morning fast, was 1.5 μM at 6 months and 1.7 μM at 12 months; assuming that about 1.3% of the genistein pool was unconjugated, this would be expected to correspond to a free genistein concentration of about 20 nM, presumably sufficient to activate ERbeta.

Most other studies of this type have administered isoflavones as glycosides, often in conjunction with soy protein. It is conceivable that variations in the bioavailability of genistin are at least partially responsible for the inconsistent findings of these studies. In future, such studies should measure serum free genistein levels to assess the bioavailability of the administered isoflavones. Isoflavone supplementation has also been assessed in premenopausal women and in men; no impact on bone metabolism or bone density was noted in these studies.^{160;161}

Since East Asian women often consume ample amounts of soy isoflavones in their habitual diets, several studies have attempted to correlate habitual isoflavone intake or serum isoflavone level with postmenopausal bone density and/or bone metabolism in such women. Two studies – one each from Japan and China - have reported that women in the highest quartile of soy intake, as contrasted to the lowest quartile, had higher bone density.^{162;163} Several other studies, including those in the U.S. or Europe, where soy intake is comparatively low, did not observe such a correlation,¹⁶⁴⁻¹⁶⁶ albeit a Southern California study had a positive outcome.¹⁶⁷

Reducing Prostate Cancer Risk

Human prostatic epithelium expresses ERbeta, but not ERalpha – whereas prostate stroma expresses ERalpha.^{168;169} There is reason to believe that ERbeta activity has an antiproliferative impact both in healthy prostate and in prostate cancers.^{170;171} Prostatic hypertrophy is common in aging ERbeta-knockout mice, whereas knockout of ERalpha has no such effect.^{172;173} Furthermore, transfection of ERbeta into human prostate cancer cell lines induces apoptosis.¹⁷⁴ As prostate cancers progress, ERbeta expression tends to decrease – consistent with the possibility that this receptor exerts a restraining effect on proliferation.^{169;175;176} In prostate cancer cell lines which express ERbeta, a variety of estrogens and anti-estrogens – including genistein and the drug raloxifene – have an antiproliferative, pro-apoptotic effect.¹⁷⁷⁻¹⁸⁰

Genistein has been shown to decrease expression of the androgen receptor in the human prostate cancer-derived LNCaP cell line, an effect mediated by activation of ERbeta.¹⁸¹ Moreover, soy phytochemical concentrates slow the growth of LNCaP in nude mice.^{180;182;183} Genistein feeding likewise down-regulates androgen receptor expression in rat prostate,¹⁸⁴ and reduces the yield of prostate cancer in carcinogen-treated rats as

well as in transgenic “TRAMP” mice that have a high spontaneous incidence of this cancer.^{185;186} Pilot clinical studies evaluating the impact of oral genistein on early stage prostate cancer have achieved a moderate reduction of PSA in a minority of patients, and an apparent reduction in cancer growth rate in others.^{187;188} Case-control studies from the Orient are reasonably consistent with the thesis that diets high in soy products are associated with lower risk for prostate cancer¹⁸⁹⁻¹⁹¹ – albeit high soy intake may be a marker for diets and lifestyles that are more traditional.

Isoflavones and the Breast – Safe and Possibly Protective

How dietary soy isoflavones influence breast cancer risk is also a matter of considerable interest. In the normal human breast, both types of estrogen receptor are expressed in epithelial cells;¹⁹² ERbeta predominates in adult human mammary fibroblasts.¹⁹³ In ERalpha knockout mice, the breast is atrophic; conversely, in ERbeta knockout mice, epithelium is hyperproliferative and the mice are prone to severe cystic breast disease as they age.^{194;195} Transfection of ERbeta into an ERalpha-expressing human breast cancer cell line (MCF-7) results in a suppression of proliferation associated with up-regulation of cdk inhibitors p21 and p27, and down-regulation of c-myc and cyclins D1 and A; however, these effects are only partially ligand dependent.¹⁹⁶ ERbeta transfection also slows the estrogen-stimulated growth of the estrogen-sensitive T47D mammary cancer cell line,¹⁹⁷ but has a pro-proliferative impact on the MDA-MB-435 tumor.¹⁹⁸ In the main, these findings suggest that ERalpha and ERbeta may have a “yin-yang” role in breast development, with ERbeta opposing the proliferative impact of ERalpha; however, they do not necessarily imply that ERbeta-specific ligands will have an antiproliferative effect.

ERbeta is expressed by the majority of human breast cancers – even those considered “estrogen negative”;^{199;200} the standard assays for breast cancer “estrogen receptors” are ERalpha-specific. The prognostic significance of ERbeta expression in breast cancer has been the subject of conflicting reports.¹⁹⁶

A number of studies have examined the impact of soy isoflavones on breast cancer risk or growth in rodents, but some of these are of limited interest owing to their use of high parenteral doses that would likely have ERalpha-agonist activity. High-dose genistein, administered pre-pubertally, induces a premature differentiation of breast tissue that diminishes susceptibility of adult rats to carcinogen-induced breast cancer;²⁰¹⁻²⁰³ this effect is also seen with estrogen administration, and there is no evidence that physiological levels of genistein could achieve a comparable effect. There are however several studies which conclude that more modest intakes of genistein can favorably influence breast cancer induction. When administered at 250 mg/kg diet, either genistein or daidzein was found to slow the onset of breast cancer in cancer-prone MMTV-neu mice; however, they did not influence the growth of established tumors.²⁰⁴ A functional food rich in unconjugated genistein slowed the growth of the MDA-MB-231 human breast cancer in nude mice; this was associated with increased apoptosis in the tumor.²⁰⁵ In the mouse mammary tumor virus-induced spontaneous breast cancer model, oral administration of biochanin A – but not daidzein – reduced tumor incidence at 15

months; this effect was not seen in germ-free animals, and thus presumably was contingent on conversion of biochanin A to genistein by intestinal bacteria.²⁰⁶

On the other hand, a number of studies show that dietary genistein can increase the growth of estrogen-dependent MCF-7 tumors in ovariectomized nude mice; this effect appears to hinge on activation of ERalpha.²⁰⁷⁻²¹⁰ This cell line seems to be exquisitely sensitive to ERalpha activation, inasmuch as genistein concentrations as low as 10 nM can modestly increase its rate of proliferation in vitro.^{207,211} Nonetheless, a much more substantial response is seen with genistein concentrations in the high nanomolar range, consistent with the known affinity of genistein for ERalpha; responsiveness at 10 nM presumably reflects that fact that activation of only a small minority of ERalpha receptors can have a discernible impact on proliferation in this cell line. Administered at 750 mg/kg of diet to rats pretreated with the carcinogen MNU, genistein increases the size of MNU-induced mammary tumors in ovariectomized rats;²¹² this dose of genistein also increases uterus size, pointing to an ERalpha-mediated effect on estrogen-sensitive tumors. In DMBA-treated mice, 1 g/kg dietary genistein increases the yield of malignant adenocarcinomas – whereas no cancers develop with ERalpha knockout mice.²¹³ Overall, these findings suggest that moderate intakes of genistein may slow the onset or progression of certain mammary tumors, but that very high intakes can be expected to boost the growth of estrogen-dependent tumors, and even moderate intakes may have the potential to at least modestly influence the growth of certain tumors that are highly sensitive to ERalpha activation.

The impact of soyfood ingestion on breast cancer incidence has been examined in a number of case-control as well as prospective epidemiological studies. Although many of these studies find no link between breast cancer and soy consumption²¹⁴⁻²¹⁸, four case-control studies found an inverse association between soy intake and risk for premenopausal breast cancer in Asian populations, and two such studies found a similar association with postmenopausal breast cancer.²¹⁹⁻²²³ One recent prospective Japanese study found that high intakes of miso or of isoflavones – but not of soyfoods per se – were associated with decreased risk for pre- or postmenopausal breast cancer.²²⁴ Other prospective studies have had a negative outcome. One case-control study in Asian-Americans focused on soy consumption during adolescence, and found that high soy intake during this time predicted a lower risk for postmenopausal breast cancer; such risk was lowest for those who maintained high soy consumption in adult life.²²³ Given the fact that most Asian diets provide suboptimal isoflavone intakes from the standpoint of ERbeta activation, these findings are reasonably consistent with the possibility that somewhat higher supplemental intakes of genistein might be protective in regard to breast cancer risk; however, they certainly don't prove this proposition. A conservative but optimistic perspective is that, as contrasted with HRT, there is little reason to suspect that genistein intakes in the high *nutritional* range would *increase* breast cancer risk – and some reason to suspect that such a measure might decrease this risk.

On the other hand, in women who have been diagnosed with estrogen-sensitive breast cancers, the possibility cannot be excluded that nutritional intakes of genistein will modestly boost cancer growth by promoting low-level activation of ERalpha.

Theoretically, selective activation of ERbeta with moderate-dose genistein might slow the growth of certain estrogen-sensitive mammary cancers which express this receptor - but this is speculative. Until further evidence is forthcoming, it might be prudent for women with estrogen-sensitive breast cancer to refrain from frequent soy ingestion or isoflavone supplementation.

Modestly Effective for Hot Flashes

A number of clinical studies have assessed the impact of supplemental soy isoflavones – with or without soy protein – on postmenopausal hot flashes. A recent overview notes that 4 of these studies had a positive outcome, 5 were negative, and one showed a positive trend that missed statistical significance.²²⁵ This overview could not include a more recent study by Squadrito and colleagues.²²⁶ These researchers nested a hot flash study into their bone metabolism study by enrolling only women who were troubled by this complication; since this study achieved free genistein concentrations sufficient to benefit bone metabolism, and since it included an HRT arm, its findings may be particularly illuminating. At baseline, the daily hot flash score was very similar in the three groups, averaging 4.5-4.7 per day. After 3 months, as contrasted with the placebo group, this score was 22% lower in the genistein group and 53% lower in the HRT group; these differences were statistically significant. The findings at 12 months were similar – relative to placebo response, the score was 24% lower with genistein and 54% lower with HRT. Response to HRT was significantly greater than that to genistein.

The available findings appear consistent with the proposition that soy isoflavone regimens which achieve an adequate plasma level of free genistein are mildly beneficial with respect to hot flashes – though less effective in this regard than HRT. The inconsistency of the results of clinical studies examining this issue, may reflect the fact that the benefit to be expected is modest, as well as the likelihood that some of the isoflavone regimens tested failed to achieve adequate free genistein concentrations in some subjects. It seems highly unlikely that about half of the clinical studies to date examining this issue would find statistically significant benefit, if in fact isoflavones had no genuine potential for controlling hot flashes. Soy isoflavones have access to the brain, and certain regions of the hypothalamus express ERbeta;²²⁷⁻²²⁹ conceivably, some of these receptors mediate the impact of genistein on hot flashes.

Potential for Prevention of Glomerulosclerosis and Left Ventricular Hypertrophy

Chronic renal disease, of either diabetic or non-diabetic origin, tends to progress less rapidly in women than in men.²³⁰⁻²³² The relative protection enjoyed by women appears to be confined to the premenopausal period, and thus is likely to be mediated by estrogen.^{232;233} Indeed, estrogen ameliorates, whereas ovariectomy exacerbates, the progression of glomerulosclerosis in various rodent models of this disorder.²³⁴⁻²⁴⁰ Contrary findings have been reported in certain hyperlipidemic strains of rodents, presumably because estrogen treatment exacerbates nephrotoxic hyperlipidemia in these animals;²⁴¹⁻²⁴⁴ an estrogen-evoked increase in growth hormone secretion can also exert a

countervailing negative effect in this regard.²⁴⁵⁻²⁴⁷ However, these latter findings do not appear to be germane to the impact of endogenously-produced estrogen in women.

The various agents and circumstances which provoke glomerulosclerosis – such as hyperglycemia, glomerular hypertension, angiotensin II, advanced glycation endproducts, thromboxane, and oxidized LDL – appear to do so by boosting glomerular production of transforming growth factor-beta (TGF- β); activation of AP-1 response elements in the TGF- β promoter, often in response to PKC/MAP kinase activation, plays a role in these inductions.²⁴⁸⁻²⁶¹ This increased autocrine/paracrine production of TGF- β , in turn, activates mesangial receptors for this hormone, leading to increased production of various ground substance proteins – including collagen types I and IV, laminin, and fibronectin – decreased production of the collagenases MMP-2 and MMP-9, and increased production of protease inhibitors such as TIMP-1;²⁶²⁻²⁷² the net effect is an accumulation of mesangial ground substance and a thickening of glomerular basement membranes. TGF-beta is also a mediator of the proteinuria characteristically seen in glomerular disorders.²⁷³⁻²⁷⁷ Injection of anti-TGF- β antibodies into diabetic rodents can prevent and in some measure reverse glomerulosclerosis – demonstrating the central role of TGF- β in this syndrome.²⁷⁷⁻²⁸⁰

Mesangial cells express both ERalpha and ERbeta;^{232,281} it is noteworthy that expression of both types of ER is diminished in this mesangial cells of a strain of mouse prone to glomerulosclerosis.²³² Physiological concentrations of estrogen have been shown to suppress the response of mesangial cells to TGF- β .^{282,283} This effect appears to reflect the ability of activated estrogen receptors of both types to bind to, and suppress the transactivating activity of, the transcription factor SMAD3,²⁸⁴ whose phosphorylation and activation by TGF- β receptors mediates most effects of TGF- β .²⁸⁵ (Not surprisingly, SMAD3-knockout mice are virtually immune to diabetic glomerulopathy.)²⁷⁶ Moreover, estrogen can also suppress glomerular production of TGF- β , by boosting the NO production of glomerular endothelial cells.²⁸⁶ NO's suppressive impact on TGF- β production is poorly understood; the fact that eNOS inhibitors up-regulate glomerular TGF- β synthesis suggests that this mechanism is of physiological significance. It does not appear to be known whether glomerular endothelial cells express ERbeta, or whether this receptor can enhance eNOS activity in these cells. A further theoretical possibility is that activated ERbeta could inhibit transcription of TGF- β by interfering with AP-1 activity in its promoter; on the other hand, ERalpha would be expected to enhance AP-1 activity.^{15;20} In any case, there is reason to suspect that high-physiological concentrations of genistein, via activation of ERbeta, could inhibit the effects of TGF- β on mesangial cells, thereby helping to prevent glomerulosclerosis. Whether activation of ERbeta could also increase glomerular NO production and/or interfere with AP-1 activity – resulting in suppression of glomerular TGF- β production – is more speculative.

