

AMPK Activation – Protean Potential for Boosting Healthspan

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Abstract

AMP-activated kinase (AMPK) is activated when the cellular (AMP+ADP)/ATP ratio rises; it therefore serves as a detector of cellular “fuel deficiency”. AMPK activation is suspected to mediate some of the health-protective effects of long term calorie restriction. Several drugs and nutraceuticals which slightly and safely impede the efficiency of mitochondrial ATP generation – most notably metformin and berberine – can be employed as clinical AMPK activators, and hence have been suggested to have potential as calorie restriction mimetics for extending healthspan. In fact, an overview of current evidence indicates that AMPK activators may: reduce risk for atherosclerosis, heart attack, and stroke; help to prevent ventricular hypertrophy and manage congestive failure; ameliorate metabolic syndrome, reduce risk for type 2 diabetes, and aid glycemic control in diabetics; reduce risk for weight gain; decrease risk for a number of common cancers while improving prognosis in cancer therapy; decrease risk for dementia and possibly other neurodegenerative disorders; help to preserve the proper structure of bone and cartilage; and possibly aid in the prevention and control of autoimmunity. It is therefore reasonable to speculate that long term use of AMPK activators may indeed have a notably favorable impact on healthspan, and that such agents could reasonably be employed as adjuvants to lifestyle strategies – such as modified alternate-day fasting or carbohydrate-concentrated diets – intended to replicate the benefits of daily calorie restriction. While metformin and berberine appear to have the greatest utility as clinical AMPK activators – as reflected by their efficacy in diabetes management – regular ingestion of vinegar, as well as moderate alcohol consumption, may also achieve a modest degree of health-protective AMPK activation.

AMPK – Cellular Monitor of Fuel Availability

AMP-activated kinase (AMPK) is sometimes described as the “fuel gauge” of the cell, inasmuch as it is activated by an increase in the cellular ratio of AMP+ADP to ATP.¹⁻³ AMPK is a heterotrimer, consisting of α , β , and γ subunits. The α subunit develops serine/threonine kinase activity when its Thr172 is phosphorylated by various upstream kinases. The γ subunit has a regulatory function; it possesses 2 sites each of which can bind either AMP, ADP, or ATP. At one of these sites, AMP binding allosterically boosts the kinase activity of the activated enzyme. At the other site, binding of either AMP or ADP suppresses the ability of phosphatases to remove the phosphate from Thr172 and hence deactivate the enzyme.¹ The chief activating upstream kinases targeting AMPK are LKB1, which is constitutively active and widely expressed, and the calmodulin-dependent kinase kinases (CaMKK), whose activity is stimulated by an increase in free intracellular calcium. Acute activation of AMPK is therefore seen when the (AMP+ADP)/ATP ratio rises, or when intracellular free calcium rises – conditions which often signal cellular stress. Modulation of the activity of the phosphatases which target Thr172 of the α subunit – PP2A and PP2C – can also influence AMPK activity.⁴

A number of drugs, phytochemicals, and hormones have the potential to activate AMPK. Many of these agents boost the (AMP+ADP)/ATP ratio by impeding the efficiency of mitochondrial electron transport

(metformin, berberine, thiazolidinediones, dinitrophenol), inhibiting mitochondrial ATP synthase (resveratrol, quercetin) or inhibiting glycolysis (2-deoxyglucose).³ Hormones and drugs which increase intracellular free calcium can also function as AMPK activators; for example, the ability of HDL particles to activate AMPK in endothelial cells reflects, in part, an increase in calcium influx that activates CaMKK.⁵ The natural metabolite AICAR is converted intracellularly to an analog of AMP which mimics AMP's activating impact on AMPK. And pharmaceutical companies are striving to develop drugs – of which A769662 is a prototype – which likewise can activate AMPK by binding to it directly.⁶ The ability of the hormone adiponectin to activate AMPK has recently been traced to the ceramidase activity of the activated adiponectin receptor; ceramide suppresses AMPK activity by activating PP2A.^{4,7} Analogously, lipoic acid has the potential to support AMPK activity by countering oxidant-mediated activation of neutral sphingomyelinase, whose product is ceramide.^{8,9}

