

Nutraceutical and Drug Measures for Expanding the T Regulatory Lymphocyte Pool

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Abstract

T regulatory cells, owing to their down-regulatory impact on inflammatory mechanisms, can aid control of autoimmunity, certain neurodegenerative disorders, atherosclerosis, and metabolic syndrome. Safe and practical measures for expanding the Treg pool may therefore have broad utility for providing protection from a host of common health disorders. Measures which appear likely to be useful in this regard include: subcutaneous or oral administration of heat shock proteins, or epitopes thereof; the cyanobacterial protein phycocyanin and its chromophore phycocyanobilin; vitamin D; PPARgamma agonists such as pioglitazone; GABA or GABAergic drugs; and possibly a vegan diet. Functional combinations of these measures which best promote Treg induction and activity may have considerable potential in prevention and therapy.

Treg Expansion May have Broad Clinical Utility

T regulatory (Treg) cells play a key role in the prevention and control of autoimmunity and chronic inflammatory syndromes, including certain neurodegenerative disorders.^{1,2} Tregs also oppose atherogenesis and can stabilize plaque, likely decreasing risk for heart attack, stroke, and aneurysms.³⁻¹¹ Notably, Treg levels tend to be decreased in patients with unstable angina, as opposed to patients with stable angina or healthy controls. And Tregs in visceral adipose tissue, via production of IL-10, can dampen adipose inflammation and thereby help to preserve good systemic insulin sensitivity and prevent the metabolic syndrome.¹²⁻²⁰ Hence, practical strategies for expanding the body's pool of Tregs could confer a wide range of health benefits.

Vaccination with Heat Shock Proteins

Oral or subcutaneous vaccination with heat shock proteins, or epitopes derived therefrom, has been shown to promote Treg activity, likely because a significant proportion of thymic-derived nTregs recognize hsp-derived epitopes.²¹⁻²³ These Tregs may be induced to proliferate and activated when dendritic cells present them with these epitopes; they then may migrate to inflamed tissues, where they can tolerize dendritic cells presenting hsp epitopes, as well as interacting directly with T cells to suppress their pro-inflammatory activity. Oral administration of bacterial heat shock proteins – as with the lyophilized E.coli extract Subreum (OM-89) – has shown clinical utility in rheumatoid arthritis comparable to that of DMARD drugs.²⁴⁻²⁹ Oral administration of hsp60 (or an epitope derived from it) to LDL receptor-deficient mice, markedly suppressed development of atherosclerosis while promoting Treg induction.³⁰ Parenterally administered hsp epitopes are effective in rodent models of autoimmunity,^{21-23,31-34} and subcutaneous injection of hsp-enriched monocytes, heat-shocked *ex vivo* and then incubated at physiological temperature for a further 24-48 hours prior to injection, has been reported anecdotally to achieve clinical benefit in a wide range of autoimmune disorders (the MAM-14 strategy).³⁵

Phycocyanobilin – A Bilirubin Mimetic

Bilirubin, as well as its homologue phycocyanobilin (PhyCB), a major component of cyanobacteria used as food such as spirulina, promote the conversion of CD4⁺ lymphocytes to foxp3⁺ Tregs, in vitro and in vivo.³⁶⁻⁴⁰ This may explain why induction of heme oxygenase-1, which generates bilirubin's precursor biliverdin, has a similar effect.^{36;41-43} Bilirubin is an endogenous activating ligand for the aryl hydrocarbon receptor (AhR),⁴⁴⁻⁴⁷ which can promote the induction of T regulatory cells (both foxp3⁺, and foxp3⁻ producing IL-10);⁴⁸⁻⁵¹ this likely explains the impact of bilirubin on Treg induction (Libor Vitek, personal communication). Activating ligands of AhR have shown therapeutic efficacy in rodent models of multiple sclerosis, colitis, type 1 diabêtes, autoimmune uveitis, peanut allergy, and graft rejection.⁵²⁻⁵⁸ It is reasonable to predict that phycocyanorubin, produced within cells by bilirubin reductase-mediated reduction of PhyCB, is likewise an activating ligand for AhR. Oral administration of phycocyanin or its chromophore PhyCB may therefore be useful for Treg expansion, and oral phycocyanin is beneficial in rodent models of autoimmunity and chronic inflammation.⁵⁹⁻⁶⁴ The antioxidant efficacy of these agents may also contribute to their anti-inflammatory impact.^{65;66} The nutraceutical diindolylmethane likewise has agonist activity for AhR, and has potential for control of autoimmunity.⁶⁷⁻⁶⁹ There is recent evidence that the kynurenine generated by indolamine 2,3-dioxygenase activity also serves as an endogenous ligand for AhR.^{70;71}

Vitamin D

Low vitamin D status has been linked to increased risk for various autoimmune disorders, as well as metabolic syndrome and atherogenic disease.⁷²⁻⁷⁵ Activated dendritic cells express vitamin D 1- α -hydroxylase activity, and hence can convert cholecalciferol to the active hormone calcitriol; the extent to which this occurs will be proportional to the circulating concentration of vitamin D.⁷⁶ Dendritic cells exposed to calcitriol develop a tolerogenic phenotype – expressing IL-10 and IDO – and can interact with naïve CD4⁺ lymphocytes to convert them to Tregs.^{77;78} Calcitriol can also act directly on lymphocytes in the microenvironment of these dendritic cells; a vitamin D response element has been identified in the first intron of the Foxp3 gene, and promotes its transcription.⁷⁹ A physiologically significant dose of vitamin D – 140,000 IU monthly – has been reported to expand the pool of CD4⁺CD25⁺Foxp3⁺ lymphocytes in healthy humans.⁸⁰

PPARgamma Agonists

There is recent evidence that PPARgamma agonists, such as the drug pioglitazone, can act both on dendritic cells and naïve CD4⁺ lymphocytes to promote generation of iTregs.⁸¹⁻⁸⁷ A key effect of these agents is to boost the capacity of dendritic cells to synthesize retinoic acid, which promotes induction of iTregs; the maturation of dendritic cells is also opposed, rendering them more tolerogenic.^{87;88} PPARgamma activity within lymphocytes boosts Foxp3 expression by modulating methylation of DNA in the Foxp3 promoter region.⁸⁵ PPARgamma is highly expressed in the Tregs that congregate in visceral adipose tissue, and likely contributes to the favorable impact of thiazolidinedione therapy on insulin sensitivity in diabetics.^{15;16} Pioglitazone administration was found to be beneficial in rodent models of autoimmune arthritis and uveitis, and to stabilize plaque in atheroma-prone mice.^{84;86;89} Moreover, a controlled clinical trial found that this drug was a useful adjuvant to methotrexate therapy in rheumatoid arthritis.⁸⁹

GABA and GABAergic Agents

Oral administration of gamma-amino-butyric acid (GABA) is protective in a mouse models of type 1 diabetes (NOD) and rheumatoid arthritis, owing in part to anti-inflammatory effects mediated by GABAergic receptors on macrophages and T cells.⁹⁰⁻⁹² GABAergic drugs (topiramate, vigabatrin) ameliorate the course of experimental autoimmune encephalomyelitis, a mouse model of multiple sclerosis.⁹³ GABA administration (2 g /l in drinking water) likewise aids maintenance of insulin sensitivity in fat-fed mice; this latter study reported an increase in the CD4+Foxp3+ pool in these mice.⁹⁴ How GABA promotes Treg induction requires further clarification, but may reflect a tolerizing effect on dendritic cells.⁹³ Hence, ample oral doses of GABA, or treatment with GABAergic drugs, might have clinical potential for promoting Treg activity.⁹⁴ GABA per se has the advantage that it does not cross the blood-brain barrier, and hence would not produce central side effects; but from the standpoint of convenience, GABAergic drugs likely have better pharmacokinetics. It has credibly been proposed that oral GABA could be useful for prevention or treatment of atherosclerosis.⁹⁵

Fasting and Vegan Diet

Fasting increases the Treg pool in mice, and this has been shown to reflect an acute reduction in circulating levels of leptin, which opposes Treg induction and exerts various other pro-inflammatory effects.⁹⁶ A fast of several days duration, followed by adoption of a vegan diet, has been found to be clinically beneficial in rheumatoid arthritis.⁹⁷⁻⁹⁹ The relatively low risk for autoimmune disorders in sub-Saharan Africans and East Asians during the early to mid-twentieth century remains unexplained.¹⁰⁰⁻¹⁰² It has been speculated that the leanness and relatively low intake of essential amino acids associated with long-term consumption of vegan diets may promote Treg expression; this hypothesis has not yet been tested.¹⁰¹

Multifocal Protocols for Treg Induction

A protocol consisting of hsp vaccination, complemented with adjuvant measures that might include vitamin D, phycocyanin, pioglitazone, GABA or GABAergic drugs, and a vegan diet, may therefore have clinical potential for treating autoimmune disorders, neurodegenerative conditions driven in part by activated microglia, metabolic syndrome/diabetes, and for stabilizing atherosclerotic plaque (thereby lessening risk for vascular accidents). Subcutaneous hsp vaccination (as in MAM-14) could be used episodically, whereas oral administration of hsps could be employed in the intervals between such therapy. Rodent models of autoimmunity, neurodegeneration, metabolic syndrome, and atherosclerosis might be employed to determine what combinations of these options might have the most important impact on Treg expression and activity, and on the progression of these syndromes.

References

1. Allan SE, Broady R, Gregori S et al. CD4+ T-regulatory cells: toward therapy for human diseases. *Immunol Rev* 2008;223:391-421.
2. Gendelman HE, Appel SH. Neuroprotective activities of regulatory T cells. *Trends Mol Med* 2011;17(12):687-688.
3. Yang K, Li D, Luo M, Hu Y. Generation of HSP60-specific regulatory T cell and effect on atherosclerosis. *Cell Immunol* 2006;243(2):90-95.
4. Cheng X, Yu X, Ding YJ et al. The Th17/Treg imbalance in patients with acute coronary syndrome. *Clin Immunol* 2008;127(1):89-97.
5. Entin-Meer M, Afek A, George J. Regulatory T-cells, FoxP3 and atherosclerosis. *Adv Exp Med Biol* 2009;665:106-114.
6. Yin M, Zhang J, Wang Y et al. Deficient CD4+CD25+ T regulatory cell function in patients with abdominal aortic aneurysms. *Arterioscler Thromb Vasc Biol* 2010;30(9):1825-1831.
7. Foks AC, Frodermann V, ter BM et al. Differential effects of regulatory T cells on the initiation and regression of atherosclerosis. *Atherosclerosis* 2011;218(1):53-60.
8. George J, Schwartzberg S, Medvedovsky D et al. Regulatory T cells and IL-10 levels are reduced in patients with vulnerable coronary plaques. *Atherosclerosis* 2012;222(2):519-523.
9. Dietel B, Cicha I, Voskens CJ, Verhoeven E, Achenbach S, Garlichs CD. Decreased numbers of regulatory T cells are associated with human atherosclerotic lesion vulnerability and inversely correlate with infiltrated mature dendritic cells. *Atherosclerosis* 2013;230(1):92-99.
10. Ait-Oufella H, Wang Y, Herbin O et al. Natural Regulatory T Cells Limit Angiotensin II-Induced Aneurysm Formation and Rupture in Mice. *Arterioscler Thromb Vasc Biol* 2013;33(10):2374-2379.
11. Zhang WC, Wang J, Shu YW et al. Impaired thymic export and increased apoptosis account for regulatory T cell defects in patients with non-ST segment elevation acute coronary syndrome. *J Biol Chem* 2012;287(41):34157-34166.

12. Eller K, Kirsch A, Wolf AM et al. Potential role of regulatory T cells in reversing obesity-linked insulin resistance and diabetic nephropathy. *Diabetes* 2011;60(11):2954-2962.
13. Wagner NM, Brandhorst G, Czepluch F et al. Circulating regulatory T cells are reduced in obesity and may identify subjects at increased metabolic and cardiovascular risk. *Obesity (Silver Spring)* 2013;21(3):461-468.
14. Ilan Y, Maron R, Tukupah AM et al. Induction of regulatory T cells decreases adipose inflammation and alleviates insulin resistance in ob/ob mice. *Proc Natl Acad Sci U S A* 2010;107(21):9765-9770.
15. Cipolletta D, Feuerer M, Li A et al. PPAR-gamma is a major driver of the accumulation and phenotype of adipose tissue Treg cells. *Nature* 2012;486(7404):549-553.
16. Hamaguchi M, Sakaguchi S. Regulatory T cells expressing PPAR-gamma control inflammation in obesity. *Cell Metab* 2012;16(1):4-6.
17. Chen X, Wu Y, Wang L. Fat-resident Tregs: an emerging guard protecting from obesity-associated metabolic disorders. *Obes Rev* 2013;14(7):568-578.
18. Feuerer M, Herrero L, Cipolletta D et al. Lean, but not obese, fat is enriched for a unique population of regulatory T cells that affect metabolic parameters. *Nat Med* 2009;15(8):930-939.
19. Cipolletta D, Kolodin D, Benoist C, Mathis D. Tissue-resident Foxp3+CD4+ T cells that impact organismal metabolism. *Semin Immunol* 2011;23(6):431-437.
20. Pettersson US, Walden TB, Carlsson PO, Jansson L, Phillipson M. Female mice are protected against high-fat diet induced metabolic syndrome and increase the regulatory T cell population in adipose tissue. *PLoS ONE* 2012;7(9):e46057.
21. Hauet-Broere F, Wieten L, Guichelaar T, Berlo S, van der Zee R, van EW. Heat shock proteins induce T cell regulation of chronic inflammation. *Ann Rheum Dis* 2006;65 Suppl 3:iii65-iii68.
22. Keijzer C, Wieten L, van HM, van der Zee R, van EW, Broere F. Heat shock proteins are therapeutic targets in autoimmune diseases and other chronic inflammatory conditions. *Expert Opin Ther Targets* 2012;16(9):849-857.
23. van Herwijnen MJ, Wieten L, van der Zee R et al. Regulatory T cells that recognize a ubiquitous stress-inducible self-antigen are long-lived suppressors of autoimmune arthritis. *Proc Natl Acad Sci U S A* 2012;109(35):14134-14139.

24. Vischer TL. A double blind multicentre study of OM-8980 and auranofin in rheumatoid arthritis. *Ann Rheum Dis* 1988;47(7):582-587.
25. Hauzeur JP, Appelboom T. Double-blind, placebo-controlled study of OM-8980 in rheumatoid arthritis. *Rheumatol Int* 1989;9(2):71-76.
26. Verstraeten A, Sileghem A, Dequeker J. OM-8980 and D-penicillamine in the treatment of rheumatoid arthritis. A 12-month double-blind randomized study. *Scand J Rheumatol* 1990;19(6):422-431.
27. Vischer TL. Follow-up with OM-8980 after a double-blind study of OM-8980 and auranofin in rheumatoid arthritis. *Clin Rheumatol* 1990;9(3):356-361.
28. Rosenthal M, Bahous I, Ambrosini G. Longterm treatment of rheumatoid arthritis with OM-8980. A retrospective study. *J Rheumatol* 1991;18(12):1790-1793.
29. Muller-Fassbender HR. A 6-month randomized dose range study of OM-8980 in rheumatoid arthritis. *Br J Rheumatol* 1993;32(8):746-750.
30. van Puijvelde GH, van ET, van Wanrooij EJ et al. Induction of oral tolerance to HSP60 or an HSP60-peptide activates T cell regulation and reduces atherosclerosis. *Arterioscler Thromb Vasc Biol* 2007;27(12):2677-2683.
31. Billetta R, Ghahramani N, Morrow O et al. Epitope-specific immune tolerization ameliorates experimental autoimmune encephalomyelitis. *Clin Immunol* 2012;145(2):94-101.
32. Ma YJ, Lu Y, Hou J et al. Vaccination of non-obese diabetic mice with a fragment of peptide P277 attenuates insulin-dependent diabetes mellitus. *Int Immunopharmacol* 2011;11(9):1298-1302.
33. Coelho V, Faria AM. HSP60: issues and insights on its therapeutic use as an immunoregulatory agent. *Front Immunol* 2011;2:97.
34. Wieten L, Berlo SE, Ten Brink CB et al. IL-10 is critically involved in mycobacterial HSP70 induced suppression of proteoglycan-induced arthritis. *PLoS ONE* 2009;4(1):e4186.
35. McCarty MF, Al-Harbi SA. Vaccination with heat-shocked mononuclear cells as a strategy for treating neurodegenerative disorders driven by microglial inflammation. *Med Hypotheses* 2013.

36. Lee SS, Gao W, Mazzola S et al. Heme oxygenase-1, carbon monoxide, and bilirubin induce tolerance in recipients toward islet allografts by modulating T regulatory cells. *FASEB J* 2007;21(13):3450-3457.
37. Liu Y, Li P, Lu J et al. Bilirubin possesses powerful immunomodulatory activity and suppresses experimental autoimmune encephalomyelitis. *J Immunol* 2008;181(3):1887-1897.
38. Rocuts F, Zhang X, Yan J et al. Bilirubin promotes de novo generation of T regulatory cells. *Cell Transplant* 2010;19(4):443-451.
39. Penton-Rol G, Martinez-Sanchez G, Cervantes-Llanos M et al. C-Phycocyanin ameliorates experimental autoimmune encephalomyelitis and induces regulatory T cells. *Int Immunopharmacol* 2011;11(1):29-38.
40. McCarty MF. Clinical potential of phycocyanobilin for induction of T regulatory cells in the management of inflammatory disorders. *Med Hypotheses* 2011;77(6):1031-1033.
41. Xia ZW, Xu LQ, Zhong WW et al. Heme oxygenase-1 attenuates ovalbumin-induced airway inflammation by up-regulation of foxp3 T-regulatory cells, interleukin-10, and membrane-bound transforming growth factor- 1. *Am J Pathol* 2007;171(6):1904-1914.
42. Sun L, Shi T, Qiao H et al. Hepatic overexpression of heme oxygenase-1 improves liver allograft survival by expanding T regulatory cells. *J Surg Res* 2011;166(2):e187-e194.
43. Li JG, Zhuan-Sun YX, Wen B et al. Human Mesenchymal Stem Cells Elevate CD4+CD25+CD127low/- Regulatory T Cells of Asthmatic Patients via Heme Oxygenase-1. *Iran J Allergy Asthma Immunol* 2013;12(3):228-235.
44. Sinal CJ, Bend JR. Aryl hydrocarbon receptor-dependent induction of cyp1a1 by bilirubin in mouse hepatoma hepa 1c1c7 cells. *Mol Pharmacol* 1997;52(4):590-599.
45. Phelan D, Winter GM, Rogers WJ, Lam JC, Denison MS. Activation of the Ah receptor signal transduction pathway by bilirubin and biliverdin. *Arch Biochem Biophys* 1998;357(1):155-163.
46. Bock KW, Kohle C. Contributions of the Ah receptor to bilirubin homeostasis and its antioxidative and atheroprotective functions. *Biol Chem* 2010;391(6):645-653.
47. Togawa H, Shinkai S, Mizutani T. Induction of human UGT1A1 by bilirubin through AhR dependent pathway. *Drug Metab Lett* 2008;2(4):231-237.

48. Marshall NB, Vorachek WR, Stepan LB, Mourich DV, Kerkvliet NI. Functional characterization and gene expression analysis of CD4⁺ CD25⁺ regulatory T cells generated in mice treated with 2,3,7,8-tetrachlorodibenzo-p-dioxin. *J Immunol* 2008;181(4):2382-2391.
49. Quintana FJ, Basso AS, Iglesias AH et al. Control of T(reg) and T(H)17 cell differentiation by the aryl hydrocarbon receptor. *Nature* 2008;453(7191):65-71.
50. Gandhi R, Kumar D, Burns EJ et al. Activation of the aryl hydrocarbon receptor induces human type 1 regulatory T cell-like and Foxp3(+) regulatory T cells. *Nat Immunol* 2010;11(9):846-853.
51. Marshall NB, Kerkvliet NI. Dioxin and immune regulation: emerging role of aryl hydrocarbon receptor in the generation of regulatory T cells. *Ann N Y Acad Sci* 2010;1183:25-37.
52. Quintana FJ, Murugaiyan G, Farez MF et al. An endogenous aryl hydrocarbon receptor ligand acts on dendritic cells and T cells to suppress experimental autoimmune encephalomyelitis. *Proc Natl Acad Sci U S A* 2010;107(48):20768-20773.
53. Singh NP, Singh UP, Singh B, Price RL, Nagarkatti M, Nagarkatti PS. Activation of aryl hydrocarbon receptor (AhR) leads to reciprocal epigenetic regulation of FoxP3 and IL-17 expression and amelioration of experimental colitis. *PLoS ONE* 2011;6(8):e23522.
54. Kerkvliet NI, Stepan LB, Vorachek W et al. Activation of aryl hydrocarbon receptor by TCDD prevents diabetes in NOD mice and increases Foxp3⁺ T cells in pancreatic lymph nodes. *Immunotherapy* 2009;1(4):539-547.
55. Zhang L, Ma J, Takeuchi M et al. Suppression of experimental autoimmune uveoretinitis by inducing differentiation of regulatory T cells via activation of aryl hydrocarbon receptor. *Invest Ophthalmol Vis Sci* 2010;51(4):2109-2117.
56. Schulz VJ, Smit JJ, Willemsen KJ et al. Activation of the aryl hydrocarbon receptor suppresses sensitization in a mouse peanut allergy model. *Toxicol Sci* 2011;123(2):491-500.
57. Benson JM, Shepherd DM. Aryl hydrocarbon receptor activation by TCDD reduces inflammation associated with Crohn's disease. *Toxicol Sci* 2011;120(1):68-78.
58. Cai LJ, Yu DW, Gao Y, Yang C, Zhou HM, Chen ZH. Activation of aryl hydrocarbon receptor prolongs survival of fully mismatched cardiac allografts. *J Huazhong Univ Sci Technolog Med Sci* 2013;33(2):199-204.

59. Romay C, Gonzalez R, Ledon N, Ramirez D, Rimbau V. C-phycoerythrin: a biliprotein with antioxidant, anti-inflammatory and neuroprotective effects. *Curr Protein Pept Sci* 2003;4(3):207-216.
60. Ramirez D, Gonzalez R, Merino N, Rodriguez S, Ancheta O. Inhibitory effects of Spirulina in zymosan-induced arthritis in mice. *Mediators Inflamm* 2002;11(2):75-79.
61. Kumar N, Singh S, Patro N, Patro I. Evaluation of protective efficacy of Spirulina platensis against collagen-induced arthritis in rats. *Inflammopharmacology* 2009;17(3):181-190.
62. Rasool M, Sabina EP, Lavanya B. Anti-inflammatory effect of Spirulina fusiformis on adjuvant-induced arthritis in mice. *Biol Pharm Bull* 2006;29(12):2483-2487.
63. Patro N, Sharma A, Kariya K, Patro I. Spirulina platensis protects neurons via suppression of glial activation and peripheral sensitization leading to restoration of motor function in collagen-induced arthritic rats. *Indian J Exp Biol* 2011;49(10):739-748.
64. Coskun ZK, Kerem M, Gurbuz N et al. The study of biochemical and histopathological effects of spirulina in rats with TNBS-induced colitis. *Bratisl Lek Listy* 2011;112(5):235-243.
65. McCarty MF. Clinical potential of Spirulina as a source of phycoerythrin. *J Med Food* 2007;10(4):566-570.
66. Zheng J, Inoguchi T, Sasaki S et al. Phycoerythrin and phycoerythrin from Spirulina platensis protect against diabetic nephropathy by inhibiting oxidative stress. *Am J Physiol Regul Integr Comp Physiol* 2013;304(2):R110-R120.
67. Chen I, McDougal A, Wang F, Safe S. Aryl hydrocarbon receptor-mediated antiestrogenic and antitumorigenic activity of diindolylmethane. *Carcinogenesis* 1998;19(9):1631-1639.
68. Rouse M, Singh NP, Nagarkatti PS, Nagarkatti M. Indoles mitigate the development of experimental autoimmune encephalomyelitis by induction of reciprocal differentiation of regulatory T cells and Th17 cells. *Br J Pharmacol* 2013;169(6):1305-1321.
69. Huang Z, Jiang Y, Yang Y et al. 3,3'-Diindolylmethane alleviates oxazolone-induced colitis through Th2/Th17 suppression and Treg induction. *Mol Immunol* 2013;53(4):335-344.
70. Nguyen NT, Kimura A, Nakahama T et al. Aryl hydrocarbon receptor negatively regulates dendritic cell immunogenicity via a kynurenine-dependent mechanism. *Proc Natl Acad Sci U S A* 2010;107(46):19961-19966.

71. Mezrich JD, Fechner JH, Zhang X, Johnson BP, Burlingham WJ, Bradfield CA. An interaction between kynurenine and the aryl hydrocarbon receptor can generate regulatory T cells. *J Immunol* 2010;185(6):3190-3198.
72. Yang CY, Leung PS, Adamopoulos IE, Gershwin ME. The implication of vitamin d and autoimmunity: a comprehensive review. *Clin Rev Allergy Immunol* 2013;45(2):217-226.
73. Pludowski P, Holick MF, Pilz S et al. Vitamin D effects on musculoskeletal health, immunity, autoimmunity, cardiovascular disease, cancer, fertility, pregnancy, dementia and mortality-a review of recent evidence. *Autoimmun Rev* 2013;12(10):976-989.
74. Boucher BJ. Is vitamin D status relevant to metabolic syndrome? *Dermatoendocrinol* 2012;4(2):212-224.
75. Anagnostis P, Athyros VG, Adamidou F, Florentin M, Karagiannis A. Vitamin D and cardiovascular disease: a novel agent for reducing cardiovascular risk? *Curr Vasc Pharmacol* 2010;8(5):720-730.
76. Jeffery LE, Wood AM, Qureshi OS et al. Availability of 25-hydroxyvitamin D(3) to APCs controls the balance between regulatory and inflammatory T cell responses. *J Immunol* 2012;189(11):5155-5164.
77. Farias AS, Spagnol GS, Bordeaux-Rego P et al. Vitamin D3 induces IDO(+) tolerogenic DCs and enhances Treg, reducing the severity of EAE. *CNS Neurosci Ther* 2013;19(4):269-277.
78. van der Aar AM, Sibiryak DS, Bakdash G et al. Vitamin D3 targets epidermal and dermal dendritic cells for induction of distinct regulatory T cells. *J Allergy Clin Immunol* 2011;127(6):1532-1540.
79. Kang SW, Kim SH, Lee N et al. 1,25-Dihydroxyvitamin D3 promotes FOXP3 expression via binding to vitamin D response elements in its conserved noncoding sequence region. *J Immunol* 2012;188(11):5276-5282.
80. Prietl B, Treiber G, Mader JK et al. High-dose cholecalciferol supplementation significantly increases peripheral CD4 Tregs in healthy adults without negatively affecting the frequency of other immune cells. *Eur J Nutr* 2013.
81. Wohlfert EA, Nichols FC, Nevis E, Clark RB. Peroxisome proliferator-activated receptor gamma (PPARgamma) and immunoregulation: enhancement of regulatory T cells through PPARgamma-dependent and -independent mechanisms. *J Immunol* 2007;178(7):4129-4135.

82. Wang W, Zhu Z, Zhu B, Ma Z. Peroxisome proliferator-activated receptor-gamma agonist induces regulatory T cells in a murine model of allergic rhinitis. *Otolaryngol Head Neck Surg* 2011;144(4):506-513.
83. Zhao W, Berthier CC, Lewis EE, McCune WJ, Kretzler M, Kaplan MJ. The peroxisome-proliferator activated receptor-gamma agonist pioglitazone modulates aberrant T cell responses in systemic lupus erythematosus. *Clin Immunol* 2013;149(1):119-132.
84. Okunuki Y, Usui Y, Nakagawa H et al. Peroxisome proliferator-activated receptor-gamma agonist pioglitazone suppresses experimental autoimmune uveitis. *Exp Eye Res* 2013.
85. Lei J, Hasegawa H, Matsumoto T, Yasukawa M. Peroxisome proliferator-activated receptor alpha and gamma agonists together with TGF-beta convert human CD4+. *J Immunol* 2010;185(12):7186-7198.
86. Shen Y, Yuan Z, Yin A et al. Antiatherogenic effect of pioglitazone on uremic apolipoprotein E knockout mice by modulation of the balance of regulatory and effector T cells. *Atherosclerosis* 2011;218(2):330-338.
87. Housley WJ, O'Connor CA, Nichols F et al. PPARgamma regulates retinoic acid-mediated DC induction of Tregs. *J Leukoc Biol* 2009;86(2):293-301.
88. Klotz L, Dani I, Edenhofer F et al. Peroxisome proliferator-activated receptor gamma control of dendritic cell function contributes to development of CD4+ T cell anergy. *J Immunol* 2007;178(4):2122-2131.
89. Shahin D, Toraby EE, Abdel-Malek H, Boshra V, Elsamanoudy AZ, Shaheen D. Effect of peroxisome proliferator-activated receptor gamma agonist (pioglitazone) and methotrexate on disease activity in rheumatoid arthritis (experimental and clinical study). *Clin Med Insights Arthritis Musculoskelet Disord* 2011;4:1-10.
90. Tian J, Lu Y, Zhang H, Chau CH, Dang HN, Kaufman DL. Gamma-aminobutyric acid inhibits T cell autoimmunity and the development of inflammatory responses in a mouse type 1 diabetes model. *J Immunol* 2004;173(8):5298-5304.
91. Soltani N, Qiu H, Aleksic M et al. GABA exerts protective and regenerative effects on islet beta cells and reverses diabetes. *Proc Natl Acad Sci U S A* 2011;108(28):11692-11697.
92. Tian J, Yong J, Dang H, Kaufman DL. Oral GABA treatment downregulates inflammatory responses in a mouse model of rheumatoid arthritis. *Autoimmunity* 2011;44(6):465-470.

93. Bhat R, Axtell R, Mitra A et al. Inhibitory role for GABA in autoimmune inflammation. *Proc Natl Acad Sci U S A* 2010;107(6):2580-2585.
94. Tian J, Dang HN, Yong J et al. Oral treatment with gamma-aminobutyric acid improves glucose tolerance and insulin sensitivity by inhibiting inflammation in high fat diet-fed mice. *PLoS ONE* 2011;6(9):e25338.
95. Yang Y, Luo H, Cheng LX, Liu K. Inhibitory role for GABA in atherosclerosis. *Med Hypotheses* 2013.
96. Liu Y, Yu Y, Matarese G, La CA. Cutting edge: fasting-induced hypoleptinemia expands functional regulatory T cells in systemic lupus erythematosus. *J Immunol* 2012;188(5):2070-2073.
97. Kjeldsen-Kragh J, Haugen M, Borchgrevink CF et al. Controlled trial of fasting and one-year vegetarian diet in rheumatoid arthritis. *Lancet* 1991;338(8772):899-902.
98. Kjeldsen-Kragh J. Rheumatoid arthritis treated with vegetarian diets. *Am J Clin Nutr* 1999;70(3 Suppl):594S-600S.
99. McDougall J, Bruce B, Spiller G, Westerdahl J, McDougall M. Effects of a very low-fat, vegan diet in subjects with rheumatoid arthritis. *J Altern Complement Med* 2002;8(1):71-75.
100. Burkitt DP, Trowell HC. *Refined Carbohydrate Foods and Disease* London: Academic Press; 1975.
101. McCarty, M. F. A Vegan Diet of Modest Protein Content, by Down-Regulating Akt-mTORC1 Activity in Lymphocytes, May Aid Induction of T Regulatory Cells . 2013.
Ref Type: Unpublished Work
102. McCarty MF. Upregulation of lymphocyte apoptosis as a strategy for preventing and treating autoimmune disorders: a role for whole-food vegan diets, fish oil and dopamine agonists. *Med Hypotheses* 2001;57(2):258-275.