In fact, many rodent studies conclude that soy-based diets – in comparison to casein-based diets – are associated with slower progression of glomerulosclerosis.²⁸⁷⁻²⁹⁵ Two recent clinical studies in type 2 diabetics likewise report that proteinuria is lower during soy protein supplementation than during casein supplementation.^{296;297} At least a portion of this effect reflects the fact that soy is of “poorer quality” than casein. It is well

established that diets rich in “high quality” protein promote glomerulosclerosis by increasing glomerular pressure and thereby increasing glomerular filtration rate; this presumably represents a homeostatic response which helps the body to cope with increased nitrogenous waste.^{291;298;299} Soy and other plant proteins have a less substantial effect in this regard.³⁰⁰⁻³⁰² Thus, diets low but adequate in protein content have been used clinically to slow the progression of glomerular disease;^{303;304} in particular, quasi-vegan diets, some featuring soy protein, have been used for this purpose.^{301;305-308}

The possibility that phytochemical components of soy – such as isoflavones – contribute to the nephroprotection afforded by soy-based diets has been suggested by Valasquez and Bhatena.^{291;309} Indeed, a subsequent study with hypercholesterolemic rats prone to glomerulosclerosis demonstrated that addition of an isoflavone-rich soy ethanol extract to their casein-based diets ameliorated subsequent renal damage; the interpretation of this study is complicated by the fact that serum lipids declined in the isoflavone-supplemented rats.³¹⁰ Also of particular interest in this regard is a study by Neugarten and colleagues.³¹¹ These researchers demonstrated that, in concentrations as low as 1-10 nM, genistein markedly inhibits synthesis of both type I and type IV collagen by murine mesangial cells; the authors propose that ERbeta mediates this effect. Conceivably, this finding simply reflects the fact that genistein-activated ERbeta can block the impact of autocrine TGF- β on mesangial cells.²⁸⁴ Whether physiological concentrations of genistein can also influence glomerular production of TGF- β remains to be seen. Of related interest is a report that a diet supplemented with red clover isoflavones decreases production of TGF- β by prostatic epithelium in mice.³¹²

If subsequent animal and clinical studies prove that soy isoflavones can indeed reduce risk for, or slow progression of, glomerulosclerosis, it won't necessarily follow that a diet high in soy protein should be recommended. Most likely, the most nephroprotective diet will be a vegan diet relatively low in protein content, supplemented with soy isoflavones. In this regard, adding soy protein to a diet already rich in casein did not protect hypercholesterolemic rats from glomerular injury – albeit addition of a comparable surplus of casein exacerbated this injury.²⁹³

The intracellular signaling mechanisms which mediate left ventricular hypertrophy (LVH) appear to be very similar to those that evoke glomerulosclerosis;^{313;314} moreover, NO has an antagonistic impact on development of LVH analogous to its impact on glomerulosclerosis.³¹⁵ Premenopausal women are relatively protected from LVH, and estrogen replacement has been shown to limit expansion of ventricular mass in postmenopausal women and ovariectomized rodents at risk for this disorder.³¹⁶⁻³²³ Cardiac myocytes and fibroblasts express both isoforms of the estrogen receptor, but, in neonatal rat cardiac myocytes, only ERbeta has an inductive effect on eNOS.^{324;325} A very recent study examining ERalpha- and ERbeta-knockout mice demonstrates that ERbeta mediates the protective effect of estrogen on cardiac hypertrophy.³²⁶ Thus, it is reasonable to suspect that genistein has the potential to protect postmenopausal women, and possibly men, from LVH.

Directions for Future Research

The thesis that ERbeta mediates the favorable physiological effects of moderate-dose isoflavones on bone metabolism and endothelial function can best be tested in ovariectomized ERbeta knockout rodents, using dietary concentrations of isoflavones that will achieve a free genistein plasma concentration not in excess of 50 nM. If these effects largely persist in the ERbeta knockouts, the thesis of this paper will be falsified, and it will be necessary to identify further molecular targets that respond to free isoflavones in the low nanomolar range. In light of current evidence, however, the contention that ERbeta is the key target of dietary isoflavones is credible and brings a satisfying unity to the diverse research literature on these compounds.

The fact that daidzein has a relatively low affinity for both ERbeta and ERalpha, may seem difficult to square with a handful of reports indicating that daidzein, or its methoxylated derivative formononetin, can be physiologically active.³²⁷⁻³³⁰ GI bacterial action has the potential to convert these compounds to equol, the S-isomer of which has recently been shown to be a potent and selective agonist for ERbeta, with an affinity almost as high as that of genistein.^{331,332} The R-isomer, on the other hand, is relatively selective for ERalpha, and has a higher affinity for this receptor than does genistein ($K_i = 50$ nM). Too little is currently known about the extent of glucuronidation of plasma equol, or the relative abundance of the two isoforms, to make firm predictions regarding the possible contribution of equol to the physiological effects of soy isoflavones. Further complicating this issue is the fact that capacity to convert daidzein to equol varies a great deal from person to person, to the extent that people have been categorized as equol “producers” or “non-producers”.^{331,333} Since soy isoflavone supplementation, at least in nutritional doses, is non-uterotrophic, it is reasonable to conclude that R-equol makes little contribution to the physiological effects of such supplementation. Whether S-equol has a more significant physiological role – thus rationalizing claims that daidzein or formononetin are clinically active – must be clarified in future research.

The fact that clinical studies with soy isoflavone supplementation have yielded inconsistent results may reflect, at least in part, variations in the plasma levels of free genistein (and possibly equol) achieved by the diverse supplementation regimens that have been assessed. Is genistin truly as effective as equimolar intakes of genistein during longterm administration in most subjects? Or do variations in GI metabolism of genistin render supplementation with genistein a more fail-safe proposition? Too few studies have assessed the long-term impact of various isoflavone regimens on equilibrium concentrations of free genistein; we should bear in mind that acute pharmacokinetic studies do not take into account possible adaptive changes in enzyme expression that could influence achieved plasma levels. In light of the markedly beneficial effects on endothelial function and bone metabolism achieved by Squadrito et al. with 54 mg genistein daily, efforts to confirm these results in larger and more diverse populations are clearly warranted.

As a safer substitute for HRT in postmenopausal women, supplemental genistein would appear to have great promise; in Squadrito’s studies, the impact of supplemental genistein

on endothelium and bone was fully comparable to that of HRT, and the relief from hot flashes noted was worthwhile though less substantial. Whether premenopausal women would benefit is less clear, since the impact of genistein on endothelium or bone may be modest compared to that of ambient estrogen levels. However, the epidemiological and rodent literature provides just a hint that, by shifting the balance toward ERbeta activity in mammary tissue, premenopausal soy isoflavone ingestion may in fact be protective in regard to breast cancer risk. In this regard, more rodent chemoprevention studies, using moderate genistein doses that will selectively activate ERbeta, would be desirable; in regard to epidemiology, more attention should be focused on the possible impact of adolescent isoflavone intake on subsequent breast cancer risk.

Could supplemental genistein improve endothelial function and bone metabolism in men? Too few studies have examined this issue to allow any conclusions to be drawn. However, even if genistein cannot protect men in these respects, its likely impact on prostate health may make genistein supplementation a very worthwhile option for men. Ongoing clinical studies with genistein in early prostate cancer may shed further light on this issue. The fact that ERbeta expression tends to be lost as prostate cancer progresses probably means that, as a therapy for pre-existing prostate cancer, genistein will have at best transient efficacy; thus, its greater potential may be for chemoprevention.

With respect to expectations that isoflavone-rich diets may decrease risk for certain common cancers, a proviso is in order. Commentators frequently note that risks for certain “Western” cancers are comparatively low in East Asian cultures which make frequent use of soy products. However, these rates are equally low in many other Third World rural societies in Africa and South America where soy consumption is minimal.³³⁴⁻³³⁶ The traditional diets of these societies tend to be low in fat and animal products, and moderate in total protein; lifelong consumption of such diets is likely to be associated with reduced serum levels of insulin and of free IGF-I – now known to have important cancer promotional activity.³³⁷⁻³³⁹ High intakes of soy protein can actually boost serum IGF-I,³⁴⁰⁻³⁴² thus, heavy use of soy protein per se may be inadvisable from the standpoint of cancer risk. These considerations suggest that ample intakes of soy isoflavones may best be achieved through supplementation rather than through heavy consumption of protein-rich soy products. Furthermore, we should take care not to encourage the delusion that simply adding soy products – or soy isoflavones – to a meat-rich omnivore diet will reproduce the full measure of cancer protection associated with quasi-vegan Third World diets. The popular focus on soy protein runs the risk of obscuring the broader and deeper truth that diets featuring “low quality” plant protein can have an important anti-promotional impact on many types of cancer.^{339;343;344}

References

1. Kuiper GG, Enmark E, Pelto-Huikko M, Nilsson S, Gustafsson JA. Cloning of a novel receptor expressed in rat prostate and ovary. *Proc.Natl.Acad.Sci U.S.A* 1996;93:5925-30.
2. Kuiper GG, Gustafsson JA. The novel estrogen receptor-beta subtype: potential role in the cell- and promoter-specific actions of estrogens and anti-estrogens. *FEBS Lett.* 1997;410:87-90.
3. Kuiper GG, Carlsson B, Grandien K, Enmark E, Haggblad J, Nilsson S *et al.* Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptors alpha and beta. *Endocrinology* 1997;138:863-70.
4. Kuiper GG, Lemmen JG, Carlsson B, Corton JC, Safe SH, van der Saag PT *et al.* Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology* 1998;139:4252-63.
5. Barkhem T, Carlsson B, Nilsson Y, Enmark E, Gustafsson J, Nilsson S. Differential response of estrogen receptor alpha and estrogen receptor beta to partial estrogen agonists/antagonists. *Mol.Pharmacol* 1998;54:105-12.
6. Routledge EJ, White R, Parker MG, Sumpter JP. Differential effects of xenoestrogens on coactivator recruitment by estrogen receptor (ER) alpha and ERbeta. *J Biol Chem* 2000;275:35986-93.
7. An J, Tzagarakis-Foster C, Scharschmidt TC, Lomri N, Leitman DC. Estrogen receptor beta-selective transcriptional activity and recruitment of coregulators by phytoestrogens. *J Biol Chem* 2001;276:17808-14.
8. Morito K, Hirose T, Kinjo J, Hirakawa T, Okawa M, Nohara T *et al.* Interaction of phytoestrogens with estrogen receptors alpha and beta. *Biol Pharm.Bull.* 2001;24:351-56.
9. Liu J, Knappenberger KS, Kack H, Andersson G, Nilsson E, Dartsch C *et al.* A homogeneous in vitro functional assay for estrogen receptors: coactivator recruitment. *Mol.Endocrinol.* 2003;17:346-55.
10. Muthyala RS, Ju YH, Sheng S, Williams LD, Doerge DR, Katzenellenbogen BS *et al.* Equol, a natural estrogenic metabolite from soy isoflavones: convenient preparation and resolution of R- and S-equols and their differing binding and biological activity through estrogen receptors alpha and beta. *Bioorg.Med Chem* 2004;12:1559-67.
11. Bovee TF, Helsdingen RJ, Rietjens IM, Keijer J, Hoogenboom RL. Rapid yeast estrogen bioassays stably expressing human estrogen receptors alpha and beta, and green fluorescent protein: a comparison of different compounds with both receptor types. *J Steroid Biochem Mol.Biol* 2004;91:99-109.

12. Mueller SO, Simon S, Chae K, Metzler M, Korach KS. Phytoestrogens and their human metabolites show distinct agonistic and antagonistic properties on estrogen receptor alpha (ERalpha) and ERbeta in human cells. *Toxicol.Sci* 2004;80:14-25.
13. Kostelac D, Rechkemmer G, Briviba K. Phytoestrogens modulate binding response of estrogen receptors alpha and beta to the estrogen response element. *J Agric.Food Chem* 2003;51:7632-35.
14. Kurebayashi S, Miyashita Y, Hirose T, Kasayama S, Akira S, Kishimoto T. Characterization of mechanisms of interleukin-6 gene repression by estrogen receptor. *J Steroid Biochem Mol.Biol* 1997;60:11-17.
15. Paech K, Webb P, Kuiper GG, Nilsson S, Gustafsson J, Kushner PJ *et al.* Differential ligand activation of estrogen receptors ERalpha and ERbeta at AP1 sites. *Science* 1997;277:1508-10.
16. Saville B, Wormke M, Wang F, Nguyen T, Enmark E, Kuiper G *et al.* Ligand-, cell-, and estrogen receptor subtype (alpha/beta)-dependent activation at GC-rich (Sp1) promoter elements. *J Biol Chem* 2000;275:5379-87.
17. Sabbah M, Courilleau D, Mester J, Redeuilh G. Estrogen induction of the cyclin D1 promoter: involvement of a cAMP response-like element. *Proc.Natl.Acad.Sci U.S.A* 1999;96:11217-22.
18. Kushner PJ, Agard DA, Greene GL, Scanlan TS, Shiau AK, Uht RM *et al.* Estrogen receptor pathways to AP-1. *J Steroid Biochem Mol.Biol* 2000;74:311-17.
19. Wu JJ, Geimonen E, Andersen J. Increased expression of estrogen receptor beta in human uterine smooth muscle at term. *Eur.J Endocrinol.* 2000;142:92-99.
20. Maruyama S, Fujimoto N, Asano K, Ito A. Suppression by estrogen receptor beta of AP-1 mediated transactivation through estrogen receptor alpha. *J Steroid Biochem Mol.Biol* 2001;78:177-84.
21. Liu MM, Albanese C, Anderson CM, Hilty K, Webb P, Uht RM *et al.* Opposing action of estrogen receptors alpha and beta on cyclin D1 gene expression. *J Biol Chem* 2002;277:24353-60.
22. Kanda N, Watanabe S. 17Beta-estradiol inhibits MCP-1 production in human keratinocytes. *J Invest Dermatol.* 2003;120:1058-66.
23. Adlercreutz H, Markkanen H, Watanabe S. Plasma concentrations of phyto-oestrogens in Japanese men. *Lancet* 1993;342:1209-10.
24. Adlercreutz H, Fotsis T, Lampe J, Wahala K, Makela T, Brunow G *et al.* Quantitative determination of lignans and isoflavonoids in plasma of omnivorous

- and vegetarian women by isotope dilution gas chromatography-mass spectrometry. *Scand.J Clin Lab Invest Suppl* 1993;215:5-18.
25. Setchell KD, Brown NM, Desai P, Zimmer-Nechemias L, Wolfe BE, Brashear WT *et al.* Bioavailability of pure isoflavones in healthy humans and analysis of commercial soy isoflavone supplements. *J Nutr* 2001;131:1362S-75S.
 26. Morabito N, Crisafulli A, Vergara C, Gaudio A, Lasco A, Frisina N *et al.* Effects of genistein and hormone-replacement therapy on bone loss in early postmenopausal women: a randomized double-blind placebo-controlled study. *J Bone Miner.Res* 2002;17:1904-12.
 27. Lapcik O, Hampl R, Hill M, Wahala K, Maharik NA, Adlercreutz H. Radioimmunoassay of free genistein in human serum. *J Steroid Biochem Mol.Biol* 1998;64:261-68.
 28. Nagel SC, vom Saal FS, Welshons WV. The effective free fraction of estradiol and xenoestrogens in human serum measured by whole cell uptake assays: physiology of delivery modifies estrogenic activity. *Proc.Soc.Exp Biol Med* 1998;217:300-09.
 29. Akiyama T, Ishida J, Nakagawa S, Ogawara H, Watanabe S, Itoh N *et al.* Genistein, a specific inhibitor of tyrosine-specific protein kinases. *J Biol Chem* 1987;262:5592-95.
 30. Markovits J, Linassier C, Fosse P, Couprie J, Pierre J, Jacquemin-Sablon A *et al.* Inhibitory effects of the tyrosine kinase inhibitor genistein on mammalian DNA topoisomerase II. *Cancer Res* 1989;49:5111-17.
 31. Suetsugi M, Su L, Karlsberg K, Yuan YC, Chen S. Flavone and isoflavone phytoestrogens are agonists of estrogen-related receptors. *Mol.Cancer Res* 2003;1:981-91.
 32. Pau CY, Pau KY, Spies HG. Putative estrogen receptor beta and alpha mRNA expression in male and female rhesus macaques. *Mol.Cell Endocrinol.* 1998;146:59-68.
 33. Alvaro D, Alpini G, Onori P, Perego L, Svegliata BG, Franchitto A *et al.* Estrogens stimulate proliferation of intrahepatic biliary epithelium in rats. *Gastroenterology* 2000;119:1681-91.
 34. Taylor AH, Al Azzawi F. Immunolocalisation of oestrogen receptor beta in human tissues. *J Mol.Endocrinol.* 2000;24:145-55.
 35. Pelletier G. Localization of androgen and estrogen receptors in rat and primate tissues. *Histol.Histopathol.* 2000;15:1261-70.

36. Hillisch A, Peters O, Kosemund D, Muller G, Walter A, Schneider B *et al.* Dissecting physiological roles of estrogen receptor alpha and beta with potent selective ligands from structure-based design. *Mol.Endocrinol.* 2004;18:1599-609.
37. Vehkavaara S, Silveira A, Hakala-Ala-Pietila T, Virkamaki A, Hovatta O, Hamsten A *et al.* Effects of oral and transdermal estrogen replacement therapy on markers of coagulation, fibrinolysis, inflammation and serum lipids and lipoproteins in postmenopausal women. *Thromb.Haemost.* 2001;85:619-25.
38. Hemelaar M, van der Mooren MJ, Mijatovic V, Bouman AA, Schijf CP, Kroeks MV *et al.* Oral, more than transdermal, estrogen therapy improves lipids and lipoprotein(a) in postmenopausal women: a randomized, placebo-controlled study. *Menopause.* 2003;10:550-58.
39. Nanda S, Gupta N, Mehta HC, Sangwan K. Effect of oestrogen replacement therapy on serum lipid profile. *Aust.N.Z.J Obstet.Gynaecol.* 2003;43:213-16.
40. Schunkert H, Danser AH, Hense HW, Derkx FH, Kurzinger S, Riegger GA. Effects of estrogen replacement therapy on the renin-angiotensin system in postmenopausal women. *Circulation* 1997;95:39-45.
41. Helle SI, Omsjo IH, Hughes SC, Botta L, Huls G, Holly JM *et al.* Effects of oral and transdermal oestrogen replacement therapy on plasma levels of insulin-like growth factors and IGF binding proteins 1 and 3: a cross-over study. *Clin Endocrinol.(Oxf)* 1996;45:727-32.
42. Paassilta M, Karjalainen A, Kervinen K, Savolainen MJ, Heikkinen J, Backstrom AC *et al.* Insulin-like growth factor binding protein-1 (IGFBP-1) and IGF-I during oral and transdermal estrogen replacement therapy: relation to lipoprotein(a) levels. *Atherosclerosis* 2000;149:157-62.
43. Cardim HJ, Lopes CM, Giannella-Neto D, da Fonseca AM, Pinotti JA. The insulin-like growth factor-I system and hormone replacement therapy. *Fertil.Steril.* 2001;75:282-87.
44. Biglia N, Ambroggio S, Ponzzone R, Sgro L, Ujcic E, Dato FA *et al.* Modification of serum IGF-I, IGFBPs and SHBG levels by different HRT regimens. *Maturitas* 2003;45:283-91.
45. Scarabin PY, Alhenc-Gelas M, Plu-Bureau, Taisne P, Agher R, Aiach M. Effects of oral and transdermal estrogen/progesterone regimens on blood coagulation and fibrinolysis in postmenopausal women. A randomized controlled trial. *Arterioscler.Thromb.Vasc.Biol* 1997;17:3071-78.
46. Lowe GD, Upton MN, Rumley A, McConnachie A, O'Reilly DS, Watt GC. Different effects of oral and transdermal hormone replacement therapies on factor

- IX, APC resistance, t-PA, PAI and C-reactive protein--a cross-sectional population survey. *Thromb.Haemost.* 2001;86:550-56.
47. Lowe GD. Hormone replacement therapy: prothrombotic vs. protective effects. *Pathophysiol.Haemost.Thromb.* 2002;32:329-32.
 48. Post MS, Christella M, Thomassen LG, van der Mooren MJ, van Baal WM, Rosing J *et al.* Effect of oral and transdermal estrogen replacement therapy on hemostatic variables associated with venous thrombosis: a randomized, placebo-controlled study in postmenopausal women. *Arterioscler.Thromb.Vasc.Biol* 2003;23:1116-21.
 49. Oger E, Alhenc-Gelas M, Lacut K, Blouch MT, Roudaut N, Kerlan V *et al.* Differential effects of oral and transdermal estrogen/progesterone regimens on sensitivity to activated protein C among postmenopausal women: a randomized trial. *Arterioscler.Thromb.Vasc.Biol* 2003;23:1671-76.
 50. Scarabin PY, Oger E, Plu-Bureau. Differential association of oral and transdermal oestrogen-replacement therapy with venous thromboembolism risk. *Lancet* 2003;362:428-32.
 51. Greaves KA, Parks JS, Williams JK, Wagner JD. Intact dietary soy protein, but not adding an isoflavone-rich soy extract to casein, improves plasma lipids in ovariectomized cynomolgus monkeys. *J Nutr* 1999;129:1585-92.
 52. Lichtenstein AH, Jalbert SM, Adlercreutz H, Goldin BR, Rasmussen H, Schaefer EJ *et al.* Lipoprotein response to diets high in soy or animal protein with and without isoflavones in moderately hypercholesterolemic subjects. *Arterioscler.Thromb.Vasc.Biol* 2002;22:1852-58.
 53. Weggemans RM, Trautwein EA. Relation between soy-associated isoflavones and LDL and HDL cholesterol concentrations in humans: a meta-analysis. *Eur.J Clin Nutr* 2003;57:940-46.
 54. Demonty I, Lamarche B, Jones PJ. Role of isoflavones in the hypocholesterolemic effect of soy. *Nutr Rev.* 2003;61:189-203.
 55. Adams KF, Newton KM, Chen C, Emerson SS, Potter JD, White E *et al.* Soy isoflavones do not modulate circulating insulin-like growth factor concentrations in an older population in an intervention trial. *J Nutr* 2003;133:1316-19.
 56. Teede HJ, Dalais FS, McGrath BP. Dietary soy containing phytoestrogens does not have detectable estrogenic effects on hepatic protein synthesis in postmenopausal women. *Am J Clin Nutr* 2004;79:396-401.
 57. Wang HH, Afdhal NH, Wang DQ. Estrogen receptor alpha, but not beta, plays a major role in 17beta-estradiol-induced murine cholesterol gallstones. *Gastroenterology* 2004;127:239-49.

58. Carroll KK. Hypercholesterolemia and atherosclerosis: effects of dietary protein. *Fed Proc* 1982;41:2792-96.
59. Sirtori CR, Gianazza E, Manzoni C, Lovati MR, Murphy PA. Role of isoflavones in the cholesterol reduction by soy proteins in the clinic. *Am J Clin Nutr* 1997;65:166-67.
60. Sirtori CR, Agradi E, Conti F, Mantero O, Gatti E. Soybean-protein diet in the treatment of type-II hyperlipoproteinaemia. *Lancet* 1977;1:275-77.
61. Anderson JW, Johnstone BM, Cook-Newell ME. Meta-analysis of the effects of soy protein intake on serum lipids [see comments]. *N Engl J Med* 1995;333:276-82.
62. Shah SH, Alexander KP. Hormone Replacement Therapy for Primary and Secondary Prevention of Heart Disease. *Curr.Treat.Options.Cardiovasc.Med* 2003;5:25-33.
63. Couse JF, Korach KS. Contrasting phenotypes in reproductive tissues of female estrogen receptor null mice. *Ann.N.Y.Acad.Sci* 2001;948:1-8.
64. Duncan AM, Underhill KE, Xu X, Lavalleur J, Phipps WR, Kurzer MS. Modest hormonal effects of soy isoflavones in postmenopausal women. *J Clin Endocrinol.Metab* 1999;84:3479-84.
65. Kurzer MS. Hormonal effects of soy in premenopausal women and men. *J Nutr* 2002;132:570S-3S.
66. Balk JL, Whiteside DA, Naus G, DeFerrari E, Roberts JM. A pilot study of the effects of phytoestrogen supplementation on postmenopausal endometrium. *J Soc.Gynecol.Investig.* 2002;9:238-42.
67. Penotti M, Fabio E, Modena AB, Rinaldi M, Omodei U, Vigano P. Effect of soy-derived isoflavones on hot flushes, endometrial thickness, and the pulsatility index of the uterine and cerebral arteries. *Fertil.Steril.* 2003;79:1112-17.
68. Sammartino A, Di Carlo C, Mandato VD, Bifulco G, Di Stefano M, Nappi C. Effects of genistein on the endometrium: ultrasonographic evaluation. *Gynecol.Endocrinol.* 2003;17:45-49.
69. Murray MJ, Meyer WR, Lessey BA, Oi RH, DeWire RE, Fritz MA. Soy protein isolate with isoflavones does not prevent estradiol-induced endometrial hyperplasia in postmenopausal women: a pilot trial. *Menopause.* 2003;10:456-64.
70. Santell RC, Chang YC, Nair MG, Helferich WG. Dietary genistein exerts estrogenic effects upon the uterus, mammary gland and the hypothalamic/pituitary axis in rats. *J Nutr* 1997;127:263-69.

71. Chambliss KL, Shaul PW. Estrogen modulation of endothelial nitric oxide synthase. *Endocr.Rev.* 2002;23:665-86.
72. McNeill AM, Kim N, Duckles SP, Krause DN, Kontos HA. Chronic estrogen treatment increases levels of endothelial nitric oxide synthase protein in rat cerebral microvessels. *Stroke* 1999;30:2186-90.
73. Yang S, Bae L, Zhang L. Estrogen increases eNOS and NOx release in human coronary artery endothelium. *J Cardiovasc.Pharmacol* 2000;36:242-47.
74. Stirone C, Chu Y, Sunday L, Duckles SP, Krause DN. 17 Beta-estradiol increases endothelial nitric oxide synthase mRNA copy number in cerebral blood vessels: quantification by real-time polymerase chain reaction. *Eur.J Pharmacol* 2003;478:35-38.
75. Haynes MP, Li L, Sinha D, Russell KS, Hisamoto K, Baron R *et al.* Src kinase mediates phosphatidylinositol 3-kinase/Akt-dependent rapid endothelial nitric-oxide synthase activation by estrogen. *J Biol Chem* 2003;278:2118-23.
76. Chambliss KL, Yuhanna IS, Anderson RG, Mendelsohn ME, Shaul PW. ERbeta has nongenomic action in caveolae. *Mol.Endocrinol.* 2002;16:938-46.
77. Wagner AH, Schroeter MR, Hecker M. 17beta-estradiol inhibition of NADPH oxidase expression in human endothelial cells. *FASEB J* 2001;15:2121-30.
78. Evans MJ, Harris HA, Miller CP, Karathanasis SK, Adelman SJ. Estrogen receptors alpha and beta have similar activities in multiple endothelial cell pathways. *Endocrinology* 2002;143:3785-95.
79. Rubanyi GM, Freay AD, Kauser K, Sukovich D, Burton G, Lubahn DB *et al.* Vascular estrogen receptors and endothelium-derived nitric oxide production in the mouse aorta. Gender difference and effect of estrogen receptor gene disruption. *J Clin Invest* 1997;99:2429-37.
80. Tan E, Gurjar MV, Sharma RV, Bhalla RC. Estrogen receptor-alpha gene transfer into bovine aortic endothelial cells induces eNOS gene expression and inhibits cell migration. *Cardiovasc.Res* 1999;43:788-97.
81. Geary GG, McNeill AM, Ospina JA, Krause DN, Korach KS, Duckles SP. Selected contribution: cerebrovascular nos and cyclooxygenase are unaffected by estrogen in mice lacking estrogen receptor-alpha. *J Appl.Physiol* 2001;91:2391-99.
82. Rubanyi GM, Kauser K, Johns A. Role of estrogen receptors in the vascular system. *Vascul.Pharmacol* 2002;38:81-88.

83. Darblade B, Pendaries C, Krust A, Dupont S, Fouque MJ, Rami J *et al.* Estradiol alters nitric oxide production in the mouse aorta through the alpha-, but not beta-, estrogen receptor. *Circ.Res* 2002;90:413-19.
84. Widder J, Pelzer T, Poser-Klein C, Hu K, Jazbutyte V, Fritzemeier KH *et al.* Improvement of endothelial dysfunction by selective estrogen receptor-alpha stimulation in ovariectomized SHR. *Hypertension* 2003;42:991-96.
85. Muller-Delp JM, Lubahn DB, Nichol KE, Philips BJ, Price EM, Curran EM *et al.* Regulation of nitric oxide-dependent vasodilation in coronary arteries of estrogen receptor-alpha-deficient mice. *Am J Physiol Heart Circ.Physiol* 2003;285:H2150-H2157.
86. Nuedling S, Karas RH, Mendelsohn ME, Katzenellenbogen JA, Katzenellenbogen BS, Meyer R *et al.* Activation of estrogen receptor beta is a prerequisite for estrogen-dependent upregulation of nitric oxide synthases in neonatal rat cardiac myocytes. *FEBS Lett.* 2001;502:103-08.
87. Kakui K, Itoh H, Sagawa N, Yura S, Korita D, Takemura M *et al.* Augmented endothelial nitric oxide synthase (eNOS) protein expression in human pregnant myometrium: possible involvement of eNOS promoter activation by estrogen via both estrogen receptor (ER)alpha and ERbeta. *Mol.Hum.Reprod.* 2004;10:115-22.
88. Zhu Y, Bian Z, Lu P, Karas RH, Bao L, Cox D *et al.* Abnormal vascular function and hypertension in mice deficient in estrogen receptor beta. *Science* 2002;295:505-08.
89. Makela S, Savolainen H, Aavik E, Myllarniemi M, Strauss L, Taskinen E *et al.* Differentiation between vasculoprotective and uterotrophic effects of ligands with different binding affinities to estrogen receptors alpha and beta. *Proc.Natl.Acad.Sci U.S.A* 1999;96:7077-82.
90. Squadrito F, Altavilla D, Morabito N, Crisafulli A, D'Anna R, Corrado F *et al.* The effect of the phytoestrogen genistein on plasma nitric oxide concentrations, endothelin-1 levels and endothelium dependent vasodilation in postmenopausal women. *Atherosclerosis* 2002;163:339-47.
91. Squadrito F, Altavilla D, Crisafulli A, Saitta A, Cucinotta D, Morabito N *et al.* Effect of genistein on endothelial function in postmenopausal women: a randomized, double-blind, controlled study. *Am J Med* 2003;114:470-76.
92. Steinberg FM, Guthrie NL, Villablanca AC, Kumar K, Murray MJ. Soy protein with isoflavones has favorable effects on endothelial function that are independent of lipid and antioxidant effects in healthy postmenopausal women. *Am J Clin Nutr* 2003;78:123-30.
93. Lissin LW, Oka R, Lakshmi S, Cooke JP. Isoflavones improve vascular reactivity in post-menopausal women with hypercholesterolemia. *Vasc.Med* 2004;9:26-30.

94. Vanhoutte PM. Say NO to ET. *J Auton.Nerv.Syst.* 2000;81:271-77.
95. Simons LA, Von Konigsmark M, Simons J, Celermajer DS. Phytoestrogens do not influence lipoprotein levels or endothelial function in healthy, postmenopausal women. *Am J Cardiol.* 2000;85:1297-301.
96. Hale G, Paul-Labrador M, Dwyer JH, Merz CN. Isoflavone supplementation and endothelial function in menopausal women. *Clin Endocrinol.(Oxf)* 2002;56:693-701.
97. Nestel PJ, Yamashita T, Sasahara T, Pomeroy S, Dart A, Komesaroff P *et al.* Soy isoflavones improve systemic arterial compliance but not plasma lipids in menopausal and perimenopausal women. *Arterioscler.Thromb.Vasc.Biol* 1997;17:3392-98.
98. Squadrito F, Altavilla D, Squadrito G, Saitta A, Cucinotta D, Minutoli L *et al.* Genistein supplementation and estrogen replacement therapy improve endothelial dysfunction induced by ovariectomy in rats. *Cardiovasc.Res* 2000;45:454-62.
99. Catania MA, Crupi A, Firenzuoli F, Parisi A, Sturiale A, Squadrito F *et al.* Oral administration of a soy extract improves endothelial dysfunction in ovariectomized rats. *Planta Med* 2002;68:1142-44.
100. Khemapech S, Monsiri K, Patumraj S, Siriviriyakul P. Genistein replacement therapy for vasodilation disorder in bilateral ovariectomized rats. *Clin Hemorheol.Microcirc.* 2003;29:271-77.
101. Izumi T, Piskula MK, Osawa S, Obata A, Tobe K, Saito M *et al.* Soy isoflavone aglycones are absorbed faster and in higher amounts than their glucosides in humans. *J Nutr* 2000;130:1695-99.
102. Setchell KD, Brown NM, Desai P, Zimmer-Nechemias L, Wolfe BE, Brashear WT *et al.* Bioavailability of pure isoflavones in healthy humans and analysis of commercial soy isoflavone supplements. *J Nutr* 2001;131:1362S-75S.
103. Richelle M, Pridmore-Merten S, Bodenstab S, Enslin M, Offord EA. Hydrolysis of isoflavone glycosides to aglycones by beta-glycosidase does not alter plasma and urine isoflavone pharmacokinetics in postmenopausal women. *J Nutr* 2002;132:2587-92.
104. Zubik L, Meydani M. Bioavailability of soybean isoflavones from aglycone and glucoside forms in American women. *Am J Clin Nutr* 2003;77:1459-65.
105. Setchell KD, Brown NM, Zimmer-Nechemias L, Brashear WT, Wolfe BE, Kirschner AS *et al.* Evidence for lack of absorption of soy isoflavone glycosides in humans, supporting the crucial role of intestinal metabolism for bioavailability. *Am J Clin Nutr* 2002;76:447-53.

106. Day AJ, DuPont MS, Ridley S, Rhodes M, Rhodes MJ, Morgan MR *et al.* Deglycosylation of flavonoid and isoflavonoid glycosides by human small intestine and liver beta-glucosidase activity. *FEBS Lett.* 1998;436:71-75.
107. Murota K, Shimizu S, Miyamoto S, Izumi T, Obata A, Kikuchi M *et al.* Unique uptake and transport of isoflavone aglycones by human intestinal caco-2 cells: comparison of isoflavonoids and flavonoids. *J Nutr* 2002;132:1956-61.
108. Xu X, Harris KS, Wang HJ, Murphy PA, Hendrich S. Bioavailability of soybean isoflavones depends upon gut microflora in women. *J Nutr* 1995;125:2307-15.
109. Walker HA, Dean TS, Sanders TA, Jackson G, Ritter JM, Chowienczyk PJ. The phytoestrogen genistein produces acute nitric oxide-dependent dilation of human forearm vasculature with similar potency to 17beta-estradiol. *Circulation* 2001;103:258-62.
110. Teede HJ, Dalais FS, Kotsopoulos D, Liang YL, Davis S, McGrath BP. Dietary soy has both beneficial and potentially adverse cardiovascular effects: a placebo-controlled study in men and postmenopausal women. *J Clin Endocrinol.Metab* 2001;86:3053-60.
111. McCrohon JA, Walters WA, Robinson JT, McCredie RJ, Turner L, Adams MR *et al.* Arterial reactivity is enhanced in genetic males taking high dose estrogens. *J Am Coll.Cardiol.* 1997;29:1432-36.
112. New G, Duffy SJ, Harper RW, Meredith IT. Long-term oestrogen therapy is associated with improved endothelium-dependent vasodilation in the forearm resistance circulation of biological males. *Clin Exp Pharmacol Physiol* 2000;27:25-33.
113. Sader MA, McCredie RJ, Griffiths KA, Wishart SM, Handelsman DJ, Celermajer DS. Oestradiol improves arterial endothelial function in healthy men receiving testosterone. *Clin Endocrinol.(Oxf)* 2001;54:175-81.
114. Kimura M, Sudhir K, Jones M, Simpson E, Jefferis AM, Chin-Dusting JP. Impaired acetylcholine-induced release of nitric oxide in the aorta of male aromatase-knockout mice: regulation of nitric oxide production by endogenous sex hormones in males. *Circ.Res* 2003;93:1267-71.
115. Hanke H, Hanke S, Ickrath O, Lange K, Bruck B, Muck AO *et al.* Estradiol concentrations in premenopausal women with coronary heart disease. *Coron.Artery Dis.* 1997;8:511-15.
116. Christian RC, Harrington S, Edwards WD, Oberg AL, Fitzpatrick LA. Estrogen status correlates with the calcium content of coronary atherosclerotic plaques in women. *J Clin Endocrinol.Metab* 2002;87:1062-67.

117. Bairey Merz CN, Johnson BD, Sharaf BL, Bittner V, Berga SL, Braunstein GD *et al.* Hypoestrogenemia of hypothalamic origin and coronary artery disease in premenopausal women: a report from the NHLBI-sponsored WISE study. *J Am Coll.Cardiol.* 2003;41:413-19.
118. Vasquez-Vivar J, Kalyanaraman B, Martasek P, Hogg N, Masters BS, Karoui H *et al.* Superoxide generation by endothelial nitric oxide synthase: the influence of cofactors. *Proc.Natl.Acad.Sci U.S.A* 1998;95:9220-25.
119. Milstien S, Katusic Z. Oxidation of tetrahydrobiopterin by peroxynitrite: implications for vascular endothelial function. *Biochem Biophys Res Commun.* 1999;263:681-84.
120. Vasquez-Vivar J, Kalyanaraman B, Martasek P. The role of tetrahydrobiopterin in superoxide generation from eNOS: enzymology and physiological implications. *Free Radic.Res* 2003;37:121-27.
121. Stroes ES, van Faassen EE, Yo M, Martasek P, Boer P, Govers R *et al.* Folic acid reverts dysfunction of endothelial nitric oxide synthase. *Circ.Res* 2000;86:1129-34.
122. Verhaar MC, Stroes E, Rabelink TJ. Folates and cardiovascular disease. *Arterioscler.Thromb.Vasc.Biol.* 2002;22:6-13.
123. Hyndman ME, Verma S, Rosenfeld RJ, Anderson TJ, Parsons HG. Interaction of 5-methyltetrahydrofolate and tetrahydrobiopterin on endothelial function. *Am J Physiol Heart Circ.Physiol* 2002;282:H2167-H2172.
124. Doshi SN, McDowell IF, Moat SJ, Payne N, Durrant HJ, Lewis MJ *et al.* Folic acid improves endothelial function in coronary artery disease via mechanisms largely independent of homocysteine lowering. *Circulation JID - 0147763* 2002;105:22-26.
125. McCarty MF. Coping with endothelial superoxide: potential complementarity of arginine and high-dose folate. *Med Hypotheses* 2004;63:709-18.
126. Cooke JP, Oka RK. Atherogenesis and the arginine hypothesis. *Curr Atheroscler Rep JID - 100897685* 2001;3:252-59.
127. Boger RH. The emerging role of asymmetric dimethylarginine as a novel cardiovascular risk factor. *Cardiovasc.Res* 2003;59:824-33.
128. Vidal O, Kindblom LG, Ohlsson C. Expression and localization of estrogen receptor-beta in murine and human bone. *J Bone Miner.Res* 1999;14:923-29.
129. Bord S, Horner A, Beavan S, Compston J. Estrogen receptors alpha and beta are differentially expressed in developing human bone. *J Clin Endocrinol.Metab* 2001;86:2309-14.

130. Batra GS, Hainey L, Freemont AJ, Andrew G, Saunders PT, Hoyland JA *et al.* Evidence for cell-specific changes with age in expression of oestrogen receptor (ER) alpha and beta in bone fractures from men and women. *J Pathol.* 2003;200:65-73.
131. Monroe DG, Getz BJ, Johnsen SA, Riggs BL, Khosla S, Spelsberg TC. Estrogen receptor isoform-specific regulation of endogenous gene expression in human osteoblastic cell lines expressing either ERalpha or ERbeta. *J Cell Biochem* 2003;90:315-26.
132. Lindberg MK, Moverare S, Skrtic S, Gao H, Dahlman-Wright K, Gustafsson JA *et al.* Estrogen receptor (ER)-beta reduces ERalpha-regulated gene transcription, supporting a "ying yang" relationship between ERalpha and ERbeta in mice. *Mol.Endocrinol.* 2003;17:203-08.
133. Windahl SH, Hollberg K, Vidal O, Gustafsson JA, Ohlsson C, Andersson G. Female estrogen receptor beta-/- mice are partially protected against age-related trabecular bone loss. *J Bone Miner.Res* 2001;16:1388-98.
134. Sims NA, Clement-Lacroix P, Minet D, Fraslon-Vanhulle C, Gaillard-Kelly M, Resche-Rigon M *et al.* A functional androgen receptor is not sufficient to allow estradiol to protect bone after gonadectomy in estradiol receptor-deficient mice. *J Clin Invest* 2003;111:1319-27.
135. Sims NA, Dupont S, Krust A, Clement-Lacroix P, Minet D, Resche-Rigon M *et al.* Deletion of estrogen receptors reveals a regulatory role for estrogen receptors-beta in bone remodeling in females but not in males. *Bone* 2002;30:18-25.
136. Cao L, Bu R, Oakley JI, Kalla SE, Blair HC. Estrogen receptor-beta modulates synthesis of bone matrix proteins in human osteoblast-like MG63 cells. *J Cell Biochem* 2003;89:152-64.
137. Koka S, Petro TM, Reinhardt RA. Estrogen inhibits interleukin-1beta-induced interleukin-6 production by human osteoblast-like cells. *J Interferon Cytokine Res* 1998;18:479-83.
138. Bord S, Ireland DC, Beavan SR, Compston JE. The effects of estrogen on osteoprotegerin, RANKL, and estrogen receptor expression in human osteoblasts. *Bone* 2003;32:136-41.
139. Chen X, Garner SC, Quarles LD, Anderson JJ. Effects of genistein on expression of bone markers during MC3T3-E1 osteoblastic cell differentiation. *J Nutr Biochem* 2003;14:342-49.
140. Viereck V, Grundker C, Blaschke S, Siggelkow H, Emons G, Hofbauer LC. Phytoestrogen genistein stimulates the production of osteoprotegerin by human trabecular osteoblasts. *J Cell Biochem* 2002;84:725-35.

141. Aguirre J, BATTERY L, O'Shaughnessy M, Afzal F, Fernandez dM, I, Hukkanen M *et al.* Endothelial nitric oxide synthase gene-deficient mice demonstrate marked retardation in postnatal bone formation, reduced bone volume, and defects in osteoblast maturation and activity. *Am J Pathol.* 2001;158:247-57.
142. Samuels A, Perry MJ, Gibson RL, Colley S, Tobias JH. Role of endothelial nitric oxide synthase in estrogen-induced osteogenesis. *Bone* 2001;29:24-29.
143. McFarlane SI, Muniyappa R, Shin JJ, Bahtiyar G, Sowers JR. Osteoporosis and cardiovascular disease: brittle bones and banded arteries, is there a link? *Endocrine.* 2004;23:1-10.
144. Anderson JJ, Ambrose WW, Garner SC. Biphasic effects of genistein on bone tissue in the ovariectomized, lactating rat model. *Proc.Soc.Exp Biol Med* 1998;217:345-50.
145. Fanti P, Monier-Faugere MC, Geng Z, Schmidt J, Morris PE, Cohen D *et al.* The phytoestrogen genistein reduces bone loss in short-term ovariectomized rats. *Osteoporos.Int* 1998;8:274-81.
146. Ishimi Y, Miyaura C, Ohmura M, Onoe Y, Sato T, Uchiyama Y *et al.* Selective effects of genistein, a soybean isoflavone, on B-lymphopoiesis and bone loss caused by estrogen deficiency. *Endocrinology* 1999;140:1893-900.
147. Wu J, Wang XX, Takasaki M, Ohta A, Higuchi M, Ishimi Y. Cooperative effects of exercise training and genistein administration on bone mass in ovariectomized mice. *J Bone Miner.Res* 2001;16:1829-36.
148. Nakajima D, Kim CS, Oh TW, Yang CY, Naka T, Igawa S *et al.* Suppressive effects of genistein dosage and resistance exercise on bone loss in ovariectomized rats. *J Physiol Anthropol.Appl.Human Sci* 2001;20:285-91.
149. Li B, Yu S. Genistein prevents bone resorption diseases by inhibiting bone resorption and stimulating bone formation. *Biol Pharm.Bull.* 2003;26:780-86.
150. Harris HA, Albert LM, Leathurby Y, Malamas MS, Mewshaw RE, Miller CP *et al.* Evaluation of an estrogen receptor-beta agonist in animal models of human disease. *Endocrinology* 2003;144:4241-49.
151. Potter SM, Baum JA, Teng H, Stillman RJ, Shay NF, Erdman JW, Jr. Soy protein and isoflavones: their effects on blood lipids and bone density in postmenopausal women. *Am J Clin Nutr* 1998;68:1375S-9S.
152. Uesugi T, Fukui Y, Yamori Y. Beneficial effects of soybean isoflavone supplementation on bone metabolism and serum lipids in postmenopausal Japanese women: a four-week study. *J Am Coll.Nutr* 2002;21:97-102.

153. Yamori Y, Moriguchi EH, Teramoto T, Miura A, Fukui Y, Honda KI *et al.* Soybean isoflavones reduce postmenopausal bone resorption in female Japanese immigrants in Brazil: a ten-week study. *J Am Coll.Nutr* 2002;21:560-63.
154. Chen YM, Ho SC, Lam SS, Ho SS, Woo JL. Soy isoflavones have a favorable effect on bone loss in Chinese postmenopausal women with lower bone mass: a double-blind, randomized, controlled trial. *J Clin Endocrinol.Metab* 2003;88:4740-47.
155. Chen YM, Ho SC, Lam SS, Ho SS, Woo JL. Beneficial effect of soy isoflavones on bone mineral content was modified by years since menopause, body weight, and calcium intake: a double-blind, randomized, controlled trial. *Menopause.* 2004;11:246-54.
156. Crisafulli A, Altavilla D, Squadrito G, Romeo A, Adamo EB, Marini R *et al.* Effects of the phytoestrogen genistein on the circulating soluble receptor activator of nuclear factor kappaB ligand-osteoprotegerin system in early postmenopausal women. *J Clin Endocrinol.Metab* 2004;89:188-92.
157. Hsu CS, Shen WW, Hsueh YM, Yeh SL. Soy isoflavone supplementation in postmenopausal women. Effects on plasma lipids, antioxidant enzyme activities and bone density. *J Reprod.Med* 2001;46:221-26.
158. Gallagher JC, Satpathy R, Rafferty K, Haynatzka V. The effect of soy protein isolate on bone metabolism. *Menopause.* 2004;11:290-98.
159. Kreijkamp-Kaspers S, Kok L, Grobbee DE, de Haan EH, Aleman A, Lampe JW *et al.* Effect of soy protein containing isoflavones on cognitive function, bone mineral density, and plasma lipids in postmenopausal women: a randomized controlled trial. *JAMA* 2004;292:65-74.
160. Anderson JJ, Chen X, Boass A, Symons M, Kohlmeier M, Renner JB *et al.* Soy isoflavones: no effects on bone mineral content and bone mineral density in healthy, menstruating young adult women after one year. *J Am Coll.Nutr* 2002;21:388-93.
161. Khalil DA, Lucas EA, Juma S, Smith BJ, Payton ME, Arjmandi BH. Soy protein supplementation increases serum insulin-like growth factor-I in young and old men but does not affect markers of bone metabolism. *J Nutr* 2002;132:2605-08.
162. Somekawa Y, Chiguchi M, Ishibashi T, Aso T. Soy intake related to menopausal symptoms, serum lipids, and bone mineral density in postmenopausal Japanese women. *Obstet.Gynecol.* 2001;97:109-15.
163. Ho SC, Woo J, Lam S, Chen Y, Sham A, Lau J. Soy protein consumption and bone mass in early postmenopausal Chinese women. *Osteoporos.Int* 2003;14:835-42.

164. Kardinaal AF, Morton MS, Bruggemann-Rotgans IE, van Beresteijn EC. Phyto-oestrogen excretion and rate of bone loss in postmenopausal women. *Eur.J Clin Nutr* 1998;52:850-55.
165. Kim MK, Chung BC, Yu VY, Nam JH, Lee HC, Huh KB *et al.* Relationships of urinary phyto-oestrogen excretion to BMD in postmenopausal women. *Clin Endocrinol.(Oxf)* 2002;56:321-28.
166. Nagata C, Shimizu H, Takami R, Hayashi M, Takeda N, Yasuda K. Soy product intake and serum isoflavonoid and estradiol concentrations in relation to bone mineral density in postmenopausal Japanese women. *Osteoporos.Int* 2002;13:200-04.
167. Kritz-Silverstein D, Goodman-Gruen DL. Usual dietary isoflavone intake, bone mineral density, and bone metabolism in postmenopausal women. *J Womens Health Gend.Based.Med* 2002;11:69-78.
168. Royuela M, de Miguel MP, Bethencourt FR, Sanchez-Chapado M, Fraile B, Arenas MI *et al.* Estrogen receptors alpha and beta in the normal, hyperplastic and carcinomatous human prostate. *J Endocrinol.* 2001;168:447-54.
169. Fixemer T, Remberger K, Bonkhoff H. Differential expression of the estrogen receptor beta (ERbeta) in human prostate tissue, premalignant changes, and in primary, metastatic, and recurrent prostatic adenocarcinoma. *Prostate* 2003;54:79-87.
170. Signoretti S, Loda M. Estrogen receptor beta in prostate cancer: brake pedal or accelerator? *Am J Pathol.* 2001;159:13-16.
171. Ho SM. Estrogens and anti-estrogens: key mediators of prostate carcinogenesis and new therapeutic candidates. *J Cell Biochem* 2004;91:491-503.
172. Kregel JH, Hodgin JB, Couse JF, Enmark E, Warner M, Mahler JF *et al.* Generation and reproductive phenotypes of mice lacking estrogen receptor beta. *Proc.Natl.Acad.Sci U.S.A* 1998;95:15677-82.
173. Weihua Z, Makela S, Andersson LC, Salmi S, Saji S, Webster JI *et al.* A role for estrogen receptor beta in the regulation of growth of the ventral prostate. *Proc.Natl.Acad.Sci U.S.A* 2001;98:6330-35.
174. Cheng J, Lee EJ, Madison LD, Lazennec G. Expression of estrogen receptor beta in prostate carcinoma cells inhibits invasion and proliferation and triggers apoptosis. *FEBS Lett.* 2004;566:169-72.
175. Zhu X, Leav I, Leung YK, Wu M, Liu Q, Gao Y *et al.* Dynamic regulation of estrogen receptor-beta expression by DNA methylation during prostate cancer development and metastasis. *Am J Pathol.* 2004;164:2003-12.

176. Linja MJ, Savinainen KJ, Tammela TL, Isola JJ, Visakorpi T. Expression of ERalpha and ERbeta in prostate cancer. *Prostate* 2003;55:180-86.
177. Lau KM, LaSpina M, Long J, Ho SM. Expression of estrogen receptor (ER)-alpha and ER-beta in normal and malignant prostatic epithelial cells: regulation by methylation and involvement in growth regulation. *Cancer Res* 2000;60:3175-82.
178. Kim IY, Seong dH, Kim BC, Lee DK, Remaley AT, Leach F *et al.* Raloxifene, a selective estrogen receptor modulator, induces apoptosis in androgen-responsive human prostate cancer cell line LNCaP through an androgen-independent pathway. *Cancer Res* 2002;62:3649-53.
179. Kim IY, Kim BC, Seong dH, Lee DK, Seo JM, Hong YJ *et al.* Raloxifene, a mixed estrogen agonist/antagonist, induces apoptosis in androgen-independent human prostate cancer cell lines. *Cancer Res* 2002;62:5365-69.
180. Bemis DL, Capodice JL, Desai M, Buttyan R, Katz AE. A concentrated aglycone isoflavone preparation (GCP) that demonstrates potent anti-prostate cancer activity in vitro and in vivo. *Clin Cancer Res* 2004;10:5282-92.
181. Bektic J, Berger AP, Pfeil K, Dobler G, Bartsch G, Klocker H. Androgen receptor regulation by physiological concentrations of the isoflavonoid genistein in androgen-dependent LNCaP cells is mediated by estrogen receptor beta. *Eur.Urol.* 2004;45:245-51.
182. Zhou JR, Gugger ET, Tanaka T, Guo Y, Blackburn GL, Clinton SK. Soybean phytochemicals inhibit the growth of transplantable human prostate carcinoma and tumor angiogenesis in mice. *J Nutr* 1999;129:1628-35.
183. Aronson WJ, Tymchuk CN, Elashoff RM, McBride WH, McLean C, Wang H *et al.* Decreased growth of human prostate LNCaP tumors in SCID mice fed a low-fat, soy protein diet with isoflavones. *Nutr Cancer* 1999;35:130-36.
184. Fritz WA, Wang J, Eltoum IE, Lamartiniere CA. Dietary genistein down-regulates androgen and estrogen receptor expression in the rat prostate. *Mol.Cell Endocrinol.* 2002;186:89-99.
185. Wang J, Eltoum IE, Lamartiniere CA. Dietary genistein suppresses chemically induced prostate cancer in Lobund-Wistar rats. *Cancer Lett.* 2002;186:11-18.
186. Mentor-Marcel R, Lamartiniere CA, Eltoum IE, Greenberg NM, Elgavish A. Genistein in the diet reduces the incidence of poorly differentiated prostatic adenocarcinoma in transgenic mice (TRAMP). *Cancer Res* 2001;61:6777-82.
187. Hussain M, Banerjee M, Sarkar FH, Djuric Z, Pollak MN, Doerge D *et al.* Soy isoflavones in the treatment of prostate cancer. *Nutr Cancer* 2003;47:111-17.

188. deVere White RW, Hackman RM, Soares SE, Beckett LA, Li Y, Sun B. Effects of a genistein-rich extract on PSA levels in men with a history of prostate cancer. *Urology* 2004;63:259-63.
189. Messina MJ. Emerging evidence on the role of soy in reducing prostate cancer risk. *Nutr Rev.* 2003;61:117-31.
190. Lee MM, Gomez SL, Chang JS, Wey M, Wang RT, Hsing AW. Soy and isoflavone consumption in relation to prostate cancer risk in China. *Cancer Epidemiol.Biomarkers Prev.* 2003;12:665-68.
191. Ozasa K, Nakao M, Watanabe Y, Hayashi K, Miki T, Mikami K *et al.* Serum phytoestrogens and prostate cancer risk in a nested case-control study among Japanese men. *Cancer Sci* 2004;95:65-71.
192. Pelletier G, El Alfy M. Immunocytochemical localization of estrogen receptors alpha and beta in the human reproductive organs. *J Clin Endocrinol.Metab* 2000;85:4835-40.
193. Palmieri C, Saji S, Sakaguchi H, Cheng G, Sunters A, O'Hare MJ *et al.* The expression of oestrogen receptor (ER)-beta and its variants, but not ERalpha, in adult human mammary fibroblasts. *J Mol.Endocrinol.* 2004;33:35-50.
194. Couse JF, Korach KS. Estrogen receptor null mice: what have we learned and where will they lead us? *Endocr.Rev.* 1999;20:358-417.
195. Gustafsson JA, Warner M. Estrogen receptor beta in the breast: role in estrogen responsiveness and development of breast cancer. *J Steroid Biochem Mol.Biol* 2000;74:245-48.
196. Paruthiyil S, Parmar H, Kerekatte V, Cunha GR, Firestone GL, Leitman DC. Estrogen receptor beta inhibits human breast cancer cell proliferation and tumor formation by causing a G2 cell cycle arrest. *Cancer Res* 2004;64:423-28.
197. Strom A, Hartman J, Foster JS, Kietz S, Wimalasena J, Gustafsson JA. Estrogen receptor beta inhibits 17beta-estradiol-stimulated proliferation of the breast cancer cell line T47D. *Proc.Natl.Acad.Sci U.S.A* 2004;101:1566-71.
198. Hou YF, Yuan ST, Li HC, Wu J, Lu JS, Liu G *et al.* ERbeta exerts multiple stimulative effects on human breast carcinoma cells. *Oncogene* 2004;23:5799-806.
199. Dotzlaw H, Leygue E, Watson PH, Murphy LC. Expression of estrogen receptor-beta in human breast tumors. *J Clin Endocrinol.Metab* 1997;82:2371-74.
200. Fuqua SA, Schiff R, Parra I, Moore JT, Mohsin SK, Osborne CK *et al.* Estrogen receptor beta protein in human breast cancer: correlation with clinical tumor parameters. *Cancer Res* 2003;63:2434-39.

201. Murrill WB, Brown NM, Zhang JX, Manzolillo PA, Barnes S, Lamartiniere CA. Prepubertal genistein exposure suppresses mammary cancer and enhances gland differentiation in rats. *Carcinogenesis* 1996;17:1451-57.
202. Lamartiniere CA, Murrill WB, Manzolillo PA, Zhang JX, Barnes S, Zhang X *et al.* Genistein alters the ontogeny of mammary gland development and protects against chemically-induced mammary cancer in rats. *Proc.Soc.Exp Biol Med* 1998;217:358-64.
203. Cabanes A, Wang M, Olivo S, DeAssis S, Gustafsson JA, Khan G *et al.* Prepubertal estradiol and genistein exposures up-regulate BRCA1 mRNA and reduce mammary tumorigenesis. *Carcinogenesis* 2004;25:741-48.
204. Jin Z, MacDonald RS. Soy isoflavones increase latency of spontaneous mammary tumors in mice. *J Nutr* 2002;132:3186-90.
205. Yuan L, Wagatsuma C, Yoshida M, Miura T, Mukoda T, Fujii H *et al.* Inhibition of human breast cancer growth by GCP (genistein combined polysaccharide) in xenogeneic athymic mice: involvement of genistein biotransformation by beta-glucuronidase from tumor tissues. *Mutat.Res* 2003;523-524:55-62.
206. Mizunuma H, Kanazawa K, Ogura S, Otsuka S, Nagai H. Anticarcinogenic effects of isoflavones may be mediated by genistein in mouse mammary tumor virus-induced breast cancer. *Oncology* 2002;62:78-84.
207. Hsieh CY, Santell RC, Haslam SZ, Helferich WG. Estrogenic effects of genistein on the growth of estrogen receptor-positive human breast cancer (MCF-7) cells in vitro and in vivo. *Cancer Res* 1998;58:3833-38.
208. Allred CD, Allred KF, Ju YH, Virant SM, Helferich WG. Soy diets containing varying amounts of genistein stimulate growth of estrogen-dependent (MCF-7) tumors in a dose-dependent manner. *Cancer Res* 2001;61:5045-50.
209. Ju YH, Allred CD, Allred KF, Karko KL, Doerge DR, Helferich WG. Physiological concentrations of dietary genistein dose-dependently stimulate growth of estrogen-dependent human breast cancer (MCF-7) tumors implanted in athymic nude mice. *J Nutr* 2001;131:2957-62.
210. Ju YH, Doerge DR, Allred KF, Allred CD, Helferich WG. Dietary genistein negates the inhibitory effect of tamoxifen on growth of estrogen-dependent human breast cancer (MCF-7) cells implanted in athymic mice. *Cancer Res* 2002;62:2474-77.
211. Maggiolini M, Bonofiglio D, Marsico S, Panno ML, Cenni B, Picard D *et al.* Estrogen receptor alpha mediates the proliferative but not the cytotoxic dose-dependent effects of two major phytoestrogens on human breast cancer cells. *Mol.Pharmacol* 2001;60:595-602.

212. Allred CD, Allred KF, Ju YH, Clausen LM, Doerge DR, Schantz SL *et al.* Dietary genistein results in larger MNU-induced, estrogen-dependent mammary tumors following ovariectomy of Sprague-Dawley rats. *Carcinogenesis* 2004;25:211-18.
213. Day JK, Besch-Williford C, McMann TR, Hufford MG, Lubahn DB, MacDonald RS. Dietary genistein increased DMBA-induced mammary adenocarcinoma in wild-type, but not ER alpha KO, mice. *Nutr Cancer* 2001;39:226-32.
214. Hirohata T, Shigematsu T, Nomura AM, Nomura Y, Horie A, Hirohata I. Occurrence of breast cancer in relation to diet and reproductive history: a case-control study in Fukuoka, Japan. *Natl.Cancer Inst.Monogr* 1985;69:187-90.
215. Yuan JM, Wang QS, Ross RK, Henderson BE, Yu MC. Diet and breast cancer in Shanghai and Tianjin, China. *Br.J Cancer* 1995;71:1353-58.
216. Key TJ, Sharp GB, Appleby PN, Beral V, Goodman MT, Soda M *et al.* Soya foods and breast cancer risk: a prospective study in Hiroshima and Nagasaki, Japan. *Br.J Cancer* 1999;81:1248-56.
217. Horn-Ross PL, John EM, Lee M, Stewart SL, Koo J, Sakoda LC *et al.* Phytoestrogen consumption and breast cancer risk in a multiethnic population: the Bay Area Breast Cancer Study. *Am J Epidemiol.* 2001;154:434-41.
218. Horn-Ross PL, Hoggatt KJ, West DW, Krone MR, Stewart SL, Anton H *et al.* Recent diet and breast cancer risk: the California Teachers Study (USA). *Cancer Causes Control* 2002;13:407-15.
219. Lee HP, Gourley L, Duffy SW, Esteve J, Lee J, Day NE. Risk factors for breast cancer by age and menopausal status: a case-control study in Singapore. *Cancer Causes Control* 1992;3:313-22.
220. Hirose K, Tajima K, Hamajima N, Inoue M, Takezaki T, Kuroishi T *et al.* A large-scale, hospital-based case-control study of risk factors of breast cancer according to menopausal status. *Jpn.J Cancer Res* 1995;86:146-54.
221. Wu AH, Ziegler RG, Horn-Ross PL, Nomura AM, West DW, Kolonel LN *et al.* Tofu and risk of breast cancer in Asian-Americans. *Cancer Epidemiol.Biomarkers Prev.* 1996;5:901-06.
222. Dai Q, Shu XO, Jin F, Potter JD, Kushi LH, Teas J *et al.* Population-based case-control study of soyfood intake and breast cancer risk in Shanghai. *Br.J Cancer* 2001;85:372-78.
223. Wu AH, Wan P, Hankin J, Tseng CC, Yu MC, Pike MC. Adolescent and adult soy intake and risk of breast cancer in Asian-Americans. *Carcinogenesis* 2002;23:1491-96.

224. Yamamoto S, Sobue T, Kobayashi M, Sasaki S, Tsugane S. Soy, isoflavones, and breast cancer risk in Japan. *J Natl.Cancer Inst.* 2003;95:906-13.
225. Huntley AL, Ernst E. Soy for the treatment of perimenopausal symptoms--a systematic review. *Maturitas* 2004;47:1-9.
226. Crisafulli A, Marini H, Bitto A, Altavilla D, Squadrito G, Romeo A *et al.* Effects of genistein on hot flushes in early postmenopausal women: a randomized, double-blind EPT- and placebo-controlled study. *Menopause.* 2004;11:400-04.
227. Shughrue PJ, Komm B, Merchenthaler I. The distribution of estrogen receptor-beta mRNA in the rat hypothalamus. *Steroids* 1996;61:678-81.
228. Kruijver FP, Balesar R, Espila AM, Unmehopa UA, Swaab DF. Estrogen-receptor-beta distribution in the human hypothalamus: similarities and differences with ER alpha distribution. *J Comp Neurol.* 2003;466:251-77.
229. Forsling ML, Kallo I, Hartley DE, Heinze L, Ladek R, Coen CW *et al.* Oestrogen receptor-beta and neurohypophysial hormones: functional interaction and neuroanatomical localisation. *Pharmacol Biochem Behav.* 2003;76:535-42.
230. Silbiger SR, Neugarten J. The impact of gender on the progression of chronic renal disease. *Am J Kidney Dis.* 1995;25:515-33.
231. Neugarten J, Acharya A, Silbiger SR. Effect of gender on the progression of nondiabetic renal disease: a meta-analysis. *J Am Soc.Nephrol.* 2000;11:319-29.
232. Potier M, Karl M, Zheng F, Elliot SJ, Striker GE, Striker LJ. Estrogen-related abnormalities in glomerulosclerosis-prone mice: reduced mesangial cell estrogen receptor expression and prosclerotic response to estrogens. *Am J Pathol.* 2002;160:1877-85.
233. Neugarten J, Gallo G, Silbiger S, Kasiske B. Glomerulosclerosis in aging humans is not influenced by gender. *Am J Kidney Dis.* 1999;34:884-88.
234. Sakemi T, Toyoshima H, Shouno Y, Morito F. Estrogen attenuates progressive glomerular injury in hypercholesterolemic male Imai rats. *Nephron* 1995;69:159-65.
235. Sakemi T, Ohtsuka N, Tomiyoshi Y, Morito F. Testosterone does not eliminate the attenuating effect of estrogen on progressive glomerular injury in estrogen-treated hypercholesterolemic male Imai rats. *Kidney Blood Press Res* 1997;20:51-56.
236. Xiao S, Gillespie DG, Baylis C, Jackson EK, Dubey RK. Effects of estradiol and its metabolites on glomerular endothelial nitric oxide synthesis and mesangial cell growth. *Hypertension* 2001;37:645-50.

237. Elliot SJ, Karl M, Berho M, Potier M, Zheng F, Leclercq B *et al.* Estrogen deficiency accelerates progression of glomerulosclerosis in susceptible mice. *Am J Pathol.* 2003;162:1441-48.
238. Antus B, Hamar P, Kokeny G, Szollosi Z, Mucsi I, Nemes Z *et al.* Estradiol is nephroprotective in the rat remnant kidney. *Nephrol.Dial.Transplant.* 2003;18:54-61.
239. Gross ML, Adamczak M, Rabe T, Harbi NA, Krtil J, Koch A *et al.* Beneficial Effects of Estrogens on Indices of Renal Damage in Uninephrectomized SHRsp Rats. *J Am Soc.Nephrol.* 2004;15:348-58.
240. Maric C, Sandberg K, Hinojosa-Laborde C. Glomerulosclerosis and tubulointerstitial fibrosis are attenuated with 17beta-estradiol in the aging Dahl salt sensitive rat. *J Am Soc.Nephrol.* 2004;15:1546-56.
241. Sakemi T, Ohtsuka N, Shouno Y, Morito F. Effect of ovariectomy on glomerular injury in hypercholesterolemic female Imai rats. *Nephron* 1996;72:72-78.
242. Joles JA, van Goor H, Koomans HA. Estrogen induces glomerulosclerosis in analbuminemic rats. *Kidney Int* 1998;53:862-68.
243. Stevenson FT, Wheeldon CM, Gades MD, Kaysen GA, Stern JS, van Goor H. Estrogen worsens incipient hypertriglyceridemic glomerular injury in the obese Zucker rat. *Kidney Int* 2000;57:1927-35.
244. Tomiyoshi Y, Sakemi T, Aoki S, Miyazono M. Different effects of castration and estrogen administration on glomerular injury in spontaneously hyperglycemic Otsuka Long-Evans Tokushima Fatty (OLETF) rats. *Nephron* 2002;92:860-67.
245. Sakemi T, Ohtsuka N, Shouno Y, Morito F. Ovariectomy attenuates proteinuria and glomerular injury in unilaterally nephrectomized female Sprague-Dawley rats. *Nephron* 1996;73:251-57.
246. Ohtsuka N, Sakemi T, Tomiyoshi Y, Morito F. Different effect of estrogen administration from castration on glomerular injury in unilaterally nephrectomized male Sprague-Dawley rats. *Nephron* 1997;77:445-51.
247. Ikeda Y, Sakemi T, Tomiyoshi Y, Miyazono M. Combined therapy with estrogen and testosterone eliminates the aggravating effect of estrogen replacement therapy on glomerular injury in hypercholesterolemic female Imai rats. *Kidney Blood Press Res* 2000;23:27-34.
248. Studer RK, Craven PA, DeRubertis FR. Antioxidant inhibition of protein kinase C-signaled increases in transforming growth factor-beta in mesangial cells. *Metabolism* 1997;46:918-25.

249. Toyoda M, Suzuki D, Honma M, Uehara G, Sakai T, Umezono T *et al.* High expression of PKC-MAPK pathway mRNAs correlates with glomerular lesions in human diabetic nephropathy. *Kidney Int* 2004;66:1107-14.
250. Weigert C, Sauer U, Brodbeck K, Pfeiffer A, Haring HU, Schleicher ED. AP-1 proteins mediate hyperglycemia-induced activation of the human TGF-beta1 promoter in mesangial cells. *J Am Soc.Nephrol.* 2000;11:2007-16.
251. Koya D, Jirousek MR, Lin YW, Ishii H, Kuboki K, King GL. Characterization of protein kinase C beta isoform activation on the gene expression of transforming growth factor-beta, extracellular matrix components, and prostanoids in the glomeruli of diabetic rats. *J Clin Invest* 1997;100:115-26.
252. Ishii H, Tada H, Isogai S. An aldose reductase inhibitor prevents glucose-induced increase in transforming growth factor-beta and protein kinase C activity in cultured mesangial cells. *Diabetologia* 1998;41:362-64.
253. Ikehara K, Tada H, Kuboki K, Inokuchi T. Role of protein kinase C-angiotensin II pathway for extracellular matrix production in cultured human mesangial cells exposed to high glucose levels. *Diabetes Res Clin Pract.* 2003;59:25-30.
254. Yasuda T, Kondo S, Homma T, Harris RC. Regulation of extracellular matrix by mechanical stress in rat glomerular mesangial cells. *J Clin Invest* 1996;98:1991-2000.
255. Hirakata M, Kaname S, Chung UG, Joki N, Hori Y, Noda M *et al.* Tyrosine kinase dependent expression of TGF-beta induced by stretch in mesangial cells. *Kidney Int* 1997;51:1028-36.
256. Weigert C, Brodbeck K, Klopfer K, Haring HU, Schleicher ED. Angiotensin II induces human TGF-beta 1 promoter activation: similarity to hyperglycaemia. *Diabetologia* 2002;45:890-98.
257. Chen S, Cohen MP, Lautenslager GT, Shearman CW, Ziyadeh FN. Glycated albumin stimulates TGF-beta 1 production and protein kinase C activity in glomerular endothelial cells. *Kidney Int* 2001;59:673-81.
258. Kim YS, Kim BC, Song CY, Hong HK, Moon KC, Lee HS. Advanced glycosylation end products stimulate collagen mRNA synthesis in mesangial cells mediated by protein kinase C and transforming growth factor-beta. *J Lab Clin Med* 2001;138:59-68.
259. Studer RK, Negrete H, Craven PA, DeRubertis FR. Protein kinase C signals thromboxane induced increases in fibronectin synthesis and TGF-beta bioactivity in mesangial cells. *Kidney Int* 1995;48:422-30.

260. Wu Z, Zhou Q, Lan Y, Wang Y, Xu X, Jin H. AP-1 complexes mediate oxidized LDL-induced overproduction of TGF-beta(1) in rat mesangial cells. *Cell Biochem Funct.* 2004;22:237-47.
261. Lee HS, Kim BC, Hong HK, Kim YS. LDL stimulates collagen mRNA synthesis in mesangial cells through induction of PKC and TGF-beta expression. *Am J Physiol* 1999;277:F369-F376.
262. Singh LP, Green K, Alexander M, Bassly S, Crook ED. Hexosamines and TGF-beta1 use similar signaling pathways to mediate matrix protein synthesis in mesangial cells. *Am J Physiol Renal Physiol* 2004;286:F409-F416.
263. Weiss RH, Ramirez A. TGF-beta- and angiotensin-II-induced mesangial matrix protein secretion is mediated by protein kinase C. *Nephrol.Dial.Transplant.* 1998;13:2804-13.
264. Tada H, Isogai S. The fibronectin production is increased by thrombospondin via activation of TGF-beta in cultured human mesangial cells. *Nephron* 1998;79:38-43.
265. Runyan CE, Schnaper HW, Poncelet AC. Smad3 and PKCdelta mediate TGF-beta1-induced collagen I expression in human mesangial cells. *Am J Physiol Renal Physiol* 2003;285:F413-F422.
266. Zdunek M, Silbiger S, Lei J, Neugarten J. Protein kinase CK2 mediates TGF-beta1-stimulated type IV collagen gene transcription and its reversal by estradiol. *Kidney Int* 2001;60:2097-108.
267. Singh R, Song RH, Alavi N, Pegoraro AA, Singh AK, Leehey DJ. High glucose decreases matrix metalloproteinase-2 activity in rat mesangial cells via transforming growth factor-beta1. *Exp Nephrol.* 2001;9:249-57.
268. Baricos WH, Cortez SL, Deboisblanc M, Xin S. Transforming growth factor-beta is a potent inhibitor of extracellular matrix degradation by cultured human mesangial cells. *J Am Soc.Nephrol.* 1999;10:790-95.
269. Poncelet AC, Schnaper HW. Regulation of human mesangial cell collagen expression by transforming growth factor-beta1. *Am J Physiol* 1998;275:F458-F466.
270. Marti HP, Lee L, Kashgarian M, Lovett DH. Transforming growth factor-beta 1 stimulates glomerular mesangial cell synthesis of the 72-kd type IV collagenase. *Am J Pathol.* 1994;144:82-94.
271. Xin C, Ren S, Kleuser B, Shabahang S, Eberhardt W, Radeke H *et al.* Sphingosine 1-phosphate cross-activates the Smad signaling cascade and mimics transforming growth factor-beta-induced cell responses. *J Biol Chem* 2004;279:35255-62.

272. Kaizuka M, Yamabe H, Osawa H, Okumura K, Fujimoto N. Thrombin stimulates synthesis of type IV collagen and tissue inhibitor of metalloproteinases-1 by cultured human mesangial cells. *J Am Soc.Nephrol.* 1999;10:1516-23.
273. Kopp JB, Factor VM, Mozes M, Nagy P, Sanderson N, Bottinger EP *et al.* Transgenic mice with increased plasma levels of TGF-beta 1 develop progressive renal disease. *Lab Invest* 1996;74:991-1003.
274. Ali SM, Laping NJ, Fredrickson TA, Contino LC, Olson B, Anderson K *et al.* Angiotensin-converting enzyme inhibition attenuates proteinuria and renal TGF-beta 1 mRNA expression in rats with chronic renal disease. *Pharmacology* 1998;57:20-27.
275. Sharma R, Khanna A, Sharma M, Savin VJ. Transforming growth factor-beta1 increases albumin permeability of isolated rat glomeruli via hydroxyl radicals. *Kidney Int* 2000;58:131-36.
276. Fujimoto M, Maezawa Y, Yokote K, Joh K, Kobayashi K, Kawamura H *et al.* Mice lacking Smad3 are protected against streptozotocin-induced diabetic glomerulopathy. *Biochem Biophys Res Commun.* 2003;305:1002-07.
277. Benigni A, Zoja C, Corna D, Zatelli C, Conti S, Campana M *et al.* Add-on anti-TGF-beta antibody to ACE inhibitor arrests progressive diabetic nephropathy in the rat. *J Am Soc.Nephrol.* 2003;14:1816-24.
278. Sharma K, Jin Y, Guo J, Ziyadeh FN. Neutralization of TGF-beta by anti-TGF-beta antibody attenuates kidney hypertrophy and the enhanced extracellular matrix gene expression in STZ-induced diabetic mice. *Diabetes* 1996;45:522-30.
279. Ziyadeh FN, Hoffman BB, Han DC, Iglesias-de la Cruz MC, Hong SW, Isono M *et al.* Long-term prevention of renal insufficiency, excess matrix gene expression, and glomerular mesangial matrix expansion by treatment with monoclonal antitransforming growth factor-beta antibody in db/db diabetic mice. *Proc.Natl.Acad.Sci U.S.A* 2000;97:8015-20.
280. Chen S, Iglesias-de la Cruz MC, Jim B, Hong SW, Isono M, Ziyadeh FN. Reversibility of established diabetic glomerulopathy by anti-TGF-beta antibodies in db/db mice. *Biochem Biophys Res Commun.* 2003;300:16-22.
281. Potier M, Elliot SJ, Tack I, Lenz O, Striker GE, Striker LJ *et al.* Expression and regulation of estrogen receptors in mesangial cells: influence on matrix metalloproteinase-9. *J Am Soc.Nephrol.* 2001;12:241-51.
282. Lei J, Silbiger S, Ziyadeh FN, Neugarten J. Serum-stimulated alpha 1 type IV collagen gene transcription is mediated by TGF-beta and inhibited by estradiol. *Am J Physiol* 1998;274:F252-F258.

283. Silbiger S, Lei J, Ziyadeh FN, Neugarten J. Estradiol reverses TGF-beta1-stimulated type IV collagen gene transcription in murine mesangial cells. *Am J Physiol* 1998;274:F1113-F1118.
284. Matsuda T, Yamamoto T, Muraguchi A, Saatcioglu F. Cross-talk between transforming growth factor-beta and estrogen receptor signaling through Smad3. *J Biol Chem* 2001;276:42908-14.
285. Derynck R, Zhang Y, Feng XH. Smads: transcriptional activators of TGF-beta responses. *Cell* 1998;95:737-40.
286. Craven PA, Studer RK, Felder J, Phillips S, DeRubertis FR. Nitric oxide inhibition of transforming growth factor-beta and collagen synthesis in mesangial cells. *Diabetes* 1997;46:671-81.
287. Williams AJ, Baker F, Walls J. Effect of varying quantity and quality of dietary protein intake in experimental renal disease in rats. *Nephron* 1987;46:83-90.
288. Williams AJ, Walls J. Metabolic consequences of differing protein diets in experimental renal disease. *Eur J Clin Invest* 1987;17:117-22.
289. Iwasaki K, Gleiser CA, Masoro EJ, McMahan CA, Seo EJ, Yu BP. The influence of dietary protein source on longevity and age-related disease processes of Fischer rats. *J Gerontol.* 1988;43:B5-12.
290. Aukema HM, Housini I. Dietary soy protein effects on disease and IGF-I in male and female Han:SPRD-cy rats. *Kidney Int* 2001;59:52-61.
291. Velasquez MT, Bhatena SJ. Dietary phytoestrogens: a possible role in renal disease protection. *Am J Kidney Dis.* 2001;37:1056-68.
292. Maddox DA, Alavi FK, Silbernack EM, Zawada ET. Protective effects of a soy diet in preventing obesity-linked renal disease. *Kidney Int* 2002;61:96-104.
293. Sakemi T, Ikeda Y, Shimazu K. Effect of soy protein added to casein diet on the development of glomerular injury in spontaneous hypercholesterolemic male Imai rats. *Am J Nephrol.* 2002;22:548-54.
294. Fair DE, Ogborn MR, Weiler HA, Bankovic-Calic N, Nitschmann EP, Fitzpatrick-Wong SC *et al.* Dietary soy protein attenuates renal disease progression after 1 and 3 weeks in Han:SPRD-cy weanling rats. *J Nutr* 2004;134:1504-07.
295. Trujillo J, Ramirez V, Perez J, Torre-Villalvazo I, Torres N, Tovar AR *et al.* Renal protection by soy diet in obese Zucker rats is associated with restoration of nitric oxide generation. *Am J Physiol Renal Physiol* 2004.

296. Azadbakht L, Shakerhosseini R, Atabak S, Jamshidian M, Mehrabi Y, Esmail-Zadeh A. Beneficiary effect of dietary soy protein on lowering plasma levels of lipid and improving kidney function in type II diabetes with nephropathy. *Eur.J Clin Nutr* 2003;57:1292-94.
297. Teixeira SR, Tappenden KA, Carson L, Jones R, Prabhudesai M, Marshall WP *et al*. Isolated soy protein consumption reduces urinary albumin excretion and improves the serum lipid profile in men with type 2 diabetes mellitus and nephropathy. *J Nutr* 2004;134:1874-80.
298. Brenner BM, Meyer TW, Hostetter TH. Dietary protein intake and the progressive nature of kidney disease: the role of hemodynamically mediated glomerular injury in the pathogenesis of progressive glomerular sclerosis in aging, renal ablation, and intrinsic renal disease. *N.Engl.J Med* 1982;307:652-59.
299. Klahr S, Buerkert J, Purkerson ML. Role of dietary factors in the progression of chronic renal disease. *Kidney Int* 1983;24:579-87.
300. Nakamura H, Takasawa M, Kashara S, Tsuda A, Momotsu T, Ito S *et al*. Effects of acute protein loads of different sources on renal function of patients with diabetic nephropathy. *Tohoku J Exp Med* 1989;159:153-62.
301. Kontessis P, Jones S, Dodds R, Trevisan R, Nosadini R, Fioretto P *et al*. Renal, metabolic and hormonal responses to ingestion of animal and vegetable proteins. *Kidney Int* 1990;38:136-44.
302. Anderson JW, Smith BM, Washnock CS. Cardiovascular and renal benefits of dry bean and soybean intake. *Am J Clin Nutr* 1999;70:464S-74S.
303. Fouque D, Laville M, Boissel JP, Chifflet R, Labeeuw M, Zech PY. Controlled low protein diets in chronic renal insufficiency: meta-analysis. *BMJ* 1992;304:216-20.
304. Pedrini MT, Levey AS, Lau J, Chalmers TC, Wang PH. The effect of dietary protein restriction on the progression of diabetic and nondiabetic renal diseases: a meta-analysis. *Ann.Intern.Med* 1996;124:627-32.
305. Jibani MM, Bloodworth LL, Foden E, Griffiths KD, Galpin OP. Predominantly vegetarian diet in patients with incipient and early clinical diabetic nephropathy: effects on albumin excretion rate and nutritional status. *Diabet.Med* 1991;8:949-53.
306. D'Amico G, Gentile MG. Influence of diet on lipid abnormalities in human renal disease. *Am J Kidney Dis.* 1993;22:151-57.
307. Barsotti G, Morelli E, Cupisti A, Meola M, Dani L, Giovannetti S. A low-nitrogen low-phosphorus Vegan diet for patients with chronic renal failure. *Nephron* 1996;74:390-94.

308. Soroka N, Silverberg DS, Gremland M, Birk Y, Blum M, Peer G *et al.* Comparison of a vegetable-based (soya) and an animal-based low-protein diet in predialysis chronic renal failure patients. *Nephron* 1998;79:173-80.
309. Ranich T, Bhathena SJ, Velasquez MT. Protective effects of dietary phytoestrogens in chronic renal disease. *J Ren Nutr* 2001;11:183-93.
310. Sakemi T, Ikeda Y, Shimazu K, Uesugi T. Attenuating effect of a semipurified alcohol extract of soy protein on glomerular injury in spontaneous hypercholesterolemic male Imai rats. *Am J Kidney Dis.* 2001;37:832-37.
311. Neugarten J, Acharya A, Lei J, Silbiger S. Selective estrogen receptor modulators suppress mesangial cell collagen synthesis. *Am J Physiol Renal Physiol* 2000;279:F309-F318.
312. Slater M, Brown D, Husband A. In the prostatic epithelium, dietary isoflavones from red clover significantly increase estrogen receptor beta and E-cadherin expression but decrease transforming growth factor beta1. *Prostate Cancer Prostatic Dis.* 2002;5:16-21.
313. McCarty MF. Adjuvant strategies for prevention of glomerulosclerosis. *Med.Hypotheses* 2004;submitted for publication.
314. Higuchi Y, Otsu K, Nishida K, Hirotani S, Nakayama H, Yamaguchi O *et al.* The small GTP-binding protein Rac1 induces cardiac myocyte hypertrophy through the activation of apoptosis signal-regulating kinase 1 and nuclear factor-kappa B. *J Biol Chem* 2003;278:20770-77.
315. Simko F, Simko J. The potential role of nitric oxide in the hypertrophic growth of the left ventricle. *Physiol Res* 2000;49:37-46.
316. Lim WK, Wren B, Jepson N, Roy S, Caplan G. Effect of hormone replacement therapy on left ventricular hypertrophy. *Am J Cardiol.* 1999;83:1132-4, A9.
317. du CG, Ribstein J, Pasquie JL, Mimran A. [Determinant of left ventricular hypertrophy in the hypertensive woman. Influence of hormone replacement therapy for menopause]. *Arch Mal Coeur Vaiss.* 1999;92:975-77.
318. Modena MG, Molinari R, Muia N, Jr., Castelli A, Pala F, Rossi R. Double-blind randomized placebo-controlled study of transdermal estrogen replacement therapy on hypertensive postmenopausal women. *Am J Hypertens.* 1999;12:1000-08.
319. Modena MG, Muia N, Jr., Aveta P, Molinari R, Rossi R. Effects of transdermal 17beta-estradiol on left ventricular anatomy and performance in hypertensive women. *Hypertension* 1999;34:1041-46.
320. Light KC, Hinderliter AL, West SG, Grewen KM, Steege JF, Sherwood A *et al.* Hormone replacement improves hemodynamic profile and left ventricular

- geometry in hypertensive and normotensive postmenopausal women. *J Hypertens.* 2001;19:269-78.
321. Hayward CS, Webb CM, Collins P. Effect of sex hormones on cardiac mass. *Lancet* 2001;357:1354-56.
322. Miya Y, Sumino H, Ichikawa S, Nakamura T, Kanda T, Kumakura H *et al.* Effects of hormone replacement therapy on left ventricular hypertrophy and growth-promoting factors in hypertensive postmenopausal women. *Hypertens.Res* 2002;25:153-59.
323. Agabiti-Rosei E, Muiesan ML. Left ventricular hypertrophy and heart failure in women. *J Hypertens.* 2002;20 Suppl 2:S34-S38.
324. Grohe C, Kahlert S, Lobbert K, Stimpel M, Karas RH, Vetter H *et al.* Cardiac myocytes and fibroblasts contain functional estrogen receptors. *FEBS Lett.* 1997;416:107-12.
325. Nuedling S, Karas RH, Mendelsohn ME, Katzenellenbogen JA, Katzenellenbogen BS, Meyer R *et al.* Activation of estrogen receptor beta is a prerequisite for estrogen-dependent upregulation of nitric oxide synthases in neonatal rat cardiac myocytes. *FEBS Lett.* 2001;502:103-08.
326. Skavdahl M, Steenbergen C, Clark J, Myers P, Demianenko T, Mao L *et al.* The Beta Estrogen Receptor Mediates Male-Female Differences in the Development of Pressure Overload Hypertrophy. *Am J Physiol Heart Circ.Physiol* 2004.
327. Picherit C, Coxam V, Bennetau-Pelissero C, Kati-Coulibaly S, Davicco MJ, Lebecque P *et al.* Daidzein is more efficient than genistein in preventing ovariectomy-induced bone loss in rats. *J Nutr* 2000;130:1675-81.
328. Teede HJ, McGrath BP, DeSilva L, Cehun M, Fassoulakis A, Nestel PJ. Isoflavones reduce arterial stiffness: a placebo-controlled study in men and postmenopausal women. *Arterioscler.Thromb.Vasc.Biol* 2003;23:1066-71.
329. Fonseca D, Ward WE. Daidzein together with high calcium preserve bone mass and biomechanical strength at multiple sites in ovariectomized mice. *Bone* 2004;35:489-97.
330. Woodman OL, Missen MA, Boujaoude M. Daidzein and 17 beta-estradiol enhance nitric oxide synthase activity associated with an increase in calmodulin and a decrease in caveolin-1. *J Cardiovasc.Pharmacol* 2004;44:155-63.
331. Setchell KD, Brown NM, Lydeking-Olsen E. The clinical importance of the metabolite equol-a clue to the effectiveness of soy and its isoflavones. *J Nutr* 2002;132:3577-84.

332. Muthyala RS, Ju YH, Sheng S, Williams LD, Doerge DR, Katzenellenbogen BS *et al.* Equol, a natural estrogenic metabolite from soy isoflavones: convenient preparation and resolution of R- and S-equols and their differing binding and biological activity through estrogen receptors alpha and beta. *Bioorg.Med Chem* 2004;12:1559-67.
333. Rowland IR, Wiseman H, Sanders TA, Adlercreutz H, Bowey EA. Interindividual variation in metabolism of soy isoflavones and lignans: influence of habitual diet on equol production by the gut microflora. *Nutr Cancer* 2000;36:27-32.
334. Armstrong B, Doll R. Environmental factors and cancer incidence and mortality in different countries, with special reference to dietary practices. *Int J Cancer* 1975;15:617-31.
335. Hebert JR, Hurley TG, Olendzki BC, Teas J, Ma Y, Hampl JS. Nutritional and socioeconomic factors in relation to prostate cancer mortality: a cross-national study [see comments]. *J Natl Cancer Inst* 1998;90:1637-47.
336. Hebert JR, Rosen A. Nutritional, socioeconomic, and reproductive factors in relation to female breast cancer mortality: findings from a cross-national study. *Cancer Detect Prev* 1996;20:234-44.
337. McCarty MF. Mortality from Western cancers rose dramatically among African-Americans during the 20th century: are dietary animal products to blame? *Med Hypotheses* 2001;57:169-74.
338. Giovannucci E. Nutrition, insulin, insulin-like growth factors and cancer. *Horm.Metab Res* 2003;35:694-704.
339. McCarty MF. Insulin and IGF-I as determinants of low 'Western' cancer rates in the rural third world. *Int J Epidemiol.* 2004;33:908-10.
340. Khalil DA, Lucas EA, Juma S, Smith BJ, Payton ME, Arjmandi BH. Soy protein supplementation increases serum insulin-like growth factor-I in young and old men but does not affect markers of bone metabolism. *J Nutr* 2002;132:2605-08.
341. Allen NE, Appleby PN, Davey GK, Kaaks R, Rinaldi S, Key TJ. The associations of diet with serum insulin-like growth factor I and its main binding proteins in 292 women meat-eaters, vegetarians, and vegans. *Cancer Epidemiol.Biomarkers Prev.* 2002;11:1441-48.
342. Arjmandi BH, Khalil DA, Smith BJ, Lucas EA, Juma S, Payton ME *et al.* Soy protein has a greater effect on bone in postmenopausal women not on hormone replacement therapy, as evidenced by reducing bone resorption and urinary calcium excretion. *J Clin Endocrinol.Metab* 2003;88:1048-54.

343. Schulsinger DA, Root MM, Campbell TC. Effect of dietary protein quality on development of aflatoxin B1-induced hepatic preneoplastic lesions. *J Natl Cancer Inst* 1989;81:1241-45.
344. Campbell TC, Junshi C. Diet and chronic degenerative diseases: perspectives from China. *Am J Clin Nutr* 1994;59:1153S-61S.