Since AMPK detects cellular energy deficit, its *raison d'être* is to boost the capacity of cells to generate ATP via substrate oxidation while simultaneously suppressing the activity of metabolic pathways which utilize ATP.¹⁰ Hence, AMPK boosts mitochondrial biogenesis, aids mitochondrial antioxidant protection, and increases expression and activity of glucose transporters and glycolytic enzymes; concurrently, non-essential synthesis of proteins, lipids, and carbohydrates is decreased.¹⁰⁻¹² However, some key effects of AMPK – such as activation of the endothelial nitric oxide synthase (eNOS)^{13,14} – do not readily fit into this paradigm.

Are AMPK Activators Calorie Restriction Mimetics?

AMPK has been conserved throughout the evolution of eukaryotes, and there is evidence that activation of AMPK plays an obligate role in the life-extending activity of caloric restriction in lower eukaryotes such as yeast, worms, and flies.¹⁵ Evidence is currently conflicting as to whether the fuel deficit stress induced by calorie restriction regimens in rodents is sufficient to activate AMPK in rodents.¹⁵ Such activation may however occur indirectly via up-regulation of adiponectin production in adipose tissue.¹⁶ In any case, there is good reason to suspect that chronic activation of AMPK can serve as a calorie restriction mimetic in mammals. AMPK boosts the activity of Sirt1, another evolutionarily-conserved enzyme which likewise is a mediator of the life-prolonging impact of calorie restriction in lower eukaryotes; AMPK does so by somehow increasing the expression of nicotinamide phosphoribosyl transferase (NAMPT), which is rate-limiting for the regeneration of Sirt1's obligate cofactor NAD⁺.^{17,18} In light of the media hysteria generated recently by reports that the wine phytochemical resveratrol may exert a pro-longevity effect by activating Sirt1, it should be noted that this activation now appears to be indirect, mediated via resveratrol's impact on AMPK.^{19,20} (And, ironically, pharmacokinetic studies show that absorbed resveratrol is conjugated so rapidly in humans that it has little clinical potential as an AMPK activator.^{21,22})

AMPK mimics the impact of the growth factor down-regulation associated with calorie restriction by inhibiting activity of the mammalian target of rapamycin complex 1 (mTORC1).²³ This complex, via phosphorylation of its targets p70 ribosomal S6 kinase 1 (S6K1) and 4EBP1, up-regulates protein synthesis, and plays a key role in cell proliferation; its activity is suppressed by calorie restriction.^{24,25} AMPK can also phosphorylate and thereby boost the transcriptional activity of FOXO3a, which induces expression of a number of antioxidant enzymes and other stress-resistance proteins.^{12,26,27}

By inhibiting mTORC1 activity, and also via direct phosphorylation, AMPK stimulates ULK1, a kinase which is a key trigger for the process of macroautophagy.²⁸ This “cell cleansing” process is also abetted by Sirt1 and FOXO activity.²⁹ Since macroautophagy rids the cell of aging, potentially pro-oxidative mitochondria, it is homeostatically appropriate that AMPK, Sirt1, and FOXO3a also collaborate in promoting mitochondrial biogenesis, largely by boosting the expression and pro-transcriptional activity of PPAR- γ coactivator-1 α (PCG-1 α).^{11, 30} The efficiency of macroautophagy declines with increasing age in mammals, and there is considerable speculation that the up-regulation of macroautophagy evoked by calorie restriction in many tissues contributes crucially to the life prolongation and “aging-retardation” achieved by such restriction.^{31, 32} Indeed, concurrent up-regulation of macroautophagy, mitochondrial biogenesis and expression of mitochondrial antioxidant proteins – as promoted by AMPK activity – could be expected to keep cells structurally and functionally pristine, and is emerging as a central motif in the pro-longevity impact of calorie restriction. Not surprisingly, treatment of mice with the AMPK-activating drug metformin has been shown to replicate a number of the effects of long-term calorie restriction on hepatic gene expression.³³

At a systemic level, AMPK activation can modestly lower serum glucose and insulin levels by down-regulating hepatic gluconeogenesis (as discussed below) and hence slowing hepatic glucose output; this reduction in insulin can be expected to decrease IGF-I bioactivity by up-regulating hepatic production of IGFBP-1.³⁴⁻³⁶ These effects are similar to the impact of calorie restriction on serum levels of glucose, insulin, and free IGF-I – though more modest in magnitude.

Down-regulation of insulin/IGF-I signaling (or the homologous pathways in lower organisms) is thought to be the key mediator of the lifespan extension associated with calorie restriction in eukaryotes.³⁷ Such signaling activates mTORC1 and S6K1, while inhibiting FOXO activity; AMPK has a countervailing impact in these regards. Agents or measures which inhibit mTORC1 or S6K1, or which activate AMPK, have indeed been reported to increase mean and maximal lifespan in certain strains of rodents.³⁸⁻⁴² However, the survival-prolonging impact of AMPK activators metformin and resveratrol has not been observed in some healthy rodent strains fed healthful diets – whereas it is more notable in cancer-prone strains or in rodents fed diets that induce obesity and insulin resistance.^{43, 44} Nonetheless, an improvement in markers for healthspan has been observed in resveratrol-treated mice even when lifespan has not been influenced.⁴⁴ Hence, although it seems unlikely that AMPK activation can replicate the full lifespan-lengthening impact of calorie restriction, it may have considerable potential for promoting increased *healthspan*, and arguably might be able to amplify the longevity benefits of modest degrees of daily calorie restriction or of more practical dietary strategies (e.g. modified alternate-day fasting, carbohydrate-concentrated diets) that episodically minimize serum levels of glucose, insulin, and free IGF-I.⁴⁵⁻⁴⁸

Indeed, independent of any impact of AMPK on the aging process per se, there is considerable reason to suspect that agents which can safely activate AMPK in humans – most notably metformin and berberine⁴⁹⁻⁵² – have the potential to boost healthspan via a bewildering variety of protective effects: preventing atherosclerosis, heart attack, and stroke; preventing cardiac hypertrophy and aiding management of congestive failure; ameliorating metabolic syndrome while antagonizing weight gain; reducing risk for type 2 diabetes and aiding its metabolic control; reducing risk for many cancers and improving the outcome of cancer therapies; postponing or preventing onset of dementia and possibly other neurodegenerative disorders; aiding preservation of cartilage and of bone density; and reducing risk

for, or aiding control of, autoimmune disorders. This rather audacious claim is supported, in part, by epidemiology and clinical trials focusing on metformin use in diabetics; other pertinent support comes from rodent and cell culture studies.

Vascular Protection

The first strong clue that metformin might have unusual utility for promoting vascular health emerged from the United Kingdom Prospective Diabetes Study, a prospective controlled study which examined long-term health outcomes in obese diabetics randomly allocated to therapy with metformin, a sulfonylurea, insulin, or dietary control. Even though patients treated with metformin achieved glycemic control no better than that achieved by patients treated with sulfonylureas or insulin, their mortality over ten years of follow-up from MI, stroke, or all-causes was significantly lower.⁵³ Subsequent epidemiological studies have concluded that, as compared to patients achieving comparable metabolic control with other agents, metformin-treated patients are less likely to die from MI, stroke, or congestive failure.⁵⁴⁻⁵⁹

Studies with rodents and cultured cells demonstrate that metformin and other AMPK activators can act directly on vascular endothelium, foam cells, and cardiomyocytes in ways that could be expected to diminish risk for atherosclerosis, thrombotic events, and ventricular hypertrophy. Much of this protection stems from the ability of AMPK to activate eNOS via direct phosphorylations of Ser633 and Ser1177; the activating phosphorylation of Ser633 is not duplicated by Akt activity.^{13, 14} NO, in the modest physiological concentrations produced by eNOS, is well known to exert ant-atherosclerotic, anti-thrombotic, vasodilatory, and anti-hypertrophic effects crucial for preserving healthful structure and function of the vascular system.^{60, 61} Recent studies show that the activation of eNOS induced by HDL and by adiponectin in endothelial cells – thought to play a key role in the vascular protection afforded by these agents – is mediated via activation of AMPK.^{5, 62, 63} Likewise, the ability of exercise-induced shear stress to stimulate eNOS and boost its expression is mediated in part via AMPK; the impact on eNOS expression is requires AMPK-mediated induction of the transcription factor kruppel-like factor 2.^{64, 65}

Activation of NF-kappaB and increased production of oxidative stress via NAPDH oxidase complexes contribute importantly to the inflammatory endotheliopathy that is conducive to atherogenesis and thrombosis. In many studies, AMPK activation has been shown to down-regulate NF-kappaB activation and oxidative stress in various types of cells; the precise mechanisms responsible require further clarification.⁶⁶ In part, this effect reflects the fact that Sirt1 activity (as stimulated by AMPK) impairs the transcriptional activity of NF-kappaB by removing an activating acetyl group from p65.⁶⁷ A recent study with mouse and human endothelial cell cultures has demonstrated that AMPK α 2 can modestly reduce proteasome activity; this likewise results in a down-regulation of NF-kappaB activation.⁶⁸ (A similar effect of AMPK on proteasomes has been reported in fibroblasts.⁶⁹) This down-regulation of NF-kappa, in turn, results in decreased expression of various components of the NAPDH oxidase complex.⁶⁸ The possibility that AMPK may also exert a more rapid inhibitory impact on NADPH oxidase, possibly by suppressing PKC-mediated membrane translocation of p47phox, requires more evaluation.⁷⁰⁻⁷³ Another recent study found that AMPK α 2 impedes NF-kappaB activation in endothelial cells via direct phosphorylations of IKK- β ; these phosphorylations impede IKK- β 's ability to phosphorylate I κ B and thereby promote nuclear translocation of NF-kappaB.⁷⁴ Whether these findings can be generalized to other types of cells remains to be determined; in any case, it is clear that AMPK can down-regulate NF-

kappaB activity via several independent mechanisms, the importance of which may vary dependent on cell type. AMPK-mediated proteasomal inhibition can also help to preserve the proper coupling of eNOS (thereby warding off an increase in oxidative stress and a loss of NO production) by slowing proteasomal degradation of GTP cyclohydrolase in oxidatively-stressed endothelial cells; this latter enzyme is rate-limiting for the production of eNOS's essential cofactor, tetrahydrobiopterin.⁷⁵

With respect to the role of foam cells in atherosclerosis, several studies – with one contrary exception⁷⁶ – report that agents which activate AMPK tend to boost cholesterol efflux and prevent lipid accumulation in foam cells exposed to oxidized LDL.⁷⁷⁻⁷⁹ A related report indicates that AICAR-mediated AMPK activation suppresses the macrophage proliferation induced by exposure to oxidized LDL.⁸⁰

Although hepatic activation of AMPK, mediated via metformin, appears to have a modest impact at best on serum levels of LDL – a curious fact, given that AMPK has the potential, like statins, to inhibit HMG-CoA reductase⁸¹ - the AMPK activator berberine is notably effective for lowering serum LDL cholesterol.⁸²⁻⁸⁵ However, this effect appears to be independent of any inhibition of HMG-CoA reductase; rather, it reflects an increase in the half-life of LDL receptor mRNA. Fortunately, this effect is complementary to the stimulatory impact of statins on transcription of this mRNA, such that combined use of statins and berberine can achieve a very marked lowering of LDL levels.⁸⁶ It is not clear whether AMPK has anything to do with this fortuitous benefit of berberine.

Studies examining the impact of AMPK activation on models of ventricular hypertrophy and congestive failure were prompted by epidemiological studies demonstrating that diabetics experiencing heart failure were at lower risk for mortality if they were treated with metformin.^{57, 58} In rodent studies of cardiac pressure overload, concurrent treatment with AICAR, metformin, or berberine has reduced ventricular hypertrophy; conversely, mice which are genetically deficient in AMPK- α 2 or in LKB1 expression are more prone to ventricular hypertrophy in the context of overload.⁸⁷⁻⁹⁴ The protection afforded by AMPK in this regard reflects not only an effect on cardiomyocyte hypertrophy, but also an anti-fibrotic effect.^{88, 91} Although the down-regulatory impact of AMPK on protein synthesis might play some role in this effect, there is evidence that increased eNOS activity also plays a prominent role;⁹⁰ indeed, it is well established that effective NO activity tends to prevent cardiac hypertrophy.⁶¹

In a randomized clinical trial enrolling 156 patients with congestive heart failure, patients received berberine or placebo in addition to standard management.⁹⁵ After 8 weeks, improvements in ventricular ejection fraction, exercise capacity, and frequency of premature ventricular complexes were notably better in the berberine group. At a two year follow-up, mortality in the berberine group was about half as high as in the placebo group (7 vs. 13, $p < .02$). This appears to be the first formal clinical trial which evaluated an AMPK activator in chronic congestive heart failure – in patients most of whom were not diabetic.

Much of the favorable influence of AMPK activation on vascular health is likely mediated by its impact on metabolic syndrome, which we now examine.

Controlling Metabolic Syndrome and Diabetes

The root cause of metabolic syndrome is inflammation in visceral adipocytes.⁹⁶ As these adipocytes hypertrophy, they tend to become oxidatively stressed and develop an inflammatory phenotype associated with increased production and secretion of cytokines such as IL-6 and TNF-alpha, and decreased

secretion of the protective adipokine adiponectin (which, as noted, can activate AMPK). The efficiency of insulin signaling in these adipocytes is impaired, and the resulting up-regulation in adipocyte lipolysis floods the body with excessive levels of free fatty acids, most notably during post-absorptive metabolism when free fatty acids are not needed as fuel. This free fatty acid “pollution” leads to the accumulation of triglycerides and other types of “ectopic fat” in tissues such as skeletal muscle, vascular endothelium, hepatocytes, and beta cells.^{97, 98} Certain of these ectopic fat metabolites, such as ceramide and diacylglycerol, activate signaling pathways that promote oxidative stress, insulin resistance, and inflammation in the tissues that harbor them. Hence, metabolic syndrome is often associated with insulin resistance of skeletal muscle fibers, inflammatory dysfunction of vascular endothelium, non-alcoholic steatohepatitis, and, in beta cells, glucolipotoxicity that ultimately may precipitate beta cell “failure” and type 2 diabetes. These effects can be exacerbated by increased exposure to adipose-derived pro-inflammatory cytokines and by diminution of adiponectin activity. The characteristic perturbations of serum lipids seen in metabolic syndrome are largely a consequence of increased free fatty acid flux into hepatocytes, which results in hepatic secretion of an increased number of VLDL particles and apoB molecules.⁹⁹

The extent to which AMPK can suppress inflammation and oxidative stress in hypertrophied visceral adipocytes, and restore effectiveness of adipocyte insulin signaling, remains unclear. Several reports the serum IL-6 levels decline during metformin or berberine therapy in diabetics, are suggestive of an anti-inflammatory impact;^{85, 100-102} however, metformin usually does not influence adiponectin levels.¹⁰³⁻¹⁰⁵ A small amount of evidence suggests that metformin may improve the insulin sensitivity of adipocytes in diabetics.^{106, 107} Whether or not AMPK helps “get to the root” of visceral adipocyte dysfunction, it clearly can mimic the anti-lipolytic impact of insulin on adipocytes. Catecholamines and other agonists that boost cAMP in adipocytes stimulate lipolysis owing to a PKA-mediated phosphorylation and activation of the hormone-sensitive lipase. Insulin antagonizes this effect by activating a cAMP phosphodiesterase (3B) which opposes the catecholamine-induced rise in cAMP, and hence prevents activation of PKA and its downstream target hormone-sensitive lipase.¹⁰⁸ AMPK likewise antagonizes catecholamine-mediated activation of hormone-sensitive lipase, but in a different way: by phosphorylating Ser565 on this lipase, it renders the lipase a poor substrate for the activating phosphorylations catalyzed by PKA.^{109, 110} This likely explains why diurnal levels of serum free fatty acids tend to be suppressed by therapy with metformin or berberine.^{106, 107, 111} Evidently, since excess free fatty acid exposure is a key mediator of the complications of metabolic syndrome, this down-regulatory impact of AMPK activation on serum free fatty acids can be notably protective.

Fortunately, the decrease in adipocyte lipolysis associated with metformin or berberine therapy does not tend to promote weight gain, likely because one of the key effects of AMPK is to boost the efficiency of mitochondrial fatty acid oxidation. AMPK accomplishes this, in the short term, by decreasing malonyl-CoA levels via an inhibitory phosphorylation of the enzyme acetyl-CoA carboxylase and an activating phosphorylation of malonyl-CoA decarboxylase.^{112, 113} Malonyl-CoA is not only an obligate substrate for fatty acid and cholesterol synthesis, but also functions as an allosteric inhibitor of carnitine palmitoyltransferase-I, rate-limiting for the transfer of fatty acids into the mitochondrial inner matrix and fatty acid oxidation.¹¹⁴ Hence, AMPK activity tends to disinhibit fatty acid oxidation via its impact on malonyl-CoA. In the longer term, AMPK may increase the maximal capacity of tissues for fatty acid oxidation by promoting mitochondrial biogenesis. AMPK also acts to channel free fatty acids towards oxidation by suppressing activity of glycerol-3-phosphate acyltransferase, a key mediator of triglyceride

synthesis.^{112, 115} Hence, when metabolic syndrome exposes tissues to excessive levels of free fatty acids, AMPK tends to route these fatty acids to oxidation, rather than to conversion to triglycerides or the ectopic fat metabolites that are pathogenic.

In short, AMPK activation has the potential to minimize the adverse impact of ectopic fat by slowing adipocyte release of free fatty acids, and also by reducing the propensity of the fatty acids that are released to be converted to pathogenic metabolites. This mechanism may help to explain why metformin can help to prevent or postpone the onset of type 2 diabetes in at-risk subjects, as demonstrated in the Diabetes Prevention Program¹¹⁶ – it may lessen the exposure of beta cells to the ectopic fat metabolites that promote beta cell failure.¹¹⁷⁻¹¹⁹

The most important factor in the improved diabetic glycemic control imparted by metformin appears to be a down-regulation of hepatic gluconeogenesis that lessens hepatic glucose output and hence helps to moderate fasting glucose levels.¹²⁰ This reflects the fact that AMPK suppresses transcription of the genes coding for the key gluconeogenic enzymes phosphoenolpyruvate carboxykinase and fructose-diphosphatase. AMPK achieves this by phosphorylating, and thereby inhibiting the activity of, two coactivators (CREB binding protein, and CRTC2) required for efficient transcription of these genes.^{121, 122} Metformin's inhibition of CREB binding protein precisely mimics the impact of insulin in this regard.¹²¹ AMPK also boosts the synthesis of the orphan nuclear receptor “small heterodimer partner”, which likewise interferes with the transcription of these key genes.^{123, 124} Berberine likewise suppresses gluconeogenesis in diabetic rats;¹²⁵ its clinical utility for glycemic control appears to be quite comparable to that of metformin.^{85, 111}

Weight Control

AMPK activators may also help to prevent metabolic syndrome by reducing risk for weight gain. