

NADPH Oxidase May be a Primary Mediator of the Adverse Effects of Cigarette Smoke and of Complement Activation on Retinal Pigment Epithelium during Induction of Age-Related Macular Degeneration

Mark F. McCarty, Guardion Health Sciences, markfmccarty@gmail.com

Abstract

Cigarette smoking, as well as genetic variants expected to up-regulate activation of the alternative complement pathway, are clearly linked to increased risk for both forms of age-related macular degeneration (AMD). Moderately reactive soluble compounds in cigarette smoke extract (such as acrolein), and the pro-inflammatory complement metabolite C5a, have shown the capacity to activate NADPH oxidase in various tissues. Recent evidence suggests that cadmium (Cd), a key toxin in cigarette smoke, may play a pathogenic role in AMD, consistent with the protective effect of the physiological Cd antagonist zinc in the AREDS1 study; Cd likewise has the potential to activate NADPH oxidase. Retinal pigmented epithelial (RPE) cells express NADPH oxidase activity, and oxidative stress within these cells is suspected to play a mediating role in the pathogenesis of AMD. It is therefore proposed that acrolein, Cd, and C5a all promote activation of NADPH oxidase in RPE cells, and that this phenomenon is a major mediator of the adverse impact of smoking and of the alternative complement pathway on AMD risk. If this hypothesis is correct, increased serum levels of free bilirubin – which functions physiologically to inhibit NADPH oxidase – may be associated with decreased risk for AMD; this prediction is readily testable. Moreover, this hypothesis suggests that spirulina, whose chromophore phycocyanobilin can mimic the NADPH oxidase-inhibitory activity of its chemical relative bilirubin, may have utility for prevention and control of AMD. A corollary of these considerations is that dietary spirulina and supplemental zinc may have potential for alleviating the adverse impact of smoking – and of Cd exposure, to which smoking importantly contributes - on vascular and overall health.

Cigarette Smoke May Promote AMD via NADPH Oxidase Activity in RPE Cells

The adverse effects of smoking on vascular health – linked to greater total mortality than smoking's impact on cancer risk – appear to be mediated primarily, not by nicotine or tars, but by relatively stable organic compounds in the smoke capable of undergoing spontaneous addition reactions with thiols and other nucleophiles within vascular cells.¹ Notably, these compounds include alpha,beta-unsaturated aldehydes or ketones such as acrolein and crotonaldehyde, major constituents of cigarette smoke. A number of studies have observed that exposure of endothelial or vascular smooth muscle cells, *in vitro* or *in vivo*, to cigarette smoke extract (CSE) rich in such compounds leads to induction of oxidative stress generated primarily by NADPH oxidase complexes.¹⁻⁷ Concurrent inhibition of these complexes largely alleviates the adverse effects of CSE exposure on endothelial or smooth muscle function. CSE exposure has likewise been shown to boost NADPH oxidase activity in airway epithelia, tracheal smooth muscle, keratinocytes, and glioma cells.⁸⁻¹⁶ Activation of various PKC isoforms, and/or c-Src, by the reactive compounds in CSE, appears to mediate NADPH oxidase activation; activation of PKC does not appear to be secondary to activation of phospholipase C and diacylglycerol generation, and so might reflect direct interaction with PKC protein.¹⁵

It is well documented that smokers are at greatly increased risk for both the dry and wet forms of age-related macular degeneration (AMD).^{17, 18} There is also substantial reason to believe that oxidative stress within retinal pigment epithelial (RPE) cells plays a key role in the pathogenesis of both forms of AMD;¹⁹⁻²² however, few reports have attempted to assess what the primary source of this oxidative stress may be. RPE cells function as phagocytes, engulfing and degrading photoreceptor outer segments; like other phagocytic cells, they express NADPH oxidase activity.²³⁻²⁵ Moreover, in a mouse model of AMD evoked by laser disruption of Bruch's membrane, viral delivery of small interfering RNA for p22phox to the subretinal space prevents choroidal neovascularization – leading the authors to comment that “NADPH oxidase-mediated ROS production in RPE cells may play an important role in the genesis of neovascular AMD, and this pathway may represent a new target for therapeutic intervention in AMD.”²⁵ Exposure of RPE cells to CSE has been shown to promote oxidative stress in these cells that can induce VEGF expression (a key mediator of neovascular AMD) and also lead to apoptotic cell death; exogenous antioxidants such as N-acetylcysteine are protective in these respects.²⁶⁻³¹ However, how CSE evokes this oxidative stress in RPE cells has not been clarified. In light of the impact of CSE on vascular tissue, it is very reasonable to speculate that this oxidative stress stems primarily from activation of NADPH oxidase.

Cadmium May Also Activate NADPH Oxidase in RPE

High-normal body levels of the toxic heavy metal cadmium (Cd) have been linked to increased risk for vascular disease, osteoporosis, nephropathy, various cancers, and total mortality, and there is ample reason to suspect that these associations are causative.³²⁻³⁷ Smoking is a key source of Cd exposure, and the body Cd stores of long-time smokers tend to be about twice as high as those of non-smokers. The possibility that Cd accumulation in the retina contributes to the pathogenesis of AMD is suggested by several recent studies. The Cd content of the retina, retinal pigmented epithelium, and the aqueous humor is increased in eyes afflicted with AMD.^{38, 39} In a case-control study, urinary levels of Cd (considered a reasonably accurate measure of total body stores) of smokers with AMD were found to be significantly higher than those of smokers without this disorder; in non-smokers, however, urinary Cd did not differ between cases and controls.⁴⁰

Arguably, these finding might simply reflect the fact that Cd is serving as a marker for intensity of smoking exposure, which mediates AMD by other means. But a pathogenic role for Cd in this regard is also credible. Induction of oxidative stress is believed to be a key mechanism whereby Cd impairs health.⁴¹ Urinary Cd levels correlate strongly with urine levels of 8-oxo-7,8-dihydro-2'-deoxyguanosine, a standard marker for oxidative damage to DNA.⁴² In cultured hepatocytes or neurons exposed acutely to low micromolar levels of Cd, NADPH oxidase is strongly activated.⁴³⁻⁴⁵ Moreover, the hypertensive response of rats to chronic administration of Cd in drinking water is substantially blunted when the drug apocynin, which functions to inhibit activation of NADPH oxidase *in vivo*, is administered.⁴⁶ And oral administration of spirulina – which can function as an NADPH oxidase antagonist, as discussed below – provides dose-dependent protection from the teratogenicity of Cd in mice.⁴⁷ Cd exposure *in vitro* does indeed generate oxidative stress in RPE cells, though the source of this stress has not yet been defined.⁴⁸ These findings are consistent with the possibility that Cd exposure promotes AMD by up-regulating NADPH oxidase-mediated oxidative stress in RPE cells.

The mildly favorable results of the AREDS1 study of nutritional supplementation for control of AMD might also be viewed as consistent with this hypothesis. Although Cd has a very long half-life (estimated

as 10-30 years) in the body, and no effective chelation strategies for accelerating its removal are available, the pathogenic impact of the Cd already in the body can be mitigated to some degree by induction of the protein metallothionein.⁴⁹ This zinc-inducible protein functions to bind and store zinc and copper ions, but its numerous sulfhydryl groups can also bind Cd ions; in this bound form, Cd appears to be relatively benign. Supplemental zinc intakes in the range of 15-50 daily have been reported to boost the metallothionein content of the monocytes and erythrocytes of young men.^{50, 51} The nutritional supplementation tested in the AREDS1 study included an arm in which patients received 80 mg of zinc daily. Risk of progressing to advanced AMD during the study was significantly lower in those subjects received zinc plus antioxidants (20%) was significantly lower than that in this placebo group – but the risk in those receiving zinc alone was only slightly higher (22%).⁵² More remarkably, the total mortality over 6.5 years of follow-up was significantly lower in subjects randomized to receive zinc (RR, 0.73; 95% CI, 0.61-0.89)⁵³ - particularly intriguing in light of evidence that increased Cd stores correlate with increased total mortality. Other recent research suggests that, in subjects with relatively good zinc status, the correlation between body Cd and cancer mortality is attenuated.³⁷

