

Minimizing the Cancer-Promotional Activity of Cox-2 as a Central Strategy in Cancer Prevention

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Abstract

A recent meta-analysis examining long-term mortality in subjects who participated in controlled studies evaluating the impact of daily aspirin on vascular risk, has concluded that aspirin confers substantial protection from cancer mortality. Remarkably, low-dose aspirin was as effective as higher-dose regimens; hence this protection may be achievable with minimal risk. There is reason to believe that this protection stems primarily from inhibition of cox-2 in pre-neoplastic lesions. Since safe aspirin regimens can only achieve a partial and transitory inhibition of cox-2, it may be feasible to complement the cancer-protective benefit of aspirin with other measures which decrease cox-2 expression or which limit the bioactivity of cox-2-derived PGE2. Oxidative stress boosts cox-2 expression by up-regulating activation of NF-kappaB and MAP kinases; NADPH oxidase activation may thus promote carcinogenesis by increasing cox-2 expression while also amplifying oxidant-mediated mutagenesis. A prospective cohort study has observed that relatively elevated serum bilirubin levels are associated with a marked reduction in subsequent cancer mortality; this may reflect bilirubin's physiological role as a potent inhibitor of NADPH oxidase. It may be feasible to mimic this protective effect by supplementing with spirulina, a rich source of a phycobilin which shares bilirubin's ability to inhibit NADPH oxidase. Ancillary antioxidant measures – phase 2 inducing phytochemicals, melatonin, N-acetylcysteine, and astaxanthin – may also aid cox-2 down-regulation. The cancer protection often associated with high-normal vitamin D status may be attributable, in part, to the ability of the activated vitamin D receptor to decrease cox-2 expression while promoting PGE2 catabolism and suppressing the expression of PGE2 receptors. Diets with a relatively low ratio of omega-6 to long-chain omega-3 fats may achieve cancer protection by antagonizing the production and bioactivity of PGE2. Growth factors such as IGF-I increase cox-2 expression by several complementary mechanisms; hence, decreased cox-2 activity may play a role in the remarkably low mortality from “Western” cancers enjoyed by Third World cultures in which systemic growth factor activity was minimized by quasi-vegan diets complemented by leanness and excellent muscle insulin sensitivity. Practical strategies for achieving a modest degree of calorie restriction may also have potential for down-regulating cox-2 expression while decreasing cancer risk. Soy isoflavones, linked to reduced cancer risk in Asian epidemiology, may suppress cox-2 induction by activating ERbeta. In aggregate, these considerations suggest that a comprehensive lifestyle strategy targeting cox-2 expression and bioactivity may have tremendous potential for cancer prevention.

Cancer Preventive Potential of Low-Dose Aspirin

Recent meta-analyses following up long-term health outcomes in subjects who participated in controlled trials of daily aspirin use for prevention of vascular events, have revealed that daily aspirin administration is associated with a marked reduction in cancer mortality; the largest impact was seen with adenocarcinomas, most notably those of gastrointestinal origin.^{1, 2} At 20 years of follow-up, mortality from all solid cancers was 20% lower in those receiving aspirin; risk of death from adenocarcinomas was 34% lower. These findings are statistically robust, and are particularly astounding in light of the fact that these were intention-to-treat analyses (implying that subjects randomized to aspirin who were poorly compliant were nonetheless included as aspirin users), and a substantial proportion of subjects randomized to aspirin can be presumed to have discontinued aspirin use after the formal completion of their trials, which were not longer than 9 years. (It can also be presumed that some subjects randomized to placebo commenced aspirin use after trial completion, further reducing the power of the study to demonstrate benefit.) When analysis was restricted to subjects with duration of trial treatment over 7.5 years, the preventive effects were even more dramatic: deaths from all solid tumors were 31% lower, and deaths from gastrointestinal cancers were 59% lower. This suggests that continual use of daily aspirin throughout adulthood might well have an astoundingly positive impact on cancer mortality. Moreover, the meta-analyses concluded that a 75 mg daily dose was as protective as higher doses; hence, except in those individuals for which aspirin is outright contraindicated, marked protection could be achieved at a dose that would entail minimal risk for bleeding complications or other side effects. This is surely one of the most encouraging medical discoveries of recent decades.

It is probably reasonable to assume that aspirin's utility in this regard stems primarily from its ability to suppress production of cancer-promoting prostanoids by cox-2 in pre-cancerous lesions; thus, aspirin use has shown no impact on risk for colon adenocarcinomas that do not overexpress cox-2, or on risk for mortality in patients whose colon adenocarcinomas do not overexpress this enzyme.^{3, 4} The cancer-promoting activity of cox-2-derived prostanoids, most notably PGE2, is thought to reflect autocrine or paracrine stimulation of prostaglandin receptors (the four subtypes EP1-EP4), which in many tissues inhibit apoptosis via cell-type-specific mechanisms such as increased Bcl-2 expression, decreased Bax or Bim expression, Bad phosphorylation, and Akt activation; stimulation of these receptors also often promotes proliferation, invasiveness, and angiogenesis, and inhibits the cancer-scavenging activity of cytotoxic T cells or NK cells.⁵⁻¹⁵ The magnitude of protection observed in the recent meta-analyses is particularly remarkable in light of the fact that low-dose aspirin could only be expected to achieve partial and temporary inhibition of cox-2; since aspirin somewhat preferentially inhibits cox-1,¹⁶ an aspirin regimen which achieved substantial sustained inhibition of cox-2 could be expected to suppress cox-1 activity at least as strongly, and hence would be associated with high risk for gastric ulceration and kidney damage. Indeed, low-dose aspirin appears to be notably more effective for preventing adenocarcinomas of the proximal colon than those of the distal colon; Rothwell and colleagues note that, inasmuch as cox-2 expression tends to be higher in distal lesions, low-dose aspirin may have achieved insufficient inhibition of cox-2 activity in distal pre-cancerous lesions to provide optimal protection.¹

Regulation of Cox-2 Expression

This latter point suggests the possibility that adjuvant measures which suppress the expression of either cox-2 or of prostaglandin receptors, or which limit the availability of arachidonic acid as a substrate for PGE2 synthesis, might complement the utility of low-dose aspirin for cancer prevention, enabling even greater protection to be achieved. Indeed, an examination of other measures suspected to be useful for cancer prevention reveals that many of them could be expected to decrease the cancer promotional activity of cox-2 in pre-cancerous lesions. Such measures include effective antioxidant strategies – of which spirulina-derived phycocyanobilin may have the most intriguing potential – as well as vitamin D, a diet with a low ratio of omega-6 fatty acids to long-chain omega-3 fatty acids, and dietary/lifestyle measures which could be expected to minimize systemic levels of insulin and free IGF-I.

Analysis of the promoter region of the human cox-2 gene reveals binding sites for NF-kappaB, C/EBP, AP-2, SP1, Ets-1, and a CRE motif.¹⁷⁻¹⁹ Studies in various types of cells indicates that NF-kappaB can be a potent mediator of cox-2 transcription, and that the various MAP kinases – Erk1/2, JNK, and p38 – can also promote this transcription by boosting the activity of transcription factors and coactivators which target the cox-2 promoter.¹⁹⁻²⁶ It is well known that oxidative stress and various pro-inflammatory cytokines can promote activation of NF-kappaB and of the MAP kinases. Growth factors such as IGF-I or EGF also boost the expression of cox-2 mRNA; this may reflect stimulation of transcription via Erk1/2, as well as an increase in the half-life of cox-2 mRNA mediated by the PI3K-Akt pathway stimulated by most growth factors.^{23, 27-30}

Bilirubin and Spirulina – Potential for Down-Regulating Cox-2 Expression

A number of studies show that oxidative stress can enhance cox-2 expression in many types of cells, acting via NF-kappaB and the MAP kinases.^{25, 26, 31-38} NADPH oxidase complexes are a key source of oxidative stress, particularly in cells exposed to pro-inflammatory cytokines, and inhibition of NADPH oxidase has been shown to blunt cox-2 induction in many cell culture studies.^{32-37, 39-44} The extent to which NADPH oxidase activation contributes to inflammatory carcinogenesis in humans, whether by cox-2 induction or other mechanisms, remains unclear. However, a very intriguing clue has emerged from epidemiology focusing on the physiological antioxidant bilirubin.

Whereas albumin-bound bilirubin functions as an important scavenging antioxidant in plasma, the intracellular concentration of free bilirubin is so low (low nanomolar range) that the important intracellular antioxidant activity of bilirubin cannot be credibly attributed to non-specific scavenging activity.⁴⁵ Recent studies reveal that, in intracellular concentrations achieved by heme-oxygenase-1 induction, bilirubin functions as a very potent inhibitor of NADPH oxidase complexes; the isoform specificity of this effect still requires clarification.⁴⁶⁻⁴⁹ This discovery provides a satisfying rationale for recent epidemiology correlating increased serum bilirubin levels with reduced risk for vascular disorders, diabetes, and diabetic complications.⁵⁰⁻⁵⁶

Only one large prospective study to-date has examined the association between serum bilirubin levels and total cancer mortality. The Belgian Inter-University Research on Nutrition and Health study measured serum at baseline and correlated this with all-cause, cardiovascular, and cancer mortality during a 10-year follow-up in over 10,000 subjects of both sexes.⁵⁷ After adjustment for various covariants, subjects with

baseline bilirubin > 0.6 mg/dl, as contrasted with those with baseline bilirubin <0.2 mg/dl, were 58% less likely to have died of cancer over the next 10 years (RR=0.42; 95% CI 0.26-0.68). Cancer mortality in the intervening bilirubin strata was intermediate, and the inverse association of bilirubin level and cancer mortality had high statistical significance (p for trend = 0.004).

More recently, the association of baseline bilirubin with lung cancer incidence was evaluated in a huge British prospective cohort study (over half a million subjects, followed up over an average of 8 years).⁵⁸ Risk for lung cancer was found to decline monotonically with increasing bilirubin levels. Male risk for lung cancer in the tenth decile of bilirubin was 70% lower than that in the first decile, and 50% lower than that in the fifth decile; the findings in women were comparable. These findings are confounded somewhat by the fact that bilirubin levels tend to be slightly lower in current smokers, possibly owing to the oxidant effect of tobacco smoke. Nonetheless, a marked inverse association of bilirubin and lung cancer risk persisted after multivariate adjustments, including smoking status. These findings are highly consistent with a trend toward markedly lower lung cancer mortality in subjects with higher bilirubin status noted in the Belgian study, which presumably missed statistical significance owing to the modest number of cases involved.

Few other epidemiological studies to-date have examined the association between bilirubin and cancer risk. Two such studies have focused on colorectal cancer; a study employing data from the Third National Health and Nutrition Examination Survey found a large protective effect with increasing bilirubin; a 1-mg/dl increase in bilirubin was associated with a relative risk of 0.257 (95% CI 0.254-0.260).⁵⁹ Perplexingly, no such protection was observed when other researchers analyzed data from the First National Health and Nutrition Examination Survey.⁶⁰ The latter study has been criticized for failing to exclude subjects with elevated liver enzymes at baseline (in whom high bilirubin may have been a marker for liver disease), but the authors indicate that their conclusion was not altered when they did such an adjustment.^{61, 62} Another pertinent study examined the association between common polymorphisms of the enzyme UGT1A1, responsible for conjugating bilirubin to glucuronic acid in the liver; reduced hepatic expression of this enzyme leads to increased circulating levels of free bilirubin. The analysis revealed that risk of head and neck cancer was significantly higher in subjects expressing the common high-expression allele of this gene.⁶³

The possible impact of bilirubin on cancer risk can also be assessed by examining the role of heme oxygenase-1 (HO-1) in this regard. This gene likewise is polymorphic; an increase in the number of GT dinucleotide repeats in its promoter region is associated with less efficient expression in response to oxidative stress.⁶⁴ Oxidant-induced expression of HO-1 would presumably often help to quell this oxidative stress via generation of bilirubin.⁴⁹ However, HO-1 induction can be a two-edged sword in this regard, as it also releases free iron from catabolized heme. Subjects carrying decreased expression alleles of this gene have been reported to be at increased risk for esophageal squamous cell carcinoma, lung adenocarcinoma, postmenopausal breast cancer, and oral squamous cell carcinoma;⁶⁵⁻⁶⁸ however, gastric cancer and melanoma were found to be more common in subjects carrying these alleles.^{69, 70}

Clearly, more epidemiological research correlating bilirubin levels and polymorphisms of UGT1A1 and HO-1 with cancer risk are warranted. But the Belgian and British prospective studies provide strong reason to suspect that high-normal bilirubin levels may provide important protection from cancer. A

suppression of cox-2 induction in tissues exposed to inflammatory stimuli likely plays a key role in this effect. However, additional complementary mechanisms are likely at work. A moderate elevation of oxidative stress, produced by constitutive activation of NADPH oxidase, is observed in many cancers, and acts to promote survival and aggressiveness by up-regulating growth factor activities (via reversible inhibition of tyrosine phosphatases) and activation of NF-kappaB (which can oppose apoptosis by a variety of mechanism in addition to cox-2 induction).⁷¹⁻⁷³ Not unlikely, NADPH oxidase plays an analogous role in many pre-cancerous lesions, thereby aiding cancer promotion.^{74, 75} Moreover, oxidants – notably peroxynitrite, and hydrogen peroxide in the presence of labile iron, can induce mutagenic damage.⁷⁶⁻⁷⁹ Urinary 8-hydroxydeoxyguanosine is a marker for such damage, and the fact that serum ferritin levels tend to correlate directly, and transferrin receptor levels correlate inversely, with this marker in humans, suggests that iron-catalyzed oxidant damage to DNA makes an important contribution to spontaneous mutagenesis in humans.⁸⁰⁻⁸² Consistent with this possibility, a recent controlled study evaluating phlebotomy-induced iron depletion observed a significant reduction of cancer incidence in the iron-depleted group.⁸³ Hence, to the extent that elevated bilirubin levels can suppress the production of hydrogen peroxide and peroxynitrite in inflamed tissues, it may help to prevent inflammatory mutagenesis. (Keeping body iron stores relatively low should also be helpful in this regard. Diets low in heme iron – such as the vegan diet recommended below – tend to moderate body iron stores.)^{84, 85}

These findings are of much more than theoretical interest, in light of recent evidence that oral administration of biliverdin can exert profound antioxidant effects in rodents that presumably are mediated by intracellular generation of bilirubin.⁸⁶ Moreover, whereas biliverdin is expensive to synthesize and is not readily available, the biliverdin derivative and homolog phycocyanobilin (PhyCB) constitutes about 0.6% by dry weight of the microalga spirulina. PhyCB is rapidly converted within cells to phycocyanorubin, a bilirubin homolog that appears to share its ability to inhibit NADPH oxidase.^{87, 88} This likely explains why oral administration of spirulina (or of phycocyanin, the spirulina protein which contains PhyCB as a chromophore) has been associated with profound and versatile anti-inflammatory and cytoprotective activity in rodent studies.⁸⁸⁻⁹¹ Hence, sufficient intakes of spirulina, or of PhyCB-enriched spirulina extracts, may have exciting practical potential for cancer prevention and for prevention and control of a range of other oxidant-driven pathologies.⁸⁸ The impact of spirulina/PhyCB on inflammatory carcinogenesis in rodents should receive further evaluation; inhibition of oral cancer and hepatic cancer with spirulina in rodents has already been reported.^{92, 93}

Complementary Antioxidant Measures for Controlling Cox-2

Of course, PhyCB is not the only nutraceutical antioxidant which might have potential for suppressing cox-2 induction or oxidative mutagenesis. In particular, phase 2 inducers such as isothiocyanates, curcumin, epigallocatechin-gallate, and lipoic acid have been shown to decrease the expression of cox-2 in cell culture or rodent studies.⁹⁴⁻⁹⁸ This may reflect induction of HO-1, but increased expression of other antioxidant enzymes and of glutathione could also be expected to blunt the impact of hydrogen peroxide on cell signaling as well as on inflammatory mutagenesis. Phase 2 inducers would also act to diminish cancer risk by promoting the conjugation and excretion of mutagenic electrophiles (i.e. activated carcinogens).^{99, 100} Nocturnal melatonin administration has similar inductive effects that might prove comparably protective;¹⁰¹ indeed, melatonin can blunt cox-2 induction in vitro, and has shown protective activity in rodent models of carcinogenesis.¹⁰²⁻¹⁰⁷ Melatonin complements the efficacy of cox-2 inhibitors

in some of these studies. Moreover, a derangement of physiological melatonin release may contribute to the increased cancer risk observed in night-shift workers.¹⁰⁸⁻¹¹² And N-acetylcysteine has the potential to complement the benefit of these inductive agents by potentiating their impact on cellular glutathione availability.¹¹³⁻¹¹⁸ Ascorbic acid, like glutathione, is a key intracellular oxidant scavenger; in populations with poor baseline ascorbate status (in whom increased ascorbate intake can raise intracellular ascorbate levels¹¹⁹), vitamin C supplementation or an increase in consumption of ascorbate-rich fruits and vegetables may be protective.

The carotenoid astaxanthin is emerging as the most effective phytonutrient antioxidant for biomembranes, with considerably greater potential than vitamin E in this regard.¹²⁰ By protecting mitochondrial membranes from oxidative damage, this nutrient has the potential to moderate the contribution of stressed mitochondria to oxidant load.¹²¹⁻¹²³ Supplemental astaxanthin has been shown to suppress induction of cox-2 in the colon or rats treated with the carcinogen DMH, an effect associated with increased apoptosis in colonic mucosa.¹²⁴ In cell culture studies, it has blunted induction of cox-2 triggered by lipopolysaccharide or hyperglycemia.^{125, 126} Moreover, in one of the first human supplementation studies with this nutrient, a reduction in a biomarker of DNA damage was observed.¹²⁷ Hence, astaxanthin may have worthwhile potential for cancer prevention and for prevention and control of other oxidant-driven disorders.¹²⁰

Unfortunately, recent U.S. clinical studies focusing on prostate cancer risk have failed to realize hopes that supplemental selenium could provide protection from cancer; this may demonstrate that bolus carcinogen administration in rodents is a poor model for spontaneous carcinogenesis in humans.¹²⁸ However, baseline selenium nutrition in most Americans is relatively good, such that supplemental selenium has little impact on expression of glutathione peroxidase or other selenium-dependent antioxidant enzymes; selenium may well have greater potential for cancer prevention in populations where baseline selenium nutrition is poor owing to low selenium soil levels.^{129, 130}

High-dose folate has intriguing antioxidant potential, owing to the fact that many tissues concentrate it against a gradient and reduce it to tetrahydrofolates which have versatile antioxidant activity.¹³¹⁻¹³³ In particular, tetrahydrofolates quench peroxynitrite-derived radicals, likely mediators of inflammatory mutagenesis; hence, the impact of high-dose folate on DNA integrity in oxidatively-stressed tissues merits attention. It is more doubtful whether folate could influence cox-2 expression. Folate status can also influence DNA repair, cell proliferation, and DNA methylation; the net impact of nutritional-dose folate supplementation on cancer risk is a matter of ongoing controversy, as available data are inconsistent. However, there is some reason to suspect that good folate status will counteract the increase in risk for breast and colon cancer associated with moderate alcohol consumption.^{134, 135}

Squamous cell carcinomas – for which low-dose aspirin does not appear to be notably protective – tend to particularly common in disadvantaged populations consuming diets low in antioxidants such as vitamin C and flavonoids, and exposed to stressors such as tobacco smoke, alcohol, or betel nut that can induce oxidative stress.¹³⁶⁻¹⁴³ Hence there is reason to suspect that increased oxidative stress may often play a mutational and promotional role in their induction. Consistent with this is evidence linking low-expression alleles of heme oxygenase-1 (one of whose key products is bilirubin) with increased risk for such cancers.^{65, 68} Effective antioxidant measures may therefore complement mini-dose aspirin for cancer

prevention in the sense that they may provide protection from certain cancers not preventable with aspirin. Squamous cell carcinoma of the esophagus is said to be responsible for about one-sixth of total worldwide cancer mortality.¹⁴⁴

Vitamin D Antagonizes Cox-2 Expression and Bioactivity

High-normal vitamin D status, usually reflecting adequate skin exposure to UV-rich sunlight, has been associated with decreased risk for various cancers in case-control and ecologic epidemiology.¹⁴⁵⁻¹⁴⁷ Evidence that vitamin D provides protection from colorectal cancer is becoming overwhelming, but there are suggestive data that vitamin D may also provide protection from cancers of the breast, bladder, esophagus, gallbladder, stomach, ovary, renal, and the uterine endometrium.^{148, 149} An ecologic analysis concluded that if all Americans achieved optimal exposure to UV light (and hence high vitamin D status), over 23,000 Americans annually would avoid premature death from cancer;¹⁵⁰ however, some prospective studies examining serum levels of 25-hydroxycholecalciferol have yielded more equivocal results.¹⁵¹ Few placebo-controlled supplementation trials with vitamin D providing doses sufficiently high to reproduce the impact of optimal UV exposure have been done to date; however, one trial targeting postmenopausal women compared vitamin D (1100 IU) + calcium (1500 mg), calcium alone, and placebo over a 4 year period. When cancers that occurred over the first year were excluded (on the presumption that many of these were latently present when the trial began), relative risk for new non-skin cancer incidence in the vitamin D + calcium group was 0.232 (CI 0.09-0.60; $p < 0.005$), whereas a trend toward reduced cancer in the calcium alone group did not achieve statistical significance. (A significant reduction in cancer risk was also seen in the vitamin D + calcium group when the first year cancers were included in the analysis: RR=0.402 (P=0.01).) This conclusion was however based on a total of only 50 cancer cases; results from large multi-center studies should be awaited eagerly.

These observations may reflect the fact that a high proportion of the epithelia which commonly give rise to cancer express 1-alpha-hydroxylase activity, and therefore are capable of autocrine production of calcitriol, the active hormone derived from circulating 25-hydroxycholecalciferol.^{152, 153} Via interaction with vitamin D receptors – likewise commonly expressed in these tissues – calcitriol modulates gene expression in ways that tend to oppose carcinogenesis.

In particular, studies focusing on malignant or non-malignant breast or prostate cells, as well as colonic epithelium *in vivo*, have demonstrated that calcitriol, or less calcemic analogs thereof, can suppress cox-2 induction.¹⁵⁴⁻¹⁵⁷ The basis of this effect is not entirely clear, although some studies have shown that calcitriol boosts the expression of MAP kinase phosphatase 5, which opposes the activation of MAP kinases, notably p38.¹⁵⁸ But calcitriol also has the complementary ancillary effects of inducing the expression of 15-prostaglandin dehydrogenase – responsible for PGE2 catabolism - and suppressing the expression of prostaglandin receptors.^{154, 157, 158} In other words, calcitriol has the potential to antagonize the bioactivity of prostaglandins, and thus presumably could complement the protection afforded by agents that inhibit cox-2 (such as aspirin) or suppress its expression. Indeed, Feldman and colleagues have demonstrated that, in prostate cancer cell lines that are moderately responsive to the anti-proliferative impact of calcitriol alone, this hormone can markedly potentiate the growth inhibitory effects of sub-optimal concentrations of NSAID drugs.¹⁵⁴

It seems likely, however, that calcitriol can work in other, complementary ways to promote apoptosis or suppress proliferation in pre-cancerous tissues. For example, in many tissues, calcitriol or less calcemic analogs thereof have been shown to suppress NF-kappaB signaling, either by reducing expression of its components, by stabilizing or inducing IkappaB, or by acting within the nucleus to antagonize the transactivational activity of NF-kappaB.¹⁵⁹⁻¹⁷¹ (Up-regulation of NF-kappaB signaling by vitamin D activity has however been demonstrated in some contexts.¹⁷²) A vitamin D-mediated increase in IGFBP-3 expression has also been reported in some tissues; IGFBP-3 counteracts the cancer-promotional activity of IGF-I, and also can exert direct antiproliferative effects in some cell lines.^{169, 173-181} In some tissues the activated vitamin D receptor may boost the transcriptional activity of anticarcinogenic FOXO factors by facilitating their interaction with Sirt1.¹⁸² And it has been suggested that vitamin D-mediated up-regulation of cadherin expression and tight junction formation could have an anti-promotional effect on epithelia.¹⁸³⁻¹⁸⁵ No doubt the cancer-preventive effects of good vitamin D status will prove to be complex and cell type specific.

Importantly, there is recent evidence that supplemental intakes as high as 10,000 IU daily may be required to achieve in almost all subjects the serum levels of 25-hydroxycholecalciferol associated with optimal cancer protection in epidemiological studies.¹⁸⁶ Fortunately, intakes below 40,000 IU daily are likely to be safe.¹⁸⁶⁻¹⁸⁸

Long-Chain Omega-3s Limit PGE2 Production and Activity

Another feasible way to blunt the cancer-promoting activity of cox-2 is to limit its access to arachidonic acid, the precursor for the prostaglandins which mediate this activity. This can be achieved by minimizing the dietary intake of omega-6 fats relative to the intake of the long-chain omega-3s (EPA, DHA) found predominantly in fish oil; EPA can act as a competitive inhibitor of arachidonate binding to cox-2, and the PGE3 and other prostanoids derived from EPA tend to have anti-inflammatory rather than cancer-promoting activity. PGE3 fails to activate the EP2/EP4 receptors, and indeed acts as a competitive antagonist of PGE2 in this regard.¹⁸⁹ Epidemiological studies focusing on cancers of the breast, prostate, and colon have concluded that diets in which the ratio of omega-6 fats to long-chain omega-3 fats are relatively low, or a low omega-6/long-chain omega-3 ratio in blood or tissues, are associated with decreased risk.¹⁹⁰⁻²⁰⁰ A similar effect has been observed in rodent models of cancer promotion.²⁰¹⁻²⁰⁸ These considerations suggest that a “Mediterranean” profile of dietary fat – in which monounsaturates are the predominant fat, and intakes of long-chain omega-3s are significant – may be favorable for cancer prevention. It should be noted that, whereas supplemental fish oil reduced rectal cell proliferation (suspected to be a surrogate of colorectal cancer risk) in subjects consuming a low-diet, no such protection was noted when such supplementation was implemented in the context of diets high in omega-6.²⁰⁹ Vegetarians should be aware that alpha-linolenic acid – the chief omega-3 in strict vegetarian diets – does not emerge as protective in some epidemiology, likely because humans convert it only inefficiently to the longer chain omega-3s that antagonize the cancer-promoting activity of cox-2.^{199, 210-212} However, stearidonic acid (18:4n-3), which could be made in high yield by bioengineered plants, is more efficiently converted to longer chain fats, and may well have potential for cancer prevention.²¹³⁻²¹⁶

Lifestyle Modulates Cancer Risk via Systemic Growth Factor Activity and Cox-2 Expression

As noted above, growth factors can up-regulate cox-2 expression, both by activating Erk1/2, and by lengthening the half-life of cox-2 mRNA via an effect mediated by the PI3K/Akt pathway. In particular, IGF-I has been shown to have this effect in various cells lines. Insulin is also pertinent to this effect – either by a direct impact on insulin receptors, or by up-regulating systemic IGF-I activity by suppressing hepatic production of the IGF-I antagonist IGFBP-1.²¹⁷⁻²¹⁹ The cancer prevention afforded by calorie restriction in rodent studies is mediated at least in part by a chronic down-regulation of circulating levels of insulin and free IGF-I,²²⁰ and there is good reason to suspect that the far lower risks for many “Western” cancers enjoyed by various Third World cultures during the last century reflected the joint influence of quasi-vegan diets of modest protein content - which tend to down-regulate hepatic production of IGF-I owing to essential amino acid restriction - and of leanness and good muscle insulin sensitivity, which minimize diurnal insulin levels.^{217-219, 221-224} Conceivably, increased levels of adiponectin, and modulation of mTOR activation and autocrine growth factor activity in cancer-prone tissues, might also contribute to the cancer protection afforded by such a lifestyle.²²⁵ For example, increased autocrine production of IGF-II appears to mediate the promotional impact of dietary casein on hepatocarcinogenesis in rodents.²²⁶

A reduction in systemic insulin/IGF-I activity could be expected to decrease cancer risk by influencing both mutagenesis and promotion. Since mistakes are made whenever DNA is replicated, reduced proliferation in cancer-susceptible tissues would be expected to slow the rate at which stem cells accumulate heritable mutations. Moreover, insulin/IGF-I signaling may promote oxidant mutagenesis by suppressing FOXO3a-mediated transcription of the genes for catalase, the mitochondrial superoxide dismutase, and other antioxidant enzymes.²²⁷⁻²³¹ And growth factor activity works in a number of complementary ways (including cox-2 expression) to suppress apoptosis – thereby increasing the risk that pre-cancerous cells will survive long enough to acquire the mutations that turn them into full-fledged malignancies. Growth factor stimulation of Akt is particularly versatile in its suppressive impact on apoptosis.²³² A virtually obligate role for liver-derived systemic IGF-I in human carcinogenesis is suggested by the near absence of cancer in individuals who are homozygous for loss of function mutations in the gene for growth hormone receptor, in whom hepatic production of IGF-I is trivial.^{233, 234} Vegans achieve a much less drastic suppression of hepatic IGF-I production.^{223, 235}

With respect to breast cancer, low systemic growth factor activities may aid its prevention in part by delaying menarche and accelerating menopause.²³⁶⁻²³⁸ This protective effect is compounded by the fact that serum estradiol levels are notably lower in rural Chinese women than in Western controls throughout life;^{222, 239} premenopausally, this may reflect the fact that IGF-I activity potentiates ovarian estrogen synthesis.²⁴⁰⁻²⁴² This latter effect is not pertinent postmenopausally, when the lower estrogen levels and activity in Chinese women may be attributable to relative leanness and the increase in sex-hormone-binding globulin levels associated with good insulin sensitivity.²³⁹ (Perhaps this explains why IGF-I levels correlate with breast cancer risk premenopausally but not postmenopausally in some Western studies.) It should be noted that early pregnancy may often contribute to reduced breast cancer risk in certain Third World societies, and multiple pregnancies diminish risk for ovarian cancer.

The cancer protection afforded by lifelong consumption of a plant-based diet, in conjunction with leanness and insulin sensitivity (which tend to be promoted by low-fat plant-based diets, owing in part to their low saturate/unsaturate ratio²⁴³) may be very substantial indeed. Figure 1 (derived from data

published by Ernt Wynder and colleagues²⁴⁴) compares age-adjusted mortalities from various “Western” cancers in the populations of the U.S. and Japan, ca. 1955. Post-war Japan at this time was still a rather poor society; a national nutrition survey in 1950 determined that the Japanese were getting only about 6% of their calories from animal products (most of this from fish), as opposed to 78% of calories from grains.²⁴⁵ Their omega-6 intake was quite low, so the modest amount of fat obtained from fish may have resulted in a low omega-6/omega-3 ratio. The Japanese at this time also tended to be quite lean, reflecting lifelong consumption of a diet of extremely low fat content; yet, unlike the Okinawans, they were not calorically restricted.²⁴⁵ Note that age-adjusted death rates from cancers of the colon, prostate, breast, and ovary were on the order of 5-10-fold lower in Japan than in the U.S. at that time; mortality from pancreatic cancer, leukemias, and lymphomas was 3-4-fold lower in Japan.

But this phenomenon was by no means isolated to Japan; Western cancers were likewise comparatively rare in other Third World societies in which lean people ate plant-based diets. A paper by Hebert and colleagues provides data from the World Health Organization that provides estimated average daily food intakes for 59 countries, ca. 1979-1981; notably, these data estimate the percentage of daily calories provided by animal products in these countries.²⁴⁶ The same paper provides age-adjusted prostate cancer mortality statistics in these same 59 countries for 1985-1989. The 6 countries on the list in which animal products were estimated to provide less than 10% of dietary calories (Egypt, Guatemala, Honduras, South Korea, Sri Lanka, and Thailand) subsequently experienced an average prostate cancer mortality rate of 1.96 per 100,000 males; in the 20 countries on the list which obtained over 1,000 kilocalories daily from animal products – including the U.S. and most Western European countries – this death rate averaged 29.91 per 100,000 males – 15-fold higher! (And the differential in *risk* for clinically significant prostate cancer was undoubtedly *higher* than that, as people in the poorer quasi-vegan countries would presumably have less access to sophisticated medical care, and thus would be less likely to achieve surgical cures of early stage cancer.)

With respect to breast cancer, consider the data that Kenneth Carroll compiled correlating average daily dietary animal fat intake (a rough surrogate for animal calorie intake) with age-adjusted breast cancer mortality in a number of countries, in the mid 1960s.²⁴⁷ (See Chart 4 in this publication.) In the 6 countries where people were estimated to get no more than 20 g daily of fat from animal products, the breast cancer mortality averaged about 3.5 deaths per 100,000 females. In contrast, in the U.S., Canada, and Australia, this death rate averaged about 21 per 100,000. No doubt persistent epidemiologists could compile a number of similar examples. Are there any counterexamples of a society which consumed a predominantly plant-based diet, in which the people were typically lean and insulin sensitive – and Western cancer rates were high? This author has yet to encounter one.

Colin Campbell, in an analysis of data derived from the ecologic China Study (China-Oxford-Cornell Diet and Health Project), concluded that plasma cholesterol was the chief correlate of the range of Western cancers, as well as diabetes and coronary disease.²⁴⁸ In quasi-vegan societies, such as those surveyed in this study, plasma cholesterol serves as a good marker for animal product consumption, since dietary cholesterol consumption reliably raises plasma cholesterol in vegans (whereas an increase of cholesterol consumption typically has minimal impact on the plasma cholesterol of omnivores).^{249, 250} Campbell notes that, within the quasi-vegan provinces of rural China, “small intakes of foods of animal origin are associated with significant increases in plasma cholesterol concentrations, which are associated,

in turn, with significant increases in chronic degenerative disease mortality rates.”²⁴⁸ Once again, the regular consumption of animal products emerges as a major determinant of Western cancer risk. (However, increased red blood cell levels of DHA, presumably indicative of fish consumption, correlated negatively with Western disease risk in this population; this likely reflects the protection afforded by fish oil, rather than fish protein.)²⁵¹

The promotional impact of high-quality protein on cancer risk has yet to win broad acceptance, in large measure because it fails to emerge in most Western epidemiology. This likely reflects the fact that the impact of essential amino acid status on the bioactivity of IGF-I and perhaps other growth factors plateaus once a certain minimal intake is reached. In the Nurses’ Health Study, the *bottom* quintile of animal protein intake corresponds to 12% of calories.²⁵² In contrast, the populations surveyed by Campbell in the China Study got, on average, less than 1% of calories from animal protein, with total protein intake accounting for about 14% of calories.²²² Consideration should be given to the possibility that the *ratio* of nonessential to essential amino acids ingested has an impact on growth factor activities; a high intake of nonessential amino acids might decrease the effective availability of essential amino acids by promoting amino acid catabolism via glucagon.²⁴³ Red meat does however emerge as a risk factor for cancer in much Western epidemiology; this could reflect the higher levels of heme iron and saturated fat in red meat. Campbell has repeatedly emphasized that exclusive reliance on Western epidemiological data bases can be counterproductive when those data bases fail to encompass lifestyles associated with marked health protection in other societies.²²²

The impact of obesity and metabolic syndrome on cancer risk – likely mediated by factors such as hyperinsulinemia, elevated free fatty acid flux, reduced adiponectin, and increased aromatase activity – is of course well documented in Western epidemiology, as is the protection associated with regular exercise. Third World populations at low risk for Western cancers tend to be quite lean by Western standards – the average BMI in the China Study population was 21, despite calorie intakes estimated to be about 30% higher than those of Americans of comparable body size.²⁵³ A plant-based diet of moderate fat content, in conjunction with a physically active lifestyle, is inherently productive of leanness²⁴³ – as repeatedly demonstrated by physicians who employ this strategy therapeutically.²⁵⁴⁻²⁵⁶

It is obvious on its face that it is vitally important to understand why age-adjusted death rates from many prominent Western cancers have been virtually an order of magnitude lower in many societies as compared to the West. Yet, with a few honorable exceptions, medical researchers have devoted relatively little attention to this crucial question. If some view the explanations attempted here as inadequate – where are the credible alternative hypotheses?

Nonetheless, it should be acknowledged that certain types of cancer – notably gastric, hepatic, bladder, and cervical – are often common in the Third World. But it seems likely that this reflects, not any inherent failing of a quasi-vegan diet, but rather poor hygiene, including poor food hygiene, that leads to chronic infections such as hepatitis, *H. pylori*, schistosomiasis, and papilloma virus, and to ingestion of mutagens produced by bacterial or mold contamination of improperly stored food. It is not accidental that gastric cancer rates tend to plunge when a society gains access to refrigeration.²⁵⁷ The commonly noted ecological association between high-salt diets and gastric cancer may reflect, in part, the inadequacy of salt as a food preservative; however, heavily salted foods, even if uncontaminated, may play a pathogenic

role in gastric cancer induction, and their regular consumption should be discouraged.²⁵⁸ In Mongolian gerbils, a diet extremely high in salt (10%) boosts the incidence of gastric cancer evoked by either concurrent *H. pylori* infection or administration of gastric carcinogens.²⁵⁹⁻²⁶¹ Intriguingly, a salty diet amplified the induction of cox-2 and iNOS in the gastric mucosa of gerbils infected with *H. pylori*.²⁶² Salt alone had no such inductive effect, a finding which accords nicely with cohort epidemiology concluding that dietary salt may represent a risk for gastric cancer only in individuals concurrently infected with *H. pylori*.²⁶³ Not unlikely, cox-2 induction contributes to the nexus between other types of chronic infection and cancer risk.^{264, 265}

Encouragingly, Campbell's analysis of the China Study noted that risks for both gastric and hepatic cancer trended lower in the more thoroughly vegan provinces; he supplemented this observation by demonstrating that low but nutritionally adequate protein intakes suppress induction of hepatic cancer by aflatoxin or hepatitis B in rats.^{248, 266-268} Vegan diets, if hygienically stored and prepared, may be lower in mutagen load than omnivore diets. They do not contain heme iron, and also do not contain the mutagenic heterocyclic amines produced in flesh exposed to high heat.^{269, 270} (These are made only in flesh foods, as creatine is an obligate precursor.) Moreover, vegan diets are devoid of animal fat, and thus tend to be lower in the fat-soluble xenobiotic compounds (some of which may be mutagenic) that accumulate in animal fat.^{271, 272}

As noted above, diets of low variety deficient in antioxidant nutrients and phytochemicals may contribute importantly to high risks for various squamous cell carcinomas (especially esophageal) in certain regions of the Third World. Obviously, whole-food vegan diets of good variety can supply these protective nutrients in abundance.

Calorie Restriction Suppresses Cox-2 Expression

As noted, calorie restriction tends to prevent or postpone cancer in rodents, largely because of its impact on growth factor activities. The sirt1 induction associated with such diets could be expected to suppress NF-kappaB activation,²⁷³⁻²⁷⁵ and thereby down-regulate cox-2 expression. Not surprisingly, calorie restriction has been reported to decrease cox-2 expression in rodents.²⁷⁶⁻²⁷⁸ Although daily calorie restriction achieved by a strict accounting of calories is likely to prove impractical for all but a tiny percentage of people, potentially more feasible regimens such as modified alternate day fasting and carbohydrate-concentrated diets have been proposed as strategies for achieving a measure of calorie restriction in humans.²⁷⁹⁻²⁸²

Soy Isoflavones – Does ERbeta Activity Down-Regulate Cox-2?

A number of epidemiological studies have found that increased intakes of soyfoods or of soy isoflavones are associated with reduced risk for prostate, breast, and colorectal cancers – the chief causes of cancer mortality among non-smokers in the West.²⁸³⁻²⁹¹ Decreased risks for ovarian, endometrial, and gastric cancers have also been correlated with increased dietary soy.^{292, 293} These findings are most consistent in East Asian studies, presumably reflecting habitual intakes of soy in East Asia that are higher and more physiologically meaningful; meta-analyses of pertinent East Asian studies observe a robust protective effect of soy (with the exception of colorectal cancer risk in men). These findings should be interpreted

cautiously, in light of the fact that a high soy intake may serve as a marker for a diet that is less Westernized.

Nonetheless, there are compelling theoretical grounds for suspecting that soy isoflavones may indeed be protective. Genistein can act as a pure agonist for both the alpha and beta isoforms of the estrogen receptor, but its affinity for the beta isoform is over an order of magnitude higher.^{294, 295} Physiological intakes of soyfoods yield plasma levels of free, unconjugated genistein in the low nanomolar range. These concentrations may be sufficient to achieve meaningful activation of ERbeta, while having minimal impact on ERalpha (hence explaining why high-soy diets don't have feminizing effects, known to be mediated by ERalpha).²⁹⁶ In contrast to ERalpha, ERbeta tends to have anti-proliferative, pro-apoptotic, and differentiating effects on tissues which express it – and all the cancers for which soy is believed to be protective arise from tissues which express ERbeta.²⁹⁷⁻²⁹⁹ In tissues which express both isoforms of the estrogen receptor, ERbeta appears to antagonize the cancer-promotional effects of ERalpha. This may rationalize the curious observation that, in East Asian studies, high soy intakes are associated with decreased risk for breast cancer and with improved prognosis in women who have breast cancer²⁸⁵ – despite the evident promotional impact of high estrogen exposure on this cancer. Colon epithelium, not ordinarily thought of as an estrogen sensitive tissue, expresses solely ERbeta.³⁰⁰ This may explain not only the favorable impact on colorectal cancer risk associated with dietary soy, but also the reduction in risk for this cancer seen in postmenopausal women who use estrogen replacement therapy.^{301, 302} With respect to gastric cancer, gastric epithelium expresses both ER isoforms, loss of expression of ERbeta correlates with poor prognosis in gastric cancer, and this malignancy is only about half as common in women as in men.³⁰³⁻³⁰⁵

How ERbeta might influence cox-2 expression is not clear. Nonetheless, there are several reports that ingestion of soy isoflavones can suppress induction of cox-2. This has been observed in the colonic mucosa of rats treated with the carcinogen dimethylhydrazine, in the uteri of ovariectomized mice treated with estrogen, and in the prostate cancers of patients administered soy isoflavones for two weeks prior to prostatectomy.³⁰⁶⁻³⁰⁸ Also pertinent is a report that an ERbeta-selective agonist can inhibit induction of cox-2 in keratinocytes exposed to UV light or TNFalpha³⁰⁹ – in nice accord with evidence that oral or topical soy isoflavones can inhibit photoaging.^{310, 311} Further studies should determine whether and how ERbeta activation can suppress cox-2 induction in various tissues which express this receptor. (Those examining the pertinent literature should be cautioned that most in vitro studies with genistein are physiologically irrelevant, owing to the fact that they employ supraphysiological micromolar concentrations that would be expected to activate both ER isoforms, and to exert additional effects such as tyrosine kinase inhibition; moreover, rodent studies involving high isoflavone intakes may be a poor guide to human experience, as these high intakes may activate ERalpha.²⁹⁶)

As a proviso, there is evidence that, within the context of a vegan diet, high intakes of soy protein may boost IGF-I levels, probably reflecting the fact that soy protein is one of the higher quality plant proteins.^{224, 312} So vegans wishing to achieve a high intake of soy isoflavones might be well advised to ingest an isoflavone supplement rather than ingesting very large amounts of soyfoods or soy protein.

A Lifestyle Protocol for Minimizing Cancer Risk

The key point of this essay is that scope for human cancer prevention may be huge. The results of the recent aspirin meta-analyses – suggesting a reduction of cancer mortality by about one-third in subjects taking aspirin for at least 7.5 years – can justly be called astounding. Yet the protection from “Western” cancers enjoyed by Japanese in 1955 – most of whom were not taking aspirin! – is even more dramatic. There is good reason to suspect that an optimally-designed cancer-preventive lifestyle, integrating a range of protective options that target cox-2 bioactivity (albeit exerting a number of complementary cancer-preventive effects), could have a huge impact on cancer risk – a benefit that is all the more valuable given the likelihood that a “magic bullet” for cancer therapy seems likely to elude us for quite some time, if not permanently.

In light of the above discussion, such a lifestyle could include:

A whole-food plant-based diet (no or minimal animal protein), low in saturated and omega-6 fats, moderate in monounsaturates (as from almonds and olive oil) and in protein, and rich in fruits, vegetables and herbs that provide phase 2 inducers (crucifera, garlic and onions, green tea, etc.) and vitamin C;

Aerobic exercise training configured in a way that promotes leanness and good insulin sensitivity;³¹³⁻³¹⁵

Daily low-dose aspirin, enteric-coated;

A supplementation regimen including vitamin D (4,000-10,000 IU daily), EPA/DHA (or stearidonic acid, when available), spirulina (or PhyCB-enriched spirulina extracts), astaxanthin, melatonin (before bedtime), nutraceutical phase 2 inducers (reasonable choices could include lipoic acid, green tea polyphenols, aged garlic extracts, sulforaphane), N-acetylcysteine, soy isoflavones, and vitamin B12;

And - for the truly dedicated - moderate calorie restriction achieved by strategies such as modified alternate day fasting or carbohydrate-concentrated dieting.

Finally – this should be too obvious to mention! – don’t smoke or drink alcohol abusively. Folate supplementation may be advisable in those who drink moderately. Heavily salted foods should not be eaten regularly, and microbial contamination of food should be avoided by refrigeration and other appropriate measures.

Importantly, this strategy - especially if complemented by moderate salt restriction and high potassium intake - could also be expected to minimize risk for vascular disorders, type 2 diabetes, and dementia, and hence would be expected to have a remarkably favorable impact on healthful lifespan.

References

- (1) Rothwell PM, Wilson M, Elwin CE, Norrving B, Algra A, Warlow CP, Meade TW. Long-term effect of aspirin on colorectal cancer incidence and mortality: 20-year follow-up of five randomised trials. *Lancet* 2010 November 20;376(9754):1741-50.
- (2) Rothwell PM, Fowkes FG, Belch JF, Ogawa H, Warlow CP, Meade TW. Effect of daily aspirin on long-term risk of death due to cancer: analysis of individual patient data from randomised trials. *Lancet* 2010 December 6.
- (3) Chan AT, Ogino S, Fuchs CS. Aspirin and the risk of colorectal cancer in relation to the expression of COX-2. *N Engl J Med* 2007 May 24;356(21):2131-42.
- (4) Chan AT, Ogino S, Fuchs CS. Aspirin use and survival after diagnosis of colorectal cancer. *JAMA* 2009 August 12;302(6):649-58.
- (5) Chun KS, Akunda JK, Langenbach R. Cyclooxygenase-2 inhibits UVB-induced apoptosis in mouse skin by activating the prostaglandin E2 receptors, EP2 and EP4. *Cancer Res* 2007 March 1;67(5):2015-21.
- (6) Chu AJ, Chou TH, Chen BD. Prevention of colorectal cancer using COX-2 inhibitors: basic science and clinical applications. *Front Biosci* 2004 September 1;9:2697-713.
- (7) Wu T, Leng J, Han C, Demetris AJ. The cyclooxygenase-2 inhibitor celecoxib blocks phosphorylation of Akt and induces apoptosis in human cholangiocarcinoma cells. *Mol Cancer Ther* 2004 March;3(3):299-307.
- (8) Mutoh M, Takahashi M, Wakabayashi K. Roles of prostanoids in colon carcinogenesis and their potential targeting for cancer chemoprevention. *Curr Pharm Des* 2006;12(19):2375-82.
- (9) Wang D, DuBois RN. Cyclooxygenase-2: a potential target in breast cancer. *Semin Oncol* 2004 February;31(1 Suppl 3):64-73.
- (10) Wendum D, Masliah J, Trugnan G, Flejou JF. Cyclooxygenase-2 and its role in colorectal cancer development. *Virchows Arch* 2004 October;445(4):327-33.
- (11) Dempke W, Rie C, Grothey A, Schmoll HJ. Cyclooxygenase-2: a novel target for cancer chemotherapy? *J Cancer Res Clin Oncol* 2001 July;127(7):411-7.
- (12) Greenhough A, Wallam CA, Hicks DJ, Moorghen M, Williams AC, Paraskeva C. The proapoptotic BH3-only protein Bim is downregulated in a subset of colorectal cancers and is repressed by antiapoptotic COX-2/PGE(2) signalling in colorectal adenoma cells. *Oncogene* 2010 June 10;29(23):3398-410.

- (13) Basu GD, Pathangey LB, Tinder TL, Lagioia M, Gendler SJ, Mukherjee P. Cyclooxygenase-2 inhibitor induces apoptosis in breast cancer cells in an in vivo model of spontaneous metastatic breast cancer. *Mol Cancer Res* 2004 November;2(11):632-42.
- (14) Lin DW, Nelson PS. The role of cyclooxygenase-2 inhibition for the prevention and treatment of prostate carcinoma. *Clin Prostate Cancer* 2003 September;2(2):119-26.
- (15) Sun Y, Tang XM, Half E, Kuo MT, Sinicrope FA. Cyclooxygenase-2 overexpression reduces apoptotic susceptibility by inhibiting the cytochrome c-dependent apoptotic pathway in human colon cancer cells. *Cancer Res* 2002 November 1;62(21):6323-8.
- (16) Vane JR, Bakhle YS, Botting RM. Cyclooxygenases 1 and 2. *Annu Rev Pharmacol Toxicol* 1998;38:97-120.
- (17) Appleby SB, Ristimaki A, Neilson K, Narko K, Hla T. Structure of the human cyclooxygenase-2 gene. *Biochem J* 1994 September 15;302 (Pt 3):723-7.
- (18) Crofford LJ, Tan B, McCarthy CJ, Hla T. Involvement of nuclear factor kappa B in the regulation of cyclooxygenase-2 expression by interleukin-1 in rheumatoid synoviocytes. *Arthritis Rheum* 1997 February;40(2):226-36.
- (19) Newton R, Kuitert LM, Bergmann M, Adcock IM, Barnes PJ. Evidence for involvement of NF-kappaB in the transcriptional control of COX-2 gene expression by IL-1beta. *Biochem Biophys Res Commun* 1997 August 8;237(1):28-32.
- (20) Bauer MK, Lieb K, Schulze-Osthoff K, Berger M, Gebicke-Haerter PJ, Bauer J, Fiebich BL. Expression and regulation of cyclooxygenase-2 in rat microglia. *Eur J Biochem* 1997 February 1;243(3):726-31.
- (21) Inoue H, Tanabe T. Transcriptional role of the nuclear factor kappa B site in the induction by lipopolysaccharide and suppression by dexamethasone of cyclooxygenase-2 in U937 cells. *Biochem Biophys Res Commun* 1998 March 6;244(1):143-8.
- (22) Subbaramaiah K, Chung WJ, Dannenberg AJ. Ceramide regulates the transcription of cyclooxygenase-2. Evidence for involvement of extracellular signal-regulated kinase/c-Jun N-terminal kinase and p38 mitogen-activated protein kinase pathways. *J Biol Chem* 1998 December 4;273(49):32943-9.
- (23) Chen LC, Chen BK, Chang JM, Chang WC. Essential role of c-Jun induction and coactivator p300 in epidermal growth factor-induced gene expression of cyclooxygenase-2 in human epidermoid carcinoma A431 cells. *Biochim Biophys Acta* 2004 July 5;1683(1-3):38-48.
- (24) Hsu MJ, Chang CK, Chen MC, Chen BC, Ma HP, Hong CY, Lin CH. Apoptosis signal-regulating kinase 1 in peptidoglycan-induced COX-2 expression in macrophages. *J Leukoc Biol* 2010 June;87(6):1069-82.
- (25) Adderley SR, FitzGerald DJ. Oxidative damage of cardiomyocytes is limited by extracellular regulated kinases 1/2-mediated induction of cyclooxygenase-2. *J Biol Chem* 1999 February 19;274(8):5038-46.

- (26) Aggeli IK, Kefaloyianni E, Beis I, Gaitanaki C. HOX-1 and COX-2: Two differentially regulated key mediators of skeletal myoblast tolerance under oxidative stress. *Free Radic Res* 2010 June;44(6):679-93.
- (27) Stoeltzing O, Liu W, Fan F, Wagner C, Stengel K, Somcio RJ, Reinmuth N, Parikh AA, Hicklin DJ, Ellis LM. Regulation of cyclooxygenase-2 (COX-2) expression in human pancreatic carcinoma cells by the insulin-like growth factor-I receptor (IGF-IR) system. *Cancer Lett* 2007 December 18;258(2):291-300.
- (28) Nguyen T, Chai J, Li A, Akahoshi T, Tanigawa T, Tarnawski AS. Novel roles of local insulin-like growth factor-1 activation in gastric ulcer healing: promotes actin polymerization, cell proliferation, re-epithelialization, and induces cyclooxygenase-2 in a phosphatidylinositol 3-kinase-dependent manner. *Am J Pathol* 2007 April;170(4):1219-28.
- (29) Cao Z, Liu LZ, Dixon DA, Zheng JZ, Chandran B, Jiang BH. Insulin-like growth factor-I induces cyclooxygenase-2 expression via PI3K, MAPK and PKC signaling pathways in human ovarian cancer cells. *Cell Signal* 2007 July;19(7):1542-53.
- (30) Di PA, Memoli A, Apicella A, Tuccillo C, di PA, Ricchi P, Acquaviva AM, Zarrilli R. IGF-II/IGF-I receptor pathway up-regulates COX-2 mRNA expression and PGE2 synthesis in Caco-2 human colon carcinoma cells. *Oncogene* 2000 November 16;19(48):5517-24.
- (31) Nakamura T, Sakamoto K. Reactive oxygen species up-regulates cyclooxygenase-2, p53, and Bax mRNA expression in bovine luteal cells. *Biochem Biophys Res Commun* 2001 June 1;284(1):203-10.
- (32) Feng L, Xia Y, Garcia GE, Hwang D, Wilson CB. Involvement of reactive oxygen intermediates in cyclooxygenase-2 expression induced by interleukin-1, tumor necrosis factor-alpha, and lipopolysaccharide. *J Clin Invest* 1995 April;95(4):1669-75.
- (33) Eligini S, Arenaz I, Barbieri SS, Faleri ML, Crisci M, Tremoli E, Colli S. Cyclooxygenase-2 mediates hydrogen peroxide-induced wound repair in human endothelial cells. *Free Radic Biol Med* 2009 May 15;46(10):1428-36.
- (34) Chiu WT, Shen SC, Chow JM, Lin CW, Shia LT, Chen YC. Contribution of reactive oxygen species to migration/invasion of human glioblastoma cells U87 via ERK-dependent COX-2/PGE(2) activation. *Neurobiol Dis* 2010 January;37(1):118-29.
- (35) Sun Y, Chen J, Rigas B. Chemopreventive agents induce oxidative stress in cancer cells leading to COX-2 overexpression and COX-2-independent cell death. *Carcinogenesis* 2009 January;30(1):93-100.
- (36) Jaimes EA, Zhou MS, Pearse DD, Puzis L, Raij L. Upregulation of cortical COX-2 in salt-sensitive hypertension: role of angiotensin II and reactive oxygen species. *Am J Physiol Renal Physiol* 2008 February;294(2):F385-F392.
- (37) Shanmugam N, Gaw G, I, Natarajan R. Molecular mechanisms of high glucose-induced cyclooxygenase-2 expression in monocytes. *Diabetes* 2004 March;53(3):795-802.

- (38) Barbieri SS, Eligini S, Brambilla M, Tremoli E, Colli S. Reactive oxygen species mediate cyclooxygenase-2 induction during monocyte to macrophage differentiation: critical role of NADPH oxidase. *Cardiovasc Res* 2003 October 15;60(1):187-97.
- (39) Nakagiri A, Murakami M. Roles of NADPH oxidase in occurrence of gastric damage and expression of cyclooxygenase-2 during ischemia/reperfusion in rat stomachs. *J Pharmacol Sci* 2009 December;111(4):352-60.
- (40) Nishanth RP, Ramakrishna BS, Jyotsna RG, Roy KR, Reddy GV, Reddy PK, Reddanna P. C-Phycocyanin inhibits MDR1 through reactive oxygen species and cyclooxygenase-2 mediated pathways in human hepatocellular carcinoma cell line. *Eur J Pharmacol* 2010 December 15;649(1-3):74-83.
- (41) Sheu ML, Chiang CK, Tsai KS, Ho FM, Weng TI, Wu HY, Liu SH. Inhibition of NADPH oxidase-related oxidative stress-triggered signaling by honokiol suppresses high glucose-induced human endothelial cell apoptosis. *Free Radic Biol Med* 2008 June 15;44(12):2043-50.
- (42) Hougee S, Hartog A, Sanders A, Graus YM, Hoijer MA, Garssen J, Van Den Berg WB, van Beuningen HM, Smit HF. Oral administration of the NADPH-oxidase inhibitor apocynin partially restores diminished cartilage proteoglycan synthesis and reduces inflammation in mice. *Eur J Pharmacol* 2006 February 15;531(1-3):264-9.
- (43) Barbieri SS, Cavalca V, Eligini S, Brambilla M, Caiani A, Tremoli E, Colli S. Apocynin prevents cyclooxygenase 2 expression in human monocytes through NADPH oxidase and glutathione redox-dependent mechanisms. *Free Radic Biol Med* 2004 July 15;37(2):156-65.
- (44) Wang T, Qin L, Liu B, Liu Y, Wilson B, Eling TE, Langenbach R, Taniura S, Hong JS. Role of reactive oxygen species in LPS-induced production of prostaglandin E2 in microglia. *J Neurochem* 2004 February;88(4):939-47.
- (45) McCarty MF. "Iatrogenic Gilbert syndrome"--a strategy for reducing vascular and cancer risk by increasing plasma unconjugated bilirubin. *Med Hypotheses* 2007;69(5):974-94.
- (46) Lanone S, Bloc S, Foresti R, Almolki A, Taille C, Callebort J, Conti M, Goven D, Aubier M, Dureuil B, El-Benna J, Motterlini R, Bockowski J. Bilirubin decreases nos2 expression via inhibition of NAD(P)H oxidase: implications for protection against endotoxic shock in rats. *FASEB J* 2005 November;19(13):1890-2.
- (47) Matsumoto H, Ishikawa K, Itabe H, Maruyama Y. Carbon monoxide and bilirubin from heme oxygenase-1 suppresses reactive oxygen species generation and plasminogen activator inhibitor-1 induction. *Mol Cell Biochem* 2006 October;291(1-2):21-8.
- (48) Jiang F, Roberts SJ, Datla S, Dusting GJ. NO modulates NADPH oxidase function via heme oxygenase-1 in human endothelial cells. *Hypertension* 2006 November;48(5):950-7.
- (49) Datla SR, Dusting GJ, Mori TA, Taylor CJ, Croft KD, Jiang F. Induction of heme oxygenase-1 in vivo suppresses NADPH oxidase derived oxidative stress. *Hypertension* 2007 October;50(4):636-42.

- (50) Schwertner HA, Jackson WG, Tolan G. Association of low serum concentration of bilirubin with increased risk of coronary artery disease. *Clin Chem* 1994 January;40(1):18-23.
- (51) Lin JP, O'Donnell CJ, Schwaiger JP, Cupples LA, Lingenhel A, Hunt SC, Yang S, Kronenberg F. Association between the UGT1A1*28 allele, bilirubin levels, and coronary heart disease in the Framingham Heart Study. *Circulation* 2006 October 3;114(14):1476-81.
- (52) Schwertner HA, Vitek L. Gilbert syndrome, UGT1A1*28 allele, and cardiovascular disease risk: possible protective effects and therapeutic applications of bilirubin. *Atherosclerosis* 2008 May;198(1):1-11.
- (53) Lin JP, Vitek L, Schwertner HA. Serum Bilirubin and Genes Controlling Bilirubin Concentrations as Biomarkers for Cardiovascular Disease. *Clin Chem* 2010 August 6.
- (54) Ohnaka K, Kono S, Inoguchi T, Yin G, Morita M, Adachi M, Kawate H, Takayanagi R. Inverse associations of serum bilirubin with high sensitivity C-reactive protein, glycated hemoglobin, and prevalence of type 2 diabetes in middle-aged and elderly Japanese men and women. *Diabetes Res Clin Pract* 2010 April;88(1):103-10.
- (55) Inoguchi T, Sasaki S, Kobayashi K, Takayanagi R, Yamada T. Relationship between Gilbert syndrome and prevalence of vascular complications in patients with diabetes. *JAMA* 2007 September 26;298(12):1398-400.
- (56) Inoguchi T, Nawata H. NAD(P)H oxidase activation: a potential target mechanism for diabetic vascular complications, progressive beta-cell dysfunction and metabolic syndrome. *Curr Drug Targets* 2005 June;6(4):495-501.
- (57) Temme EH, Zhang J, Schouten EG, Kesteloot H. Serum bilirubin and 10-year mortality risk in a Belgian population. *Cancer Causes Control* 2001 December;12(10):887-94.
- (58) Horsfall LJ, Rait G, Walters K, Swallow DM, Pereira SP, Nazareth I, Petersen I. Serum bilirubin and risk of respiratory disease and death. *JAMA* 2011 February 16;305(7):691-7.
- (59) Zucker SD, Horn PS, Sherman KE. Serum bilirubin levels in the U.S. population: gender effect and inverse correlation with colorectal cancer. *Hepatology* 2004 October;40(4):827-35.
- (60) Ioannou GN, Liou IW, Weiss NS. Serum bilirubin and colorectal cancer risk: a population-based cohort study. *Aliment Pharmacol Ther* 2006 June 1;23(11):1637-42.
- (61) Zucker SD, Benedict M, Sherman KE. Serum bilirubin and risk of colorectal cancer. *Aliment Pharmacol Ther* 2006 October 15;24(8):1257-9.
- (62) Vitek L. Bilirubin and colorectal cancer. *Aliment Pharmacol Ther* 2006 November 15;24(10):1503-4.
- (63) Lacko M, Roelofs HM, Te Morsche RH, Voogd AC, Oude Ophuis MB, Peters WH, Manni JJ. Genetic polymorphism in the conjugating enzyme UGT1A1 and the risk of head and neck cancer. *Int J Cancer* 2010 March 3.

- (64) Exner M, Minar E, Wagner O, Schillinger M. The role of heme oxygenase-1 promoter polymorphisms in human disease. *Free Radic Biol Med* 2004 October 15;37(8):1097-104.
- (65) Hu JL, Li ZY, Liu W, Zhang RG, Li GL, Wang T, Ren JH, Wu G. Polymorphism in heme oxygenase-1 (HO-1) promoter and alcohol are related to the risk of esophageal squamous cell carcinoma on Chinese males. *Neoplasma* 2010;57(1):86-92.
- (66) Kikuchi A, Yamaya M, Suzuki S, Yasuda H, Kubo H, Nakayama K, Handa M, Sasaki T, Shibahara S, Sekizawa K, Sasaki H. Association of susceptibility to the development of lung adenocarcinoma with the heme oxygenase-1 gene promoter polymorphism. *Hum Genet* 2005 April;116(5):354-60.
- (67) Hong CC, Ambrosone CB, Ahn J, Choi JY, McCullough ML, Stevens VL, Rodriguez C, Thun MJ, Calle EE. Genetic variability in iron-related oxidative stress pathways (Nrf2, NQO1, NOS3, and HO-1), iron intake, and risk of postmenopausal breast cancer. *Cancer Epidemiol Biomarkers Prev* 2007 September;16(9):1784-94.
- (68) Chang KW, Lee TC, Yeh WI, Chung MY, Liu CJ, Chi LY, Lin SC. Polymorphism in heme oxygenase-1 (HO-1) promoter is related to the risk of oral squamous cell carcinoma occurring on male areca chewers. *Br J Cancer* 2004 October 18;91(8):1551-5.
- (69) Sawa T, Mounawar M, Tatemichi M, Gilibert I, Katoh T, Ohshima H. Increased risk of gastric cancer in Japanese subjects is associated with microsatellite polymorphisms in the heme oxygenase-1 and the inducible nitric oxide synthase gene promoters. *Cancer Lett* 2008 September 28;269(1):78-84.
- (70) Okamoto I, Krogler J, Endler G, Kaufmann S, Mustafa S, Exner M, Mannhalter C, Wagner O, Pehamberger H. A microsatellite polymorphism in the heme oxygenase-1 gene promoter is associated with risk for melanoma. *Int J Cancer* 2006 September 15;119(6):1312-5.
- (71) Brar SS, Kennedy TP, Quinn M, Hoidal JR. Redox signaling of NF-kappaB by membrane NAD(P)H oxidases in normal and malignant cells. *Protoplasma* 2003 May;221(1-2):117-27.
- (72) Wu WS. The signaling mechanism of ROS in tumor progression. *Cancer Metastasis Rev* 2006 December;25(4):695-705.
- (73) McCarty MF, Barroso-Aranda J, Contreras F. A two-phase strategy for treatment of oxidant-dependent cancers. *Med Hypotheses* 2007;69(3):489-96.
- (74) Rokutan K, Kawahara T, Kuwano Y, Tominaga K, Sekiyama A, Teshima-Kondo S. NADPH oxidases in the gastrointestinal tract: a potential role of Nox1 in innate immune response and carcinogenesis. *Antioxid Redox Signal* 2006 September;8(9-10):1573-82.
- (75) Kamata T. Roles of Nox1 and other Nox isoforms in cancer development. *Cancer Sci* 2009 August;100(8):1382-8.
- (76) Sawa T, Ohshima H. Nitrate DNA damage in inflammation and its possible role in carcinogenesis. *Nitric Oxide* 2006 March;14(2):91-100.

- (77) Ohshima H, Tatemichi M, Sawa T. Chemical basis of inflammation-induced carcinogenesis. *Arch Biochem Biophys* 2003 September 1;417(1):3-11.
- (78) Mello Filho AC, Meneghini R. In vivo formation of single-strand breaks in DNA by hydrogen peroxide is mediated by the Haber-Weiss reaction. *Biochim Biophys Acta* 1984 February 24;781(1-2):56-63.
- (79) Toyokuni S. Role of iron in carcinogenesis: Cancer as a ferrototoxic disease. *Cancer Sci* 2008 October 23.
- (80) Nakano M, Kawanishi Y, Kamohara S, Uchida Y, Shiota M, Inatomi Y, Komori T, Miyazawa K, Gondo K, Yamasawa I. Oxidative DNA damage (8-hydroxydeoxyguanosine) and body iron status: a study on 2507 healthy people. *Free Radic Biol Med* 2003 October 1;35(7):826-32.
- (81) Tuomainen TP, Loft S, Nyssonen K, Punnonen K, Salonen JT, Poulsen HE. Body iron is a contributor to oxidative damage of DNA. *Free Radic Res* 2007 March;41(3):324-8.
- (82) Hori A, Mizoue T, Kasai H, Kawai K, Matsushita Y, Nanri A, Sato M, Ohta M. Body iron store as a predictor of oxidative DNA damage in healthy men and women. *Cancer Sci* 2010 February;101(2):517-22.
- (83) Zacharski LR, Chow BK, Howes PS, Shamayeva G, Baron JA, Dalman RL, Malenka DJ, Ozaki CK, Lavori PW. Reduction of iron stores and cardiovascular outcomes in patients with peripheral arterial disease: a randomized controlled trial. *JAMA* 2007 February 14;297(6):603-10.
- (84) Alexander D, Ball MJ, Mann J. Nutrient intake and haematological status of vegetarians and age-sex matched omnivores. *Eur J Clin Nutr* 1994 August;48(8):538-46.
- (85) Fleming DJ, Tucker KL, Jacques PF, Dallal GE, Wilson PW, Wood RJ. Dietary factors associated with the risk of high iron stores in the elderly Framingham Heart Study cohort. *Am J Clin Nutr* 2002 December;76(6):1375-84.
- (86) Fujii M, Inoguchi T, Sasaki S, Maeda Y, Zheng J, Kobayashi K, Takayanagi R. Bilirubin and biliverdin protect rodents against diabetic nephropathy by downregulating NAD(P)H oxidase. *Kidney Int* 2010 November;78(9):905-19.
- (87) Terry MJ, Maines MD, Lagarias JC. Inactivation of phytochrome- and phycobiliprotein-chromophore precursors by rat liver biliverdin reductase. *J Biol Chem* 1993 December 15;268(35):26099-106.
- (88) McCarty MF. Clinical potential of Spirulina as a source of phycocyanobilin. *J Med Food* 2007 December;10(4):566-70.
- (89) Romay C, Gonzalez R, Ledon N, Ramirez D, Rimbau V. C-phycoyanin: a biliprotein with antioxidant, anti-inflammatory and neuroprotective effects. *Curr Protein Pept Sci* 2003 June;4(3):207-16.

- (90) Chamorro G, Perez-Albiter M, Serrano-Garcia N, Mares-Samano JJ, Rojas P. Spirulina maxima pretreatment partially protects against 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine neurotoxicity. *Nutr Neurosci* 2006 October;9(5-6):207-12.
- (91) Riss J, Decorde K, Sutra T, Delage M, Baccou JC, Jouy N, Brune JP, Oreal H, Cristol JP, Rouanet JM. Phycobiliprotein C-Phycocyanin from *Spirulina platensis* Is Powerfully Responsible for Reducing Oxidative Stress and NADPH Oxidase Expression Induced by an Atherogenic Diet in Hamsters. *J Agric Food Chem* 2007 September 19;55(19):7962-7.
- (92) Schwartz J, Shklar G, Reid S, Trickler D. Prevention of experimental oral cancer by extracts of *Spirulina-Dunaliella* algae. *Nutr Cancer* 1988;11(2):127-34.
- (93) Ismail MF, Ali DA, Fernando A, Abdraboh ME, Gaur RL, Ibrahim WM, Raj MH, Ouhtit A. Chemoprevention of rat liver toxicity and carcinogenesis by *Spirulina*. *Int J Biol Sci* 2009;5(4):377-87.
- (94) Shehzad A, Wahid F, Lee YS. Curcumin in cancer chemoprevention: molecular targets, pharmacokinetics, bioavailability, and clinical trials. *Arch Pharm (Weinheim)* 2010 September;343(9):489-99.
- (95) Park IJ, Lee YK, Hwang JT, Kwon DY, Ha J, Park OJ. Green tea catechin controls apoptosis in colon cancer cells by attenuation of H₂O₂-stimulated COX-2 expression via the AMPK signaling pathway at low-dose H₂O₂. *Ann N Y Acad Sci* 2009 August;1171:538-44.
- (96) Huang GS, Tseng CY, Lee CH, Su SL, Lee HS. Effects of (-)-epigallocatechin-3-gallate on cyclooxygenase 2, PGE(2), and IL-8 expression induced by IL-1beta in human synovial fibroblasts. *Rheumatol Int* 2010 July;30(9):1197-203.
- (97) Ha H, Lee JH, Kim HN, Kim HM, Kwak HB, Lee S, Kim HH, Lee ZH. alpha-Lipoic acid inhibits inflammatory bone resorption by suppressing prostaglandin E2 synthesis. *J Immunol* 2006 January 1;176(1):111-7.
- (98) El-Shitany NA, El-Masry SA, El-Ghareib MA, El-Desoky K. Thiocetic acid protects against carrageenan-induced acute inflammation in rats by reduction in oxidative stress, downregulation of COX-2 mRNA and enhancement of IL-10 mRNA. *Fundam Clin Pharmacol* 2010 February;24(1):91-9.
- (99) Talalay P, Fahey JW, Holtzclaw WD, Prestera T, Zhang Y. Chemoprotection against cancer by phase 2 enzyme induction. *Toxicol Lett* 1995 December;82-83:173-9.
- (100) Wakabayashi N, Dinkova-Kostova AT, Holtzclaw WD, Kang MI, Kobayashi A, Yamamoto M, Kensler TW, Talalay P. Protection against electrophile and oxidant stress by induction of the phase 2 response: fate of cysteines of the Keap1 sensor modified by inducers. *Proc Natl Acad Sci U S A* 2004 February 17;101(7):2040-5.
- (101) Rodriguez C, Mayo JC, Sainz RM, Antolin I, Herrera F, Martin V, Reiter RJ. Regulation of antioxidant enzymes: a significant role for melatonin. *J Pineal Res* 2004 January;36(1):1-9.
- (102) Cuzzocrea S, Costantino G, Mazzon E, Caputi AP. Regulation of prostaglandin production in carrageenan-induced pleurisy by melatonin. *J Pineal Res* 1999 August;27(1):9-14.

- (103) Dong WG, Mei Q, Yu JP, Xu JM, Xiang L, Xu Y. Effects of melatonin on the expression of iNOS and COX-2 in rat models of colitis. *World J Gastroenterol* 2003 June;9(6):1307-11.
- (104) Deng WG, Tang ST, Tseng HP, Wu KK. Melatonin suppresses macrophage cyclooxygenase-2 and inducible nitric oxide synthase expression by inhibiting p52 acetylation and binding. *Blood* 2006 July 15;108(2):518-24.
- (105) Orendas P, Kassayova M, Kajo K, Ahlers I, Kubatka P, Bojkova B, Pec M, Ahlersova E. Celecoxib and melatonin in prevention of female rat mammary carcinogenesis. *Neoplasma* 2009;56(3):252-8.
- (106) Martinez-Campa C, Gonzalez A, Mediavilla MD, onso-Gonzalez C, varez-Garcia V, Sanchez-Barcelo EJ, Cos S. Melatonin inhibits aromatase promoter expression by regulating cyclooxygenases expression and activity in breast cancer cells. *Br J Cancer* 2009 November 3;101(9):1613-9.
- (107) Padillo FJ, Ruiz-Rabelo JF, Cruz A, Perea MD, Tasset I, Montilla P, Tunez I, Muntane J. Melatonin and celecoxib improve the outcomes in hamsters with experimental pancreatic cancer. *J Pineal Res* 2010 October;49(3):264-70.
- (108) Schernhammer ES, Schulmeister K. Melatonin and cancer risk: does light at night compromise physiologic cancer protection by lowering serum melatonin levels? *Br J Cancer* 2004 March 8;90(5):941-3.
- (109) Schernhammer ES, Hankinson SE. Urinary melatonin levels and breast cancer risk. *J Natl Cancer Inst* 2005 July 20;97(14):1084-7.
- (110) Megdal SP, Kroenke CH, Laden F, Pukkala E, Schernhammer ES. Night work and breast cancer risk: a systematic review and meta-analysis. *Eur J Cancer* 2005 September;41(13):2023-32.
- (111) Viswanathan AN, Hankinson SE, Schernhammer ES. Night shift work and the risk of endometrial cancer. *Cancer Res* 2007 November 1;67(21):10618-22.
- (112) Blask DE. Melatonin, sleep disturbance and cancer risk. *Sleep Med Rev* 2009 August;13(4):257-64.
- (113) Atkuri KR, Mantovani JJ, Herzenberg LA, Herzenberg LA. N-Acetylcysteine--a safe antidote for cysteine/glutathione deficiency. *Curr Opin Pharmacol* 2007 August;7(4):355-9.
- (114) Dodd S, Dean O, Copolov DL, Malhi GS, Berk M. N-acetylcysteine for antioxidant therapy: pharmacology and clinical utility. *Expert Opin Biol Ther* 2008 December;8(12):1955-62.
- (115) Estensen RD, Levy M, Klopp SJ, Galbraith AR, Mandel JS, Blomquist JA, Wattenberg LW. N-acetylcysteine suppression of the proliferative index in the colon of patients with previous adenomatous colonic polyps. *Cancer Lett* 1999 December 1;147(1-2):109-14.
- (116) De FS, Izzotti A, D'Agostini F, Balansky RM. Mechanisms of N-acetylcysteine in the prevention of DNA damage and cancer, with special reference to smoking-related end-points. *Carcinogenesis* 2001 July;22(7):999-1013.

- (117) Cotter MA, Thomas J, Cassidy P, Robinette K, Jenkins N, Florell SR, Leachman S, Samlowski WE, Grossman D. N-acetylcysteine protects melanocytes against oxidative stress/damage and delays onset of ultraviolet-induced melanoma in mice. *Clin Cancer Res* 2007 October 1;13(19):5952-8.
- (118) Balansky R, Ganchev G, Ilcheva M, Steele VE, De FS. Prevention of cigarette smoke-induced lung tumors in mice by budesonide, phenethyl isothiocyanate, and N-acetylcysteine. *Int J Cancer* 2010 March 1;126(5):1047-54.
- (119) Levine M, Conry-Cantilena C, Wang Y, Welch RW, Washko PW, Dhariwal KR, Park JB, Lazarev A, Graumlich JF, King J, Cantilena LR. Vitamin C pharmacokinetics in healthy volunteers: evidence for a recommended dietary allowance. *Proc Natl Acad Sci U S A* 1996 April 16;93(8):3704-9.
- (120) Yuan JP, Peng J, Yin K, Wang JH. Potential health-promoting effects of astaxanthin: A high-value carotenoid mostly from microalgae. *Mol Nutr Food Res* 2010 November 18.
- (121) Manabe E, Handa O, Naito Y, Mizushima K, Akagiri S, Adachi S, Takagi T, Kokura S, Maoka T, Yoshikawa T. Astaxanthin protects mesangial cells from hyperglycemia-induced oxidative signaling. *J Cell Biochem* 2008 April 15;103(6):1925-37.
- (122) Liu X, Shibata T, Hisaka S, Osawa T. Astaxanthin inhibits reactive oxygen species-mediated cellular toxicity in dopaminergic SH-SY5Y cells via mitochondria-targeted protective mechanism. *Brain Res* 2009 February 13;1254:18-27.
- (123) Wolf AM, Asoh S, Hiranuma H, Ohsawa I, Iio K, Satou A, Ishikura M, Ohta S. Astaxanthin protects mitochondrial redox state and functional integrity against oxidative stress. *J Nutr Biochem* 2010 May;21(5):381-9.
- (124) Nagendrababhu P, Sudhandiran G. Astaxanthin inhibits tumor invasion by decreasing extracellular matrix production and induces apoptosis in experimental rat colon carcinogenesis by modulating the expressions of ERK-2, NFkB and COX-2. *Invest New Drugs* 2009 October 30.
- (125) Choi SK, Park YS, Choi DK, Chang HI. Effects of astaxanthin on the production of NO and the expression of COX-2 and iNOS in LPS-stimulated BV2 microglial cells. *J Microbiol Biotechnol* 2008 December;18(12):1990-6.
- (126) Kim YJ, Kim YA, Yokozawa T. Protection against oxidative stress, inflammation, and apoptosis of high-glucose-exposed proximal tubular epithelial cells by astaxanthin. *J Agric Food Chem* 2009 October 14;57(19):8793-7.
- (127) Park JS, Chyun JH, Kim YK, Line LL, Chew BP. Astaxanthin decreased oxidative stress and inflammation and enhanced immune response in humans. *Nutr Metab (Lond)* 2010;7:18.
- (128) Lippman SM, Klein EA, Goodman PJ, Lucia MS, Thompson IM, Ford LG, Parnes HL, Minasian LM, Gaziano JM, Hartline JA, Parsons JK, Bearden JD, III, Crawford ED, Goodman GE, Claudio J, Winquist E, Cook ED, Karp DD, Walther P, Lieber MM, Kristal AR, Darke AK, Arnold KB, Ganz PA, Santella RM, Albanes D, Taylor PR, Probstfield JL, Jagpal TJ, Crowley JJ, Meyskens FL, Jr., Baker LH, Coltman CA, Jr. Effect of selenium and vitamin E on

risk of prostate cancer and other cancers: the Selenium and Vitamin E Cancer Prevention Trial (SELECT). *JAMA* 2009 January 7;301(1):39-51.

- (129) Rayman MP. Selenoproteins and human health: insights from epidemiological data. *Biochim Biophys Acta* 2009 November;1790(11):1533-40.
- (130) Flores-Mateo G, Navas-Acien A, Pastor-Barriuso R, Guallar E. Selenium and coronary heart disease: a meta-analysis. *Am J Clin Nutr* 2006 October;84(4):762-73.
- (131) Rezk BM, Haenen GR, Van d, V, Bast A. Tetrahydrofolate and 5-methyltetrahydrofolate are folates with high antioxidant activity. Identification of the antioxidant pharmacophore. *FEBS Lett* 2003 December 18;555(3):601-5.
- (132) Antoniadou C, Shirodaria C, Warrick N, Cai S, de BJ, Lee J, Leeson P, Neubauer S, Ratnatunga C, Pillai R, Refsum H, Channon KM. 5-methyltetrahydrofolate rapidly improves endothelial function and decreases superoxide production in human vessels: effects on vascular tetrahydrobiopterin availability and endothelial nitric oxide synthase coupling. *Circulation* 2006 September 12;114(11):1193-201.
- (133) McCarty MF, Barroso-Aranda J, Contreras F. High-dose folate and dietary purines promote scavenging of peroxynitrite-derived radicals - clinical potential in inflammatory disorders. *Medical Hypotheses* 2009;accepted for publication.
- (134) Larsson SC, Giovannucci E, Wolk A. Folate and risk of breast cancer: a meta-analysis. *J Natl Cancer Inst* 2007 January 3;99(1):64-76.
- (135) Giovannucci E, Rimm EB, Ascherio A, Stampfer MJ, Colditz GA, Willett WC. Alcohol, low-methionine--low-folate diets, and risk of colon cancer in men. *J Natl Cancer Inst* 1995 February 15;87(4):265-73.
- (136) Guo WD, Li JY, Blot WJ, Hsing AW, Chen JS, Fraumeni JF, Jr. Correlations of dietary intake and blood nutrient levels with esophageal cancer mortality in China. *Nutr Cancer* 1990;13(3):121-7.
- (137) Bollschweiler E, Wolfgarten E, Nowroth T, Rosendahl U, Monig SP, Holscher AH. Vitamin intake and risk of subtypes of esophageal cancer in Germany. *J Cancer Res Clin Oncol* 2002 October;128(10):575-80.
- (138) Rossi M, Garavello W, Talamini R, La VC, Franceschi S, Lagioui P, Zambon P, Dal ML, Bosetti C, Negri E. Flavonoids and risk of squamous cell esophageal cancer. *Int J Cancer* 2007 April 1;120(7):1560-4.
- (139) Brown LM, Swanson CA, Gridley G, Swanson GM, Silverman DT, Greenberg RS, Hayes RB, Schoenberg JB, Pottern LM, Schwartz AG, Liff JM, Hoover R, Fraumeni JF, Jr. Dietary factors and the risk of squamous cell esophageal cancer among black and white men in the United States. *Cancer Causes Control* 1998 October;9(5):467-74.
- (140) Franceschi S, Bidoli E, Negri E, Zambon P, Talamini R, Ruol A, Parpinel M, Levi F, Simonato L, La VC. Role of macronutrients, vitamins and minerals in the aetiology of squamous-cell carcinoma of the oesophagus. *Int J Cancer* 2000 June 1;86(5):626-31.

- (141) Steevens J, van den Brandt PA, Goldbohm RA, Schouten LJ. Selenium status and the risk of esophageal and gastric cancer subtypes: the Netherlands cohort study. *Gastroenterology* 2010 May;138(5):1704-13.
- (142) Lu H, Cai L, Mu LN, Lu QY, Zhao J, Cui Y, Sul JH, Zhou XF, Ding BG, Elashoff RM, Marshall J, Yu SZ, Jiang QW, Zhang ZF. Dietary mineral and trace element intake and squamous cell carcinoma of the esophagus in a Chinese population. *Nutr Cancer* 2006;55(1):63-70.
- (143) Terry P, Lagergren J, Ye W, Nyren O, Wolk A. Antioxidants and cancers of the esophagus and gastric cardia. *Int J Cancer* 2000 September 1;87(5):750-4.
- (144) Stoner GD, Wang LS, Chen T. Chemoprevention of esophageal squamous cell carcinoma. *Toxicol Appl Pharmacol* 2007 November 1;224(3):337-49.
- (145) Garland CF, Garland FC, Gorham ED, Lipkin M, Newmark H, Mohr SB, Holick MF. The role of vitamin D in cancer prevention. *Am J Public Health* 2006 February;96(2):252-61.
- (146) Grant WB, Garland CF, Gorham ED. An estimate of cancer mortality rate reductions in Europe and the US with 1,000 IU of oral vitamin D per day. *Recent Results Cancer Res* 2007;174:225-34.
- (147) Grant WB. Ecologic studies of solar UV-B radiation and cancer mortality rates. *Recent Results Cancer Res* 2003;164:371-7.
- (148) Gorham ED, Garland CF, Garland FC, Grant WB, Mohr SB, Lipkin M, Newmark HL, Giovannucci E, Wei M, Holick MF. Optimal vitamin D status for colorectal cancer prevention: a quantitative meta analysis. *Am J Prev Med* 2007 March;32(3):210-6.
- (149) Grant WB. How strong is the evidence that solar ultraviolet B and vitamin D reduce the risk of cancer?: An examination using Hill's criteria for causality. *Dermatoendocrinol* 2009 January;1(1):17-24.
- (150) Grant WB. An estimate of premature cancer mortality in the U.S. due to inadequate doses of solar ultraviolet-B radiation. *Cancer* 2002 March 15;94(6):1867-75.
- (151) Freedman DM, Looker AC, Chang SC, Graubard BI. Prospective study of serum vitamin D and cancer mortality in the United States. *J Natl Cancer Inst* 2007 November 7;99(21):1594-602.
- (152) Schwartz GG, Whitlatch LW, Chen TC, Lokeshwar BL, Holick MF. Human prostate cells synthesize 1,25-dihydroxyvitamin D3 from 25-hydroxyvitamin D3. *Cancer Epidemiol Biomarkers Prev* 1998 May;7(5):391-5.
- (153) Townsend K, Evans KN, Campbell MJ, Colston KW, Adams JS, Hewison M. Biological actions of extra-renal 25-hydroxyvitamin D-1alpha-hydroxylase and implications for chemoprevention and treatment. *J Steroid Biochem Mol Biol* 2005 October;97(1-2):103-9.
- (154) Moreno J, Krishnan AV, Swami S, Nonn L, Peehl DM, Feldman D. Regulation of prostaglandin metabolism by calcitriol attenuates growth stimulation in prostate cancer cells. *Cancer Res* 2005 September 1;65(17):7917-25.

- (155) Fichera A, Little N, Dougherty U, Mustafi R, Cerda S, Li YC, Delgado J, Arora A, Campbell LK, Joseph L, Hart J, Noffsinger A, Bissonnette M. A vitamin D analogue inhibits colonic carcinogenesis in the AOM/DSS model. *J Surg Res* 2007 October;142(2):239-45.
- (156) Aparna R, Subhashini J, Roy KR, Reddy GS, Robinson M, Uskokovic MR, Venkateswara RG, Reddanna P. Selective inhibition of cyclooxygenase-2 (COX-2) by 1alpha,25-dihydroxy-16-ene-23-yne-vitamin D3, a less calcemic vitamin D analog. *J Cell Biochem* 2008 August 1;104(5):1832-42.
- (157) Krishnan AV, Swami S, Feldman D. Vitamin D and breast cancer: inhibition of estrogen synthesis and signaling. *J Steroid Biochem Mol Biol* 2010 July;121(1-2):343-8.
- (158) Krishnan AV, Moreno J, Nonn L, Swami S, Peehl DM, Feldman D. Calcitriol as a chemopreventive and therapeutic agent in prostate cancer: role of anti-inflammatory activity. *J Bone Miner Res* 2007 December;22 Suppl 2:V74-V80.
- (159) Yu XP, Bellido T, Manolagas SC. Down-regulation of NF-kappa B protein levels in activated human lymphocytes by 1,25-dihydroxyvitamin D3. *Proc Natl Acad Sci U S A* 1995 November 21;92(24):10990-4.
- (160) Harant H, Andrew PJ, Reddy GS, Foglar E, Lindley IJ. 1alpha,25-dihydroxyvitamin D3 and a variety of its natural metabolites transcriptionally repress nuclear-factor-kappaB-mediated interleukin-8 gene expression. *Eur J Biochem* 1997 November 15;250(1):63-71.
- (161) Riis JL, Johansen C, Gesser B, Moller K, Larsen CG, Kragballe K, Iversen L. 1alpha,25(OH)(2)D(3) regulates NF-kappaB DNA binding activity in cultured normal human keratinocytes through an increase in IkappaBalpha expression. *Arch Dermatol Res* 2004 October;296(5):195-202.
- (162) Cohen-Lahav M, Shany S, Tobvin D, Chaimovitz C, Douvdevani A. Vitamin D decreases NFkappaB activity by increasing IkappaBalpha levels. *Nephrol Dial Transplant* 2006 April;21(4):889-97.
- (163) Bao BY, Yao J, Lee YF. 1alpha, 25-dihydroxyvitamin D3 suppresses interleukin-8-mediated prostate cancer cell angiogenesis. *Carcinogenesis* 2006 September;27(9):1883-93.
- (164) Zhang Z, Yuan W, Sun L, Szeto FL, Wong KE, Li X, Kong J, Li YC. 1,25-Dihydroxyvitamin D3 targeting of NF-kappaB suppresses high glucose-induced MCP-1 expression in mesangial cells. *Kidney Int* 2007 July;72(2):193-201.
- (165) Tan X, Wen X, Liu Y. Paricalcitol inhibits renal inflammation by promoting vitamin D receptor-mediated sequestration of NF-kappaB signaling. *J Am Soc Nephrol* 2008 September;19(9):1741-52.
- (166) Talmor Y, Bernheim J, Klein O, Green J, Rashid G. Calcitriol blunts pro-atherosclerotic parameters through NFkappaB and p38 in vitro. *Eur J Clin Invest* 2008 August;38(8):548-54.
- (167) Deb DK, Chen Y, Zhang Z, Zhang Y, Szeto FL, Wong KE, Kong J, Li YC. 1,25-Dihydroxyvitamin D3 suppresses high glucose-induced angiotensinogen expression in kidney

cells by blocking the NF- κ B pathway. *Am J Physiol Renal Physiol* 2009 May;296(5):F1212-F1218.

- (168) Mineva ND, Wang X, Yang S, Ying H, Xiao ZX, Holick MF, Sonenshein GE. Inhibition of RelB by 1,25-dihydroxyvitamin D3 promotes sensitivity of breast cancer cells to radiation. *J Cell Physiol* 2009 September;220(3):593-9.
- (169) Krishnan AV, Feldman D. Molecular pathways mediating the anti-inflammatory effects of calcitriol: implications for prostate cancer chemoprevention and treatment. *Endocr Relat Cancer* 2010 March;17(1):R19-R38.
- (170) Tse AK, Zhu GY, Wan CK, Shen XL, Yu ZL, Fong WF. 1 α ,25-Dihydroxyvitamin D3 inhibits transcriptional potential of nuclear factor kappa B in breast cancer cells. *Mol Immunol* 2010 May;47(9):1728-38.
- (171) Kwon HJ, Won YS, Suh HW, Jeon JH, Shao Y, Yoon SR, Chung JW, Kim TD, Kim HM, Nam KH, Yoon WK, Kim DG, Kim JH, Kim YS, Kim DY, Kim HC, Choi I. Vitamin D3 upregulated protein 1 suppresses TNF- α -induced NF- κ B activation in hepatocarcinogenesis. *J Immunol* 2010 October 1;185(7):3980-9.
- (172) Adams LS, Teegarden D. 1,25-dihydroxycholecalciferol inhibits apoptosis in C3H10T1/2 murine fibroblast cells through activation of nuclear factor kappaB. *J Nutr* 2004 November;134(11):2948-52.
- (173) Moriwake T, Tanaka H, Kanzaki S, Higuchi J, Seino Y. 1,25-Dihydroxyvitamin D3 stimulates the secretion of insulin-like growth factor binding protein 3 (IGFBP-3) by cultured human osteosarcoma cells. *Endocrinology* 1992 February;130(2):1071-3.
- (174) Colston KW, Perks CM, Xie SP, Holly JM. Growth inhibition of both MCF-7 and Hs578T human breast cancer cell lines by vitamin D analogues is associated with increased expression of insulin-like growth factor binding protein-3. *J Mol Endocrinol* 1998 February;20(1):157-62.
- (175) Huynh H, Pollak M, Zhang JC. Regulation of insulin-like growth factor (IGF) II and IGF binding protein 3 autocrine loop in human PC-3 prostate cancer cells by vitamin D metabolite 1,25(OH)₂D₃ and its analog EB1089. *Int J Oncol* 1998 July;13(1):137-43.
- (176) Sprenger CC, Peterson A, Lance R, Ware JL, Drivdahl RH, Plymate SR. Regulation of proliferation of prostate epithelial cells by 1,25-dihydroxyvitamin D3 is accompanied by an increase in insulin-like growth factor binding protein-3. *J Endocrinol* 2001 September;170(3):609-18.
- (177) Stewart LV, Weigel NL. Role of insulin-like growth factor binding proteins in 1 α ,25-dihydroxyvitamin D(3)-induced growth inhibition of human prostate cancer cells. *Prostate* 2005 June 15;64(1):9-19.
- (178) Matilainen M, Malinen M, Saavalainen K, Carlberg C. Regulation of multiple insulin-like growth factor binding protein genes by 1 α ,25-dihydroxyvitamin D3. *Nucleic Acids Res* 2005;33(17):5521-32.

- (179) Malinen M, Ryyanen J, Heinaniemi M, Vaisanen S, Carlberg C. Cyclical regulation of the insulin-like growth factor binding protein 3 gene in response to 1 α ,25-dihydroxyvitamin D₃. *Nucleic Acids Res* 2010 September 19.
- (180) Jogie-Brahim S, Feldman D, Oh Y. Unraveling insulin-like growth factor binding protein-3 actions in human disease. *Endocr Rev* 2009 August;30(5):417-37.
- (181) Massoner P, Colleselli D, Matscheski A, Pircher H, Geley S, Jansen DP, Klocker H. Novel mechanism of IGF-binding protein-3 action on prostate cancer cells: inhibition of proliferation, adhesion, and motility. *Endocr Relat Cancer* 2009 September;16(3):795-808.
- (182) An BS, Tavera-Mendoza LE, Dimitrov V, Wang X, Calderon MR, Wang HJ, White JH. Stimulation of Sirt1-regulated FoxO protein function by the ligand-bound vitamin D receptor. *Mol Cell Biol* 2010 October;30(20):4890-900.
- (183) Palmer HG, Gonzalez-Sancho JM, Espada J, Berciano MT, Puig I, Baulida J, Quintanilla M, Cano A, de Herreros AG, Lafarga M, Munoz A. Vitamin D(3) promotes the differentiation of colon carcinoma cells by the induction of E-cadherin and the inhibition of beta-catenin signaling. *J Cell Biol* 2001 July 23;154(2):369-87.
- (184) Wang Q, Lee D, Sysounthone V, Chandraratna RAS, Christakos S, Korah R, Wieder R. 1,25-dihydroxyvitamin D₃ and retinoic acid analogues induce differentiation in breast cancer cells with function- and cell-specific additive effects. *Breast Cancer Res Treat* 2001 May;67(2):157-68.
- (185) Xu H, McCann M, Zhang Z, Posner GH, Bingham V, El-Tanani M, Campbell FC. Vitamin D receptor modulates the neoplastic phenotype through antagonistic growth regulatory signals. *Mol Carcinog* 2009 August;48(8):758-72.
- (186) Garland CF, French CB, Baggerly LL, Heaney RP. Vitamin d supplement doses and serum 25-hydroxyvitamin d in the range associated with cancer prevention. *Anticancer Res* 2011 February;31(2):607-11.
- (187) Vieth R. Vitamin D supplementation, 25-hydroxyvitamin D concentrations, and safety. *Am J Clin Nutr* 1999 May;69(5):842-56.
- (188) Vieth R. Vitamin D and cancer mini-symposium: the risk of additional vitamin D. *Ann Epidemiol* 2009 July;19(7):441-5.
- (189) Funahashi H, Satake M, Hasan S, Sawai H, Newman RA, Reber HA, Hines OJ, Eibl G. Opposing effects of n-6 and n-3 polyunsaturated fatty acids on pancreatic cancer growth. *Pancreas* 2008 May;36(4):353-62.
- (190) Maillard V, Bougnoux P, Ferrari P, Jourdan ML, Pinault M, Lavillonniere F, Body G, Le FO, Chajes V. N-3 and N-6 fatty acids in breast adipose tissue and relative risk of breast cancer in a case-control study in Tours, France. *Int J Cancer* 2002 March 1;98(1):78-83.
- (191) Bagga D, Anders KH, Wang HJ, Glaspy JA. Long-chain n-3-to-n-6 polyunsaturated fatty acid ratios in breast adipose tissue from women with and without breast cancer. *Nutr Cancer* 2002;42(2):180-5.

- (192) Goodstine SL, Zheng T, Holford TR, Ward BA, Carter D, Owens PH, Mayne ST. Dietary (n-3)/(n-6) fatty acid ratio: possible relationship to premenopausal but not postmenopausal breast cancer risk in U.S. women. *J Nutr* 2003 May;133(5):1409-14.
- (193) Chajes V, Bougnoux P. Omega-6/omega-3 polyunsaturated fatty acid ratio and cancer. *World Rev Nutr Diet* 2003;92:133-51.
- (194) Leitzmann MF, Stampfer MJ, Michaud DS, Augustsson K, Colditz GC, Willett WC, Giovannucci EL. Dietary intake of n-3 and n-6 fatty acids and the risk of prostate cancer. *Am J Clin Nutr* 2004 July;80(1):204-16.
- (195) Kojima M, Wakai K, Tokudome S, Suzuki K, Tamakoshi K, Watanabe Y, Kawado M, Hashimoto S, Hayakawa N, Ozasa K, Toyoshima H, Suzuki S, Ito Y, Tamakoshi A. Serum levels of polyunsaturated fatty acids and risk of colorectal cancer: a prospective study. *Am J Epidemiol* 2005 March 1;161(5):462-71.
- (196) Oh K, Willett WC, Fuchs CS, Giovannucci E. Dietary marine n-3 fatty acids in relation to risk of distal colorectal adenoma in women. *Cancer Epidemiol Biomarkers Prev* 2005 April;14(4):835-41.
- (197) Ritch CR, Wan RL, Stephens LB, Taxy JB, Huo D, Gong EM, Zagaja GP, Brendler CB. Dietary fatty acids correlate with prostate cancer biopsy grade and volume in Jamaican men. *J Urol* 2007 January;177(1):97-101.
- (198) Thiebaut AC, Chajes V, Gerber M, Boutron-Ruault MC, Joulin V, Lenoir G, Berrino F, Riboli E, Benichou J, Clavel-Chapelon F. Dietary intakes of omega-6 and omega-3 polyunsaturated fatty acids and the risk of breast cancer. *Int J Cancer* 2009 February 15;124(4):924-31.
- (199) Daniel CR, McCullough ML, Patel RC, Jacobs EJ, Flanders WD, Thun MJ, Calle EE. Dietary intake of omega-6 and omega-3 fatty acids and risk of colorectal cancer in a prospective cohort of U.S. men and women. *Cancer Epidemiol Biomarkers Prev* 2009 February;18(2):516-25.
- (200) Kim S, Sandler DP, Galanko J, Martin C, Sandler RS. Intake of polyunsaturated fatty acids and distal large bowel cancer risk in whites and African Americans. *Am J Epidemiol* 2010 May 1;171(9):969-79.
- (201) Roebuck BD. Dietary fat and the development of pancreatic cancer. *Lipids* 1992 October;27(10):804-6.
- (202) Noguchi M, Minami M, Yagasaki R, Kinoshita K, Earashi M, Kitagawa H, Taniya T, Miyazaki I. Chemoprevention of DMBA-induced mammary carcinogenesis in rats by low-dose EPA and DHA. *Br J Cancer* 1997;75(3):348-53.
- (203) Singh J, Hamid R, Reddy BS. Dietary fish oil inhibits the expression of farnesyl protein transferase and colon tumor development in rodents. *Carcinogenesis* 1998 June;19(6):985-9.
- (204) Hong MY, Chapkin RS, Davidson LA, Turner ND, Morris JS, Carroll RJ, Lupton JR. Fish oil enhances targeted apoptosis during colon tumor initiation in part by downregulating Bcl-2. *Nutr Cancer* 2003;46(1):44-51.

- (205) Kimura Y. [Fish, n-3 polyunsaturated fatty acid and colorectal cancer prevention: a review of experimental and epidemiological studies]. *Nippon Koshu Eisei Zasshi* 2006 October;53(10):735-48.
- (206) van B, V, Spengelink B, Mooibroek H, Sijtsma L, Bosch D, Rietjens IM, Alink GM. An n-3 PUFA-rich microalgal oil diet protects to a similar extent as a fish oil-rich diet against AOM-induced colonic aberrant crypt foci in F344 rats. *Food Chem Toxicol* 2009 February;47(2):316-20.
- (207) Moreira AP, Sabarense CM, Dias CM, Lunz W, Natali AJ, Gloria MB, Peluzio MC. Fish oil ingestion reduces the number of aberrant crypt foci and adenoma in 1,2-dimethylhydrazine-induced colon cancer in rats. *Braz J Med Biol Res* 2009 December;42(12):1167-72.
- (208) Sarotra P, Sharma G, Kansal S, Negi AK, Aggarwal R, Sandhir R, Agnihotri N. Chemopreventive effect of different ratios of fish oil and corn oil in experimental colon carcinogenesis. *Lipids* 2010 September;45(9):785-98.
- (209) Bartram HP, Gostner A, Reddy BS, Rao CV, Scheppach W, Dusel G, Richter A, Richter F, Kasper H. Missing anti-proliferative effect of fish oil on rectal epithelium in healthy volunteers consuming a high-fat diet: potential role of the n-3:n-6 fatty acid ratio. *Eur J Cancer Prev* 1995 June;4(3):231-7.
- (210) Leitzmann MF, Stampfer MJ, Michaud DS, Augustsson K, Colditz GC, Willett WC, Giovannucci EL. Dietary intake of n-3 and n-6 fatty acids and the risk of prostate cancer. *Am J Clin Nutr* 2004 July;80(1):204-16.
- (211) Simon JA, Chen YH, Bent S. The relation of alpha-linolenic acid to the risk of prostate cancer: a systematic review and meta-analysis. *Am J Clin Nutr* 2009 May;89(5):1558S-64S.
- (212) Anderson BM, Ma DW. Are all n-3 polyunsaturated fatty acids created equal? *Lipids Health Dis* 2009;8:33.
- (213) Harris WS, DiRienzo MA, Sands SA, George C, Jones PG, Eapen AK. Stearidonic acid increases the red blood cell and heart eicosapentaenoic acid content in dogs. *Lipids* 2007 April;42(4):325-33.
- (214) Eckert H, La VB, Schweiger BJ, Kinney AJ, Cahoon EB, Clemente T. Co-expression of the borage Delta 6 desaturase and the Arabidopsis Delta 15 desaturase results in high accumulation of stearidonic acid in the seeds of transgenic soybean. *Planta* 2006 October;224(5):1050-7.
- (215) Harris WS, Lemke SL, Hansen SN, Goldstein DA, DiRienzo MA, Su H, Nemeth MA, Taylor ML, Ahmed G, George C. Stearidonic acid-enriched soybean oil increased the omega-3 index, an emerging cardiovascular risk marker. *Lipids* 2008 September;43(9):805-11.
- (216) Kelavkar UP, Hutzley J, Dhir R, Kim P, Allen KG, McHugh K. Prostate tumor growth and recurrence can be modulated by the omega-6:omega-3 ratio in diet: athymic mouse xenograft model simulating radical prostatectomy. *Neoplasia* 2006 February;8(2):112-24.
- (217) McCarty MF. Insulin and IGF-I as determinants of low "Western" cancer rates in the rural third world. *Int J Epidemiol* 2004 August;33(4):908-10.

- (218) Barnard RJ. Prevention of Cancer Through Lifestyle Changes. *Evid Based Complement Alternat Med* 2004 December;1(3):233-9.
- (219) Giovannucci E. Insulin, insulin-like growth factors and colon cancer: a review of the evidence. *J Nutr* 2001 November;131(11 Suppl):3109S-20S.
- (220) Kari FW, Dunn SE, French JE, Barrett JC. Roles for insulin-like growth factor-1 in mediating the anti-carcinogenic effects of caloric restriction. *J Nutr Health Aging* 1999;3(2):92-101.
- (221) Campbell TC. Dietary protein, growth factors, and cancer. *Am J Clin Nutr* 2007 June;85(6):1667.
- (222) Campbell TC, Campbell TM. *The China study : The most comprehensive study of nutrition ever conducted and the startling Implications for diet, weight Loss and long-term health*. Benbella Books; 2006.
- (223) Fontana L, Weiss EP, Villareal DT, Klein S, Holloszy JO. Long-term effects of calorie or protein restriction on serum IGF-1 and IGFBP-3 concentration in humans. *Aging Cell* 2008 October;7(5):681-7.
- (224) Allen NE, Appleby PN, Davey GK, Kaaks R, Rinaldi S, Key TJ. The associations of diet with serum insulin-like growth factor I and its main binding proteins in 292 women meat-eaters, vegetarians, and vegans. *Cancer Epidemiol Biomarkers Prev* 2002 November;11(11):1441-8.
- (225) McCarty MF. mTOR activity as a determinant of cancer risk - rationalizing the cancer-preventive effects of adiponectin, metformin, rapamycin, and low-protein vegan diets. *Medical Hypotheses* 2011;in submission.
- (226) Campbell TC. Dietary protein, growth factors, and cancer. *Am J Clin Nutr* 2007 June;85(6):1667.
- (227) Kops GJ, Dansen TB, Polderman PE, Saarloos I, Wirtz KW, Coffey PJ, Huang TT, Bos JL, Medema RH, Burgering BM. Forkhead transcription factor FOXO3a protects quiescent cells from oxidative stress. *Nature* 2002 September 19;419(6904):316-21.
- (228) Chiribau CB, Cheng L, Cucoranu IC, Yu YS, Clempus RE, Sorescu D. FOXO3A regulates peroxiredoxin III expression in human cardiac fibroblasts. *J Biol Chem* 2008 March 28;283(13):8211-7.
- (229) Li M, Chiu JF, Gagne J, Fukagawa NK. Age-related differences in insulin-like growth factor-1 receptor signaling regulates Akt/FOXO3a and ERK/Fos pathways in vascular smooth muscle cells. *J Cell Physiol* 2008 November;217(2):377-87.
- (230) Tan WQ, Wang K, Lv DY, Li PF. Foxo3a inhibits cardiomyocyte hypertrophy through transactivating catalase. *J Biol Chem* 2008 October 31;283(44):29730-9.
- (231) Olmos Y, Valle I, Borniquel S, Tierrez A, Soria E, Lamas S, Monsalve M. Mutual dependence of Foxo3a and PGC-1alpha in the induction of oxidative stress genes. *J Biol Chem* 2009 May 22;284(21):14476-84.

- (232) Song G, Ouyang G, Bao S. The activation of Akt/PKB signaling pathway and cell survival. *J Cell Mol Med* 2005 January;9(1):59-71.
- (233) Guevara-Aguirre J, Balasubramanian P, Guevara-Aguirre M, Wei M, Madia F, Cheng CW, Hwang D, Martin-Montalvo A, Saavedra J, Ingles S, de CR, Cohen P, Longo VD. Growth hormone receptor deficiency is associated with a major reduction in pro-aging signaling, cancer, and diabetes in humans. *Sci Transl Med* 2011 February 16;3(70):70ra13.
- (234) Steuerman R, Shevah O, Laron Z. Congenital IGF-I deficiency tends to confer protection against post-natal development of malignancies. *Eur J Endocrinol* 2011 February 3.
- (235) Fontana L, Klein S, Holloszy JO. Long-term low-protein, low-calorie diet and endurance exercise modulate metabolic factors associated with cancer risk. *Am J Clin Nutr* 2006 December;84(6):1456-62.
- (236) Sanchez A, Kissinger DG, Phillips RI. A hypothesis on the etiological role of diet on age of menarche. *Med Hypotheses* 1981 November;7(11):1339-45.
- (237) Stoll BA. Western diet, early puberty, and breast cancer risk. *Breast Cancer Res Treat* 1998 June;49(3):187-93.
- (238) Wilson ME. Premature elevation in serum insulin-like growth factor-I advances first ovulation in rhesus monkeys. *J Endocrinol* 1998 August;158(2):247-57.
- (239) Key TJ, Chen J, Wang DY, Pike MC, Boreham J. Sex hormones in women in rural China and in Britain. *Br J Cancer* 1990 October;62(4):631-6.
- (240) Behl R, Kaul R. Insulin like growth factor 1 and regulation of ovarian function in mammals. *Indian J Exp Biol* 2002 January;40(1):25-30.
- (241) Villavicencio A, Iniguez G, Johnson MC, Gabler F, Palomino A, Vega M. Regulation of steroid synthesis and apoptosis by insulin-like growth factor I and insulin-like growth factor binding protein 3 in human corpus luteum during the midluteal phase. *Reproduction* 2002 October;124(4):501-8.
- (242) Denner L, Bodenbun YH, Jiang J, Pages G, Urban RJ. Insulin-like growth factor-I activates extracellularly regulated kinase to regulate the p450 side-chain cleavage insulin-like response element in granulosa cells. *Endocrinology* 2010 June;151(6):2819-25.
- (243) McCarty MF. Dietary saturate/unsaturate ratio as a determinant of adiposity. *Med Hypotheses* 2010 July;75(1):14-6.
- (244) Wynder EL, Fujita Y, Harris RE, Hirayama T, Hiyama T. Comparative epidemiology of cancer between the United States and Japan. A second look. *Cancer* 1991 February 1;67(3):746-63.
- (245) Willcox BJ, Willcox DC, Todoriki H, Fujiyoshi A, Yano K, He Q, Curb JD, Suzuki M. Caloric restriction, the traditional Okinawan diet, and healthy aging: the diet of the world's longest-lived people and its potential impact on morbidity and life span. *Ann N Y Acad Sci* 2007 October;1114:434-55.

- (246) Hebert JR, Hurley TG, Olendzki BC, Teas J, Ma Y, Hampl JS. Nutritional and socioeconomic factors in relation to prostate cancer mortality: a cross-national study [see comments]. *J Natl Cancer Inst* 1998 November 4;90(21):1637-47.
- (247) Carroll KK. Experimental evidence of dietary factors and hormone-dependent cancers. *Cancer Res* 1975 November;35(11 Pt. 2):3374-83.
- (248) Campbell TC, Junshi C. Diet and chronic degenerative diseases: perspectives from China. *Am J Clin Nutr* 1994 May;59(5 Suppl):1153S-61S.
- (249) Hopkins PN. Effects of dietary cholesterol on serum cholesterol: a meta-analysis and review. *Am J Clin Nutr* 1992 June;55(6):1060-70.
- (250) McNamara DJ. The impact of egg limitations on coronary heart disease risk: do the numbers add up? *J Am Coll Nutr* 2000 October;19(5 Suppl):540S-8S.
- (251) Wang Y, Crawford MA, Chen J, Li J, Ghebremeskel K, Campbell TC, Fan W, Parker R, Leyton J. Fish consumption, blood docosahexaenoic acid and chronic diseases in Chinese rural populations. *Comp Biochem Physiol A Mol Integr Physiol* 2003 September;136(1):127-40.
- (252) Hu FB, Stampfer MJ, Manson JE, Rimm E, Colditz GA, Speizer FE, Hennekens CH, Willett WC. Dietary protein and risk of ischemic heart disease in women [see comments]. *Am J Clin Nutr* 1999 August;70(2):221-7.
- (253) Campbell TC, Chen J. Energy balance: interpretation of data from rural China. *Toxicol Sci* 1999 December;52(2 Suppl):87-94.
- (254) Ornish D, Brown SE, Scherwitz LW, Billings JH, Armstrong WT, Ports TA, McLanahan SM, Kirkeeide RL, Brand RJ, Gould KL. Can lifestyle changes reverse coronary heart disease? The Lifestyle Heart Trial. *Lancet JID - 2985213R* 1990 July 21;336(8708):129-33.
- (255) Ornish D. *Eat More, Weigh Less: Dr. Dean Ornish's Life Choice Program for Losing Weight Safely While Eating Abundantly*. New York: Harper; 1997.
- (256) McDougall JA. *The McDougall Program for Maximum Weight Loss*. New York: Dutton; 1994.
- (257) Howson CP, Hiyama T, Wynder EL. The decline in gastric cancer: epidemiology of an unplanned triumph. *Epidemiol Rev* 1986;8:1-27.
- (258) Wang XQ, Terry PD, Yan H. Review of salt consumption and stomach cancer risk: epidemiological and biological evidence. *World J Gastroenterol* 2009 May 14;15(18):2204-13.
- (259) Nozaki K, Shimizu N, Inada K, Tsukamoto T, Inoue M, Kumagai T, Sugiyama A, Mizoshita T, Kaminishi M, Tatematsu M. Synergistic promoting effects of *Helicobacter pylori* infection and high-salt diet on gastric carcinogenesis in Mongolian gerbils. *Jpn J Cancer Res* 2002 October;93(10):1083-9.
- (260) Kato S, Tsukamoto T, Mizoshita T, Tanaka H, Kumagai T, Ota H, Katsuyama T, Asaka M, Tatematsu M. High salt diets dose-dependently promote gastric chemical carcinogenesis in

Helicobacter pylori-infected Mongolian gerbils associated with a shift in mucin production from glandular to surface mucous cells. *Int J Cancer* 2006 October 1;119(7):1558-66.

- (261) Leung WK, Wu KC, Wong CY, Cheng AS, Ching AK, Chan AW, Chong WW, Go MY, Yu J, To KF, Wang X, Chui YL, Fan DM, Sung JJ. Transgenic cyclooxygenase-2 expression and high salt enhanced susceptibility to chemical-induced gastric cancer development in mice. *Carcinogenesis* 2008 August;29(8):1648-54.
- (262) Toyoda T, Tsukamoto T, Hirano N, Mizoshita T, Kato S, Takasu S, Ban H, Tatematsu M. Synergistic upregulation of inducible nitric oxide synthase and cyclooxygenase-2 in gastric mucosa of Mongolian gerbils by a high-salt diet and *Helicobacter pylori* infection. *Histol Histopathol* 2008 May;23(5):593-9.
- (263) Shikata K, Kiyohara Y, Kubo M, Yonemoto K, Ninomiya T, Shirota T, Tanizaki Y, Doi Y, Tanaka K, Oishi Y, Matsumoto T, Iida M. A prospective study of dietary salt intake and gastric cancer incidence in a defined Japanese population: the Hisayama study. *Int J Cancer* 2006 July 1;119(1):196-201.
- (264) Cheng AS, Chan HL, Leung WK, To KF, Go MY, Chan JY, Liew CT, Sung JJ. Expression of HBx and COX-2 in chronic hepatitis B, cirrhosis and hepatocellular carcinoma: implication of HBx in upregulation of COX-2. *Mod Pathol* 2004 October;17(10):1169-79.
- (265) El-Sheikh SS, Madaan S, Alhasso A, Abel P, Stamp G, Lalani EN. Cyclooxygenase-2: a possible target in schistosoma-associated bladder cancer. *BJU Int* 2001 December;88(9):921-7.
- (266) Dunaif GE, Campbell TC. Dietary protein level and aflatoxin B1-induced preneoplastic hepatic lesions in the rat. *J Nutr* 1987 July;117(7):1298-302.
- (267) Cheng Z, Hu J, King J, Jay G, Campbell TC. Inhibition of hepatocellular carcinoma development in hepatitis B virus transfected mice by low dietary casein. *Hepatology* 1997 November;26(5):1351-4.
- (268) Hu JF, Cheng Z, Chisari FV, Vu TH, Hoffman AR, Campbell TC. Repression of hepatitis B virus (HBV) transgene and HBV-induced liver injury by low protein diet. *Oncogene* 1997 December 4;15(23):2795-801.
- (269) Overvik E, Kleman M, Berg I, Gustafsson JA. Influence of creatine, amino acids and water on the formation of the mutagenic heterocyclic amines found in cooked meat. *Carcinogenesis* 1989 December;10(12):2293-301.
- (270) Sugimura T, Wakabayashi K, Nakagama H, Nagao M. Heterocyclic amines: Mutagens/carcinogens produced during cooking of meat and fish. *Cancer Sci* 2004 April;95(4):290-9.
- (271) Dagnelie PC, van Staveren WA, Roos AH, Tuinstra LG, Burema J. Nutrients and contaminants in human milk from mothers on macrobiotic and omnivorous diets. *Eur J Clin Nutr* 1992 May;46(5):355-66.

- (272) Arguin H, Sanchez M, Bray GA, Lovejoy JC, Peters JC, Jandacek RJ, Chaput JP, Tremblay A. Impact of adopting a vegan diet or an olestra supplementation on plasma organochlorine concentrations: results from two pilot studies. *Br J Nutr* 2010 May;103(10):1433-41.
- (273) Yeung F, Hoberg JE, Ramsey CS, Keller MD, Jones DR, Frye RA, Mayo MW. Modulation of NF-kappaB-dependent transcription and cell survival by the SIRT1 deacetylase. *EMBO J* 2004 June 16;23(12):2369-80.
- (274) Salminen A, Ojala J, Huuskonen J, Kauppinen A, Suuronen T, Kaarniranta K. Interaction of aging-associated signaling cascades: inhibition of NF-kappaB signaling by longevity factors FoxOs and SIRT1. *Cell Mol Life Sci* 2008 April;65(7-8):1049-58.
- (275) Salminen A, Kauppinen A, Suuronen T, Kaarniranta K. SIRT1 longevity factor suppresses NF-kappaB -driven immune responses: regulation of aging via NF-kappaB acetylation? *Bioessays* 2008 October;30(10):939-42.
- (276) Kim YJ, Kim HJ, No JK, Chung HY, Fernandes G. Anti-inflammatory action of dietary fish oil and calorie restriction. *Life Sci* 2006 April 18;78(21):2523-32.
- (277) Jung KJ, Lee EK, Kim JY, Zou Y, Sung B, Heo HS, Kim MK, Lee J, Kim ND, Yu BP, Chung HY. Effect of short term calorie restriction on pro-inflammatory NF-kB and AP-1 in aged rat kidney. *Inflamm Res* 2009 March;58(3):143-50.
- (278) Zhang R, Chen HZ, Liu JJ, Jia YY, Zhang ZQ, Yang RF, Zhang Y, Xu J, Wei YS, Liu DP, Liang CC. SIRT1 suppresses activator protein-1 transcriptional activity and cyclooxygenase-2 expression in macrophages. *J Biol Chem* 2010 March 5;285(10):7097-110.
- (279) Johnson JB, Laub DR, John S. The effect on health of alternate day calorie restriction: eating less and more than needed on alternate days prolongs life. *Med Hypotheses* 2006;67(2):209-11.
- (280) Johnson JB, Summer W, Cutler RG, Martin B, Hyun DH, Dixit VD, Pearson M, Nassar M, Telljohann R, Maudsley S, Carlson O, John S, Laub DR, Mattson MP. Alternate day calorie restriction improves clinical findings and reduces markers of oxidative stress and inflammation in overweight adults with moderate asthma. *Free Radic Biol Med* 2007 March 1;42(5):665-74.
- (281) Johnson JB, Laub DR. *The alternate day diet*. New York: Perigee; 2008.
- (282) Stone JJ. 2010.
Ref Type: Personal Communication
- (283) Yan L, Spitznagel EL. Soy consumption and prostate cancer risk in men: a revisit of a meta-analysis. *Am J Clin Nutr* 2009 April;89(4):1155-63.
- (284) Hwang YW, Kim SY, Jee SH, Kim YN, Nam CM. Soy food consumption and risk of prostate cancer: a meta-analysis of observational studies. *Nutr Cancer* 2009;61(5):598-606.
- (285) Dong JY, Qin LQ. Soy isoflavones consumption and risk of breast cancer incidence or recurrence: a meta-analysis of prospective studies. *Breast Cancer Res Treat* 2011 January;125(2):315-23.

- (286) Wu AH, Yu MC, Tseng CC, Pike MC. Epidemiology of soy exposures and breast cancer risk. *Br J Cancer* 2008 January 15;98(1):9-14.
- (287) Qin LQ, Xu JY, Wang PY, Hoshi K. Soyfood intake in the prevention of breast cancer risk in women: a meta-analysis of observational epidemiological studies. *J Nutr Sci Vitaminol (Tokyo)* 2006 December;52(6):428-36.
- (288) Yan L, Spitznagel EL, Bosland MC. Soy consumption and colorectal cancer risk in humans: a meta-analysis. *Cancer Epidemiol Biomarkers Prev* 2010 January;19(1):148-58.
- (289) Budhathoki S, Joshi AM, Ohnaka K, Yin G, Toyomura K, Kono S, Mibu R, Tanaka M, Kakeji Y, Maehara Y, Okamura T, Ikejiri K, Futami K, Maekawa T, Yasunami Y, Takenaka K, Ichimiya H, Terasaka R. Soy food and isoflavone intake and colorectal cancer risk: the Fukuoka Colorectal Cancer Study. *Scand J Gastroenterol* 2010 October 24.
- (290) Yang G, Shu XO, Li H, Chow WH, Cai H, Zhang X, Gao YT, Zheng W. Prospective cohort study of soy food intake and colorectal cancer risk in women. *Am J Clin Nutr* 2009 February;89(2):577-83.
- (291) Akhter M, Inoue M, Kurahashi N, Iwasaki M, Sasazuki S, Tsugane S. Dietary soy and isoflavone intake and risk of colorectal cancer in the Japan public health center-based prospective study. *Cancer Epidemiol Biomarkers Prev* 2008 August;17(8):2128-35.
- (292) Myung SK, Ju W, Choi HJ, Kim SC. Soy intake and risk of endocrine-related gynaecological cancer: a meta-analysis. *BJOG* 2009 December;116(13):1697-705.
- (293) Nagata C, Takatsuka N, Kawakami N, Shimizu H. A prospective cohort study of soy product intake and stomach cancer death. *Br J Cancer* 2002 July 1;87(1):31-6.
- (294) Kuiper GG, Carlsson B, Grandien K, Enmark E, Haggblad J, Nilsson S, Gustafsson JA. Comparison of the ligand binding specificity and transcript tissue distribution of estrogen receptors alpha and beta. *Endocrinology* 1997 March;138(3):863-70.
- (295) Kuiper GG, Lemmen JG, Carlsson B, Corton JC, Safe SH, van der Saag PT, van Der BB, Gustafsson JA. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology* 1998 October;139(10):4252-63.
- (296) McCarty MF. Isoflavones made simple - genistein's agonist activity for the beta-type estrogen receptor mediates their health benefits. *Med Hypotheses* 2006;66(6):1093-114.
- (297) Warner M, Gustafsson JA. The role of estrogen receptor beta (ERbeta) in malignant diseases--a new potential target for antiproliferative drugs in prevention and treatment of cancer. *Biochem Biophys Res Commun* 2010 May 21;396(1):63-6.
- (298) Morani A, Warner M, Gustafsson JA. Biological functions and clinical implications of oestrogen receptors alfa and beta in epithelial tissues. *J Intern Med* 2008 August;264(2):128-42.
- (299) McPherson SJ, Hussain S, Balanathan P, Hedwards SL, Niranjana B, Grant M, Chandrasiri UP, Toivanen R, Wang Y, Taylor RA, Risbridger GP. Estrogen receptor-beta activated apoptosis in

benign hyperplasia and cancer of the prostate is androgen independent and TNFalpha mediated. *Proc Natl Acad Sci U S A* 2010 February 16;107(7):3123-8.

- (300) Papaxoinis K, Triantafyllou K, Sasco AJ, Nicolopoulou-Stamati P, Ladas SD. Subsite-specific differences of estrogen receptor beta expression in the normal colonic epithelium: implications for carcinogenesis and colorectal cancer epidemiology. *Eur J Gastroenterol Hepatol* 2010 May;22(5):614-9.
- (301) Kennelly R, Kavanagh DO, Hogan AM, Winter DC. Oestrogen and the colon: potential mechanisms for cancer prevention. *Lancet Oncol* 2008 April;9(4):385-91.
- (302) La VC, Gallus S, Fernandez E. Hormone replacement therapy and colorectal cancer: an update. *J Br Menopause Soc* 2005 December;11(4):166-72.
- (303) Campbell-Thompson M, Reyher KK, Wilkinson LB. Immunolocalization of estrogen receptor alpha and beta in gastric epithelium and enteric neurons. *J Endocrinol* 2001 October;171(1):65-73.
- (304) Xu CY, Guo JL, Jiang ZN, Xie SD, Shen JG, Shen JY, Wang LB. Prognostic role of estrogen receptor alpha and estrogen receptor beta in gastric cancer. *Ann Surg Oncol* 2010 September;17(9):2503-9.
- (305) Chandanos E, Lagergren J. Oestrogen and the enigmatic male predominance of gastric cancer. *Eur J Cancer* 2008 November;44(16):2397-403.
- (306) Min WK, Sung HY, Choi YS. Suppression of colonic aberrant crypt foci by soy isoflavones is dose-independent in dimethylhydrazine-treated rats. *J Med Food* 2010 June;13(3):495-502.
- (307) Wu Y, Niwa K, Onogi K, Tang L, Mori H, Tamaya T. Effects of selective estrogen receptor modulators and genistein on the expression of ERalpha/beta and COX-1/2 in ovariectomized mouse uteri. *Eur J Gynaecol Oncol* 2007;28(2):89-94.
- (308) Swami S, Krishnan AV, Moreno J, Bhattacharyya RS, Gardner C, Brooks JD, Peehl DM, Feldman D. Inhibition of prostaglandin synthesis and actions by genistein in human prostate cancer cells and by soy isoflavones in prostate cancer patients. *Int J Cancer* 2009 May 1;124(9):2050-9.
- (309) Chang KC, Wang Y, Oh IG, Jenkins S, Freedman LP, Thompson CC, Chung JH, Nagpal S. Estrogen receptor beta is a novel therapeutic target for photoaging. *Mol Pharmacol* 2010 May;77(5):744-50.
- (310) Kim SY, Kim SJ, Lee JY, Kim WG, Park WS, Sim YC, Lee SJ. Protective effects of dietary soy isoflavones against UV-induced skin-aging in hairless mouse model. *J Am Coll Nutr* 2004 April;23(2):157-62.
- (311) Chiu TM, Huang CC, Lin TJ, Fang JY, Wu NL, Hung CF. In vitro and in vivo anti-photoaging effects of an isoflavone extract from soybean cake. *J Ethnopharmacol* 2009 October 29;126(1):108-13.

- (312) Dewell A, Weidner G, Sumner MD, Barnard RJ, Marlin RO, Daubenmier JJ, Chi C, Carroll PR, Ornish D. Relationship of dietary protein and soy isoflavones to serum IGF-1 and IGF binding proteins in the Prostate Cancer Lifestyle Trial. *Nutr Cancer* 2007;58(1):35-42.
- (313) McCarty MF. Optimizing exercise for fat loss. *Med Hypotheses* 1995 May;44(5):325-30.
- (314) Bahadori B, McCarty MF, Barroso-Aranda J, Gustin JC, Contreras F. A "mini-fast with exercise" protocol for fat loss. *Med Hypotheses* 2009 October;73(4):619-22.
- (315) Van PK, Szlufcik K, Nielens H, Pelgrim K, Deldicque L, Hesselink M, Van Veldhoven PP, Hespel P. Training in the fasted state improves glucose tolerance during fat-rich diet. *J Physiol* 2010 November 1;588(Pt 21):4289-302.