Genetic Up-Regulation of Complement Activation May Promote NADPH Oxidase Activity in RPE

AMD is known to be considerably more common in individuals who express alleles of complement factors or complement-related factors which could be expected to up-regulate activation of the alternative complement pathway.⁵⁴⁻⁵⁷ Complement metabolites are a prominent component of drusen, and RPE cells express receptors for the pro-inflammatory complement metabolites C3a and C5a; moreover, oxidative stress in RPE cells impairs their ability to express inhibitors of complement activation on their plasma membranes.⁵⁸⁻⁶⁰ Exposure of RPE cells to C5a boosts their expression of VEGF.⁶⁰ Increased plasma levels of C5a are associated with increased risk for AMD.⁶¹ In aggregate, these findings strongly suggest that activation of the alternative complement pathway on RPE cells plays a pathogenic role in AMD. In neutrophils, eosinophils, and macrophages, which also express C5a receptors, C5a stimulates NADPH oxidase activity, likely via G protein-triggered PLC-beta/PKC signaling.⁶²⁻⁶⁸ It would be of great interest to know whether C5a likewise can activate the NADPH oxidase in RPE cells; notably, the requisite G proteins, PLC-beta, and PKCs are expressed in these cells.^{69, 70}

Antioxidant Activity of Spirulina May Have Potential for Prevention and Control of AMD

It should be relatively easy to determine whether CSE, Cd, and/or C5a can activate NADPH oxidase in RPE cells. A resolution of this issue could be of some practical importance. Recent studies reveal that free bilirubin – generated within cells by heme oxygenase activity – functions physiologically as a feedback inhibitor of NADPH oxidase complexes.⁷¹⁻⁷³ Although bilirubin is highly insoluble and not readily available for clinical use, its more soluble precursor has more clinical potential. But a more practical alternative in this regard may be offered by the phycobilins – notably phycocyanobilin (PhyCB) – that are major components of cyanobacteria such as spirulina. PhyCB, a derivative and homolog of biliverdin, has recently been shown to mimic the impact of bilirubin/biliverdin on NADPH oxidase activity – likely accounting for the profound and versatile anti-inflammatory effects of oral spirulina (or of phycocyanin, the prominent spirulina protein which contains PhyCB as a chromophore) in rodent studies.⁷⁴⁻⁷⁶ Hence, if NADPH oxidase mediates much of the adverse impact of CSE, Cd, and/or complement activation on RPE cells, supplementation with spirulina or spirulina extracts may have important potential for the prevention or management of AMD.

A number of studies have linked increased serum bilirubin levels to decreased risk for vascular disorders.^{77, 78} In light of the foregoing speculations, it would be intriguing to see whether individuals with relatively high bilirubin levels might be at decreased risk for AMD.

A corollary of these considerations is that supplementation with spirulina – and with zinc, a physiological Cd antagonist - may have important potential for alleviating the adverse impact of smoking on vascular and perhaps overall health. Notably, spirulina may provide some protection for the lungs of smokers – consistent with recent evidence that people with relatively high bilirubin levels are at decreased risk for both lung cancer and chronic obstructive pulmonary disease.⁷⁹

References

- (1) Jaimes EA, DeMaster EG, Tian RX, Raji L. Stable compounds of cigarette smoke induce endothelial superoxide anion production via NADPH oxidase activation. *Arterioscler Thromb Vasc Biol* 2004 June;24(6):1031-6.
- (2) Orosz Z, Csiszar A, Labinsky N et al. Cigarette smoke-induced proinflammatory alterations in the endothelial phenotype: role of NAD(P)H oxidase activation. *Am J Physiol Heart Circ Physiol* 2007 January;292(1):H130-H139.
- (3) Shih RH, Lee IT, Hsieh HL, Kou YR, Yang CM. Cigarette smoke extract induces HO-1 expression in mouse cerebral vascular endothelial cells: involvement of c-Src/NADPH oxidase/PDGFR/JAK2/STAT3 pathway. *J Cell Physiol* 2010 November;225(3):741-50.
- (4) Shih RH, Cheng SE, Hsiao LD, Kou YR, Yang CM. Cigarette smoke extract upregulates heme oxygenase-1 via PKC/NADPH oxidase/ROS/PDGFR/PI3K/Akt pathway in mouse brain endothelial cells. *J Neuroinflammation* 2011;8:104.
- (5) Barbieri SS, Amadio P, Gianellini S, Zacchi E, Weksler BB, Tremoli E. Tobacco smoke regulates the expression and activity of microsomal prostaglandin E synthase-1: role of prostacyclin and NADPH-oxidase. *FASEB J* 2011 October;25(10):3731-40.
- (6) Mo Y, Wan R, Feng L, Chien S, Tollerud DJ, Zhang Q. Combination effects of cigarette smoke extract and ambient ultrafine particles on endothelial cells. *Toxicol In Vitro* 2012 March;26(2):295-303.
- (7) Steffen Y, Vuillaume G, Stolle K et al. Cigarette smoke and LDL cooperate in reducing nitric oxide bioavailability in endothelial cells via effects on both eNOS and NADPH oxidase. *Nitric Oxide* 2012 October 15;27(3):176-84.
- (8) Lavigne MC, Eppihimer MJ. Cigarette smoke condensate induces MMP-12 gene expression in airway-like epithelia. *Biochem Biophys Res Commun* 2005 April 29;330(1):194-203.

- (9) Cheng SE, Luo SF, Jou MJ et al. Cigarette smoke extract induces cytosolic phospholipase A2 expression via NADPH oxidase, MAPKs, AP-1, and NF-kappaB in human tracheal smooth muscle cells. *Free Radic Biol Med* 2009 April 1;46(7):948-60.
- (10) Lin CC, Lee IT, Yang YL, Lee CW, Kou YR, Yang CM. Induction of COX-2/PGE(2)/IL-6 is crucial for cigarette smoke extract-induced airway inflammation: Role of TLR4-dependent NADPH oxidase activation. *Free Radic Biol Med* 2010 January 15;48(2):240-54.
- (11) Cheng SE, Lee IT, Lin CC, Kou YR, Yang CM. Cigarette smoke particle-phase extract induces HO-1 expression in human tracheal smooth muscle cells: role of the c-Src/NADPH oxidase/MAPK/Nrf2 signaling pathway. *Free Radic Biol Med* 2010 May 15;48(10):1410-22.
- (12) Cheng SE, Lin CC, Lee IT, Hsu CK, Kou YR, Yang CM. Cigarette smoke extract regulates cytosolic phospholipase A2 expression via NADPH oxidase/MAPKs/AP-1 and p300 in human tracheal smooth muscle cells. *J Cell Biochem* 2011 February;112(2):589-99.
- (13) Talbot S, Lin JC, Lahjouji K et al. Cigarette smoke-induced kinin B1 receptor promotes NADPH oxidase activity in cultured human alveolar epithelial cells. *Peptides* 2011 July;32(7):1447-56.
- (14) Sticozzi C, Belmonte G, Pecorelli A et al. Cigarette smoke affects keratinocytes SRB1 expression and localization via H2O2 production and HNE protein adducts formation. *PLoS ONE* 2012;7(3):e33592.
- (15) Asano H, Horinouchi T, Mai Y et al. Nicotine- and tar-free cigarette smoke induces cell damage through reactive oxygen species newly generated by PKC-dependent activation of NADPH oxidase. *J Pharmacol Sci* 2012;118(2):275-87.
- (16) Mai Y, Higashi T, Terada K et al. Nicotine- and Tar-free Cigarette Smoke Extract Induces Cell Injury via Intracellular Ca(2+)-Dependent Subtype-Specific Protein Kinase C Activation. *J Pharmacol Sci* 2012 November 13.
- (17) Thornton J, Edwards R, Mitchell P, Harrison RA, Buchan I, Kelly SP. Smoking and age-related macular degeneration: a review of association. *Eye (Lond)* 2005 September;19(9):935-44.
- (18) Khan JC, Thurlby DA, Shahid H et al. Smoking and age related macular degeneration: the number of pack years of cigarette smoking is a major determinant of risk for both geographic atrophy and choroidal neovascularisation. *Br J Ophthalmol* 2006 January;90(1):75-80.
- (19) Beatty S, Koh H, Phil M, Henson D, Boulton M. The role of oxidative stress in the pathogenesis of age-related macular degeneration. *Surv Ophthalmol* 2000 September;45(2):115-34.
- (20) Beatty S, Koh H, Phil M, Henson D, Boulton M. The role of oxidative stress in the pathogenesis of age-related macular degeneration. *Surv Ophthalmol* 2000 September;45(2):115-34.
- (21) Cai J, Nelson KC, Wu M, Sternberg P, Jr., Jones DP. Oxidative damage and protection of the RPE. *Prog Retin Eye Res* 2000 March;19(2):205-21.
- (22) Imamura Y, Noda S, Hashizume K et al. Drusen, choroidal neovascularization, and retinal pigment epithelium dysfunction in SOD1-deficient mice: a model of age-related macular degeneration. *Proc Natl Acad Sci U S A* 2006 July 25;103(30):11282-7.

- (23) Miceli MV, Liles MR, Newsome DA. Evaluation of oxidative processes in human pigment epithelial cells associated with retinal outer segment phagocytosis. *Exp Cell Res* 1994 September;214(1):242-9.
- (24) Tate DJ, Jr., Miceli MV, Newsome DA. Phagocytosis and H₂O₂ induce catalase and metallothionein gene expression in human retinal pigment epithelial cells. *Invest Ophthalmol Vis Sci* 1995 June;36(7):1271-9.
- (25) Li Q, Dinculescu A, Shan Z et al. Downregulation of p22phox in Retinal Pigment Epithelial Cells Inhibits Choroidal Neovascularization in Mice. *Mol Ther* 2008 July 29.
- (26) Espinosa-Heidmann DG, Suner IJ, Catanuto P, Hernandez EP, Marin-Castano ME, Cousins SW. Cigarette smoke-related oxidants and the development of sub-RPE deposits in an experimental animal model of dry AMD. *Invest Ophthalmol Vis Sci* 2006 February;47(2):729-37.
- (27) Jia L, Liu Z, Sun L et al. Acrolein, a toxicant in cigarette smoke, causes oxidative damage and mitochondrial dysfunction in RPE cells: protection by (R)-alpha-lipoic acid. *Invest Ophthalmol Vis Sci* 2007 January;48(1):339-48.
- (28) Fujihara M, Nagai N, Sussan TE, Biswal S, Handa JT. Chronic cigarette smoke causes oxidative damage and apoptosis to retinal pigmented epithelial cells in mice. *PLoS ONE* 2008;3(9):e3119.
- (29) Bertram KM, Baglolle CJ, Phipps RP, Libby RT. Molecular regulation of cigarette smoke induced-oxidative stress in human retinal pigment epithelial cells: implications for age-related macular degeneration. *Am J Physiol Cell Physiol* 2009 November;297(5):C1200-C1210.
- (30) Pons M, Marin-Castano ME. Cigarette smoke-related hydroquinone dysregulates MCP-1, VEGF and PEDF expression in retinal pigment epithelium in vitro and in vivo. *PLoS ONE* 2011;6(2):e16722.
- (31) Yu AL, Birke K, Burger J, Welge-Lussen U. Biological effects of cigarette smoke in cultured human retinal pigment epithelial cells. *PLoS ONE* 2012;7(11):e48501.
- (32) Everett CJ, Frithsen IL. Association of urinary cadmium and myocardial infarction. *Environ Res* 2008 February;106(2):284-6.
- (33) Menke A, Muntner P, Silbergeld EK, Platz EA, Guallar E. Cadmium levels in urine and mortality among U.S. adults. *Environ Health Perspect* 2009 February;117(2):190-6.
- (34) Jarup L, Akesson A. Current status of cadmium as an environmental health problem. *Toxicol Appl Pharmacol* 2009 August 1;238(3):201-8.
- (35) Nawrot TS, Staessen JA, Roels HA et al. Cadmium exposure in the population: from health risks to strategies of prevention. *Biometals* 2010 October;23(5):769-82.
- (36) McCarty MF. Zinc and multi-mineral supplementation should mitigate the pathogenic impact of cadmium exposure. *Med Hypotheses* 2012 November;79(5):642-8.
- (37) Lin YS, Caffrey JL, Lin JW et al. Increased risk of cancer mortality associated with cadmium exposures in older Americans with low zinc intake. *J Toxicol Environ Health A* 2013;76(1):1-15.

- (38) Wills NK, Kalariya N, Sadagopa Ramanujam VM et al. Human retinal cadmium accumulation as a factor in the etiology of age-related macular degeneration. *Exp Eye Res* 2009 June 15;89(1):79-87.
- (39) Junemann AG, Stopa P, Michalke B et al. Levels of aqueous humor trace elements in patients with non-exsudative age-related macular degeneration: a case-control study. *PLoS ONE* 2013;8(2):e56734.
- (40) Erie JC, Good JA, Butz JA, Hodge DO, Pulido JS. Urinary cadmium and age-related macular degeneration. *Am J Ophthalmol* 2007 September;144(3):414-8.
- (41) Cuypers A, Plusquin M, Remans T et al. Cadmium stress: an oxidative challenge. *Biometals* 2010 October;23(5):927-40.
- (42) Engstrom KS, Vahter M, Johansson G et al. Chronic exposure to cadmium and arsenic strongly influences concentrations of 8-oxo-7,8-dihydro-2'-deoxyguanosine in urine. *Free Radic Biol Med* 2010 May 1;48(9):1211-7.
- (43) Souza V, Escobar MC, Bucio L, Hernandez E, Gomez-Quiroz LE, Gutierrez Ruiz MC. NADPH oxidase and ERK1/2 are involved in cadmium induced-STAT3 activation in HepG2 cells. *Toxicol Lett* 2009 June 22;187(3):180-6.
- (44) Chen L, Xu B, Liu L et al. Cadmium induction of reactive oxygen species activates the mTOR pathway, leading to neuronal cell death. *Free Radic Biol Med* 2011 March 1;50(5):624-32.
- (45) Martinez FK, Uribe Marin BC, Souza A, V et al. Hepatocytes display a compensatory survival response against cadmium toxicity by a mechanism mediated by EGFR and Src. *Toxicol In Vitro* 2013 April;27(3):1031-42.
- (46) Nwokocha CR, Baker A, Douglas D, McCalla G, Nwokocha M, Brown PD. Apocynin Ameliorates Cadmium-Induced Hypertension Through Elevation of Endothelium Nitric Oxide Synthase. *Cardiovasc Toxicol* 2013 May 24.
- (47) Paniagua-Castro N, Escalona-Cardoso G, Hernandez-Navarro D, Perez-Pasten R, Chamorro-Cevallos G. Spirulina (Arthrospira) protects against cadmium-induced teratogenic damage in mice. *J Med Food* 2011 April;14(4):398-404.
- (48) Kalariya NM, Wills NK, Ramana KV, Srivastava SK, van Kuijk FJ. Cadmium-induced apoptotic death of human retinal pigment epithelial cells is mediated by MAPK pathway. *Exp Eye Res* 2009 October;89(4):494-502.
- (49) Klaassen CD, Liu J, Diwan BA. Metallothionein protection of cadmium toxicity. *Toxicol Appl Pharmacol* 2009 August 1;238(3):215-20.
- (50) Sullivan VK, Burnett FR, Cousins RJ. Metallothionein expression is increased in monocytes and erythrocytes of young men during zinc supplementation. *J Nutr* 1998 April;128(4):707-13.
- (51) Park SK, Jung IC, Lee WK et al. A combination of green tea extract and l-theanine improves memory and attention in subjects with mild cognitive impairment: a double-blind placebo-controlled study. *J Med Food* 2011 April;14(4):334-43.

- (52) A randomized, placebo-controlled, clinical trial of high-dose supplementation with vitamins C and E, beta carotene, and zinc for age-related macular degeneration and vision loss: AREDS report no. 8. *Arch Ophthalmol* 2001 October;119(10):1417-36.
- (53) Clemons TE, Kurinij N, Sperduto RD. Associations of mortality with ocular disorders and an intervention of high-dose antioxidants and zinc in the Age-Related Eye Disease Study: AREDS Report No. 13. *Arch Ophthalmol* 2004 May;122(5):716-26.
- (54) Anderson DH, Radeke MJ, Gallo NB et al. The pivotal role of the complement system in aging and age-related macular degeneration: hypothesis re-visited. *Prog Retin Eye Res* 2010 March;29(2):95-112.
- (55) Despriet DD, Klaver CC, Witteman JC et al. Complement factor H polymorphism, complement activators, and risk of age-related macular degeneration. *JAMA* 2006 July 19;296(3):301-9.
- (56) Despriet DD, van Duijn CM, Oostra BA et al. Complement component C3 and risk of age-related macular degeneration. *Ophthalmology* 2009 March;116(3):474-80.
- (57) Donoso LA, Vrabec T, Kuivaniemi H. The role of complement Factor H in age-related macular degeneration: a review. *Surv Ophthalmol* 2010 May;55(3):227-46.
- (58) Nozaki M, Raisler BJ, Sakurai E et al. Drusen complement components C3a and C5a promote choroidal neovascularization. *Proc Natl Acad Sci U S A* 2006 February 14;103(7):2328-33.
- (59) Cortright DN, Meade R, Waters SM, Chenard BL, Krause JE. C5a, but not C3a, increases VEGF secretion in ARPE-19 human retinal pigment epithelial cells. *Curr Eye Res* 2009 January;34(1):57-61.
- (60) Thurman JM, Renner B, Kunchithapautham K et al. Oxidative stress renders retinal pigment epithelial cells susceptible to complement-mediated injury. *J Biol Chem* 2009 June 19;284(25):16939-47.
- (61) Reynolds R, Hartnett ME, Atkinson JP, Giclas PC, Rosner B, Seddon JM. Plasma complement components and activation fragments: associations with age-related macular degeneration genotypes and phenotypes. *Invest Ophthalmol Vis Sci* 2009 December;50(12):5818-27.
- (62) Bender JG, McPhail LC, Van Epps DE. Exposure of human neutrophils to chemotactic factors potentiates activation of the respiratory burst enzyme. *J Immunol* 1983 May;130(5):2316-23.
- (63) Dewald B, Thelen M, Baggiolini M. Two transduction sequences are necessary for neutrophil activation by receptor agonists. *J Biol Chem* 1988 November 5;263(31):16179-84.
- (64) Torres M, Forman HJ. Activation of several MAP kinases upon stimulation of rat alveolar macrophages: role of the NADPH oxidase. *Arch Biochem Biophys* 1999 June 15;366(2):231-9.
- (65) DiScipio RG, Schraufstatter IU, Sikora L, Zuraw BL, Sriramarao P. C5a mediates secretion and activation of matrix metalloproteinase 9 from human eosinophils and neutrophils. *Int Immunopharmacol* 2006 July;6(7):1109-18.
- (66) Kato M, Yamaguchi T, Tachibana A et al. An atypical protein kinase C, PKC zeta, regulates human eosinophil effector functions. *Immunology* 2005 October;116(2):193-202.

- (67) Wymann MP, Kernen P, Von T, V, Tai PC, Spry CJ, Baggiolini M. Activation of the respiratory burst in eosinophil leucocytes--a transduction sequence decoupled from cytosolic Ca²⁺ rise. *Eur J Clin Invest* 1995 January;25(1):25-31.
- (68) Jiang H, Kuang Y, Wu Y, Smrcka A, Simon MI, Wu D. Pertussis toxin-sensitive activation of phospholipase C by the C5a and fMet-Leu-Phe receptors. *J Biol Chem* 1996 June 7;271(23):13430-4.
- (69) Jiang M, Pandey S, Tran VT, Fong HK. Guanine nucleotide-binding regulatory proteins in retinal pigment epithelial cells. *Proc Natl Acad Sci U S A* 1991 May 1;88(9):3907-11.
- (70) Palma-Nicolas JP, Lopez E, Lopez-Colome AM. PKC isoenzymes differentially modulate the effect of thrombin on MAPK-dependent RPE proliferation. *Biosci Rep* 2008 December;28(6):307-17.
- (71) Lanone S, Bloc S, Foresti R et al. Bilirubin decreases nos2 expression via inhibition of NAD(P)H oxidase: implications for protection against endotoxic shock in rats. *FASEB J* 2005 November;19(13):1890-2.
- (72) Matsumoto H, Ishikawa K, Itabe H, Maruyama Y. Carbon monoxide and bilirubin from heme oxygenase-1 suppresses reactive oxygen species generation and plasminogen activator inhibitor-1 induction. *Mol Cell Biochem* 2006 October;291(1-2):21-8.
- (73) Jiang F, Roberts SJ, Datla S, Dusting GJ. NO modulates NADPH oxidase function via heme oxygenase-1 in human endothelial cells. *Hypertension* 2006 November;48(5):950-7.
- (74) McCarty MF. Clinical potential of Spirulina as a source of phycocyanobilin. *J Med Food* 2007 December;10(4):566-70.
- (75) Zheng J, Inoguchi T, Sasaki S et al. Phycocyanin and Phycocyanobilin from Spirulina Platensis Protect against Diabetic Nephropathy by Inhibiting Oxidative Stress. *Am J Physiol Regul Integr Comp Physiol* 2012 October 31.
- (76) Romay C, Gonzalez R, Ledon N, Ramirez D, Rimbau V. C-phycocyanin: a biliprotein with antioxidant, anti-inflammatory and neuroprotective effects. *Curr Protein Pept Sci* 2003 June;4(3):207-16.
- (77) Lin JP, Vitek L, Schwertner HA. Serum bilirubin and genes controlling bilirubin concentrations as biomarkers for cardiovascular disease. *Clin Chem* 2010 October;56(10):1535-43.
- (78) Horsfall LJ, Nazareth I, Petersen I. Cardiovascular events as a function of serum bilirubin levels in a large, statin-treated cohort. *Circulation* 2012 November 27;126(22):2556-64.
- (79) Horsfall LJ, Rait G, Walters K et al. Serum bilirubin and risk of respiratory disease and death. *JAMA* 2011 February 16;305(7):691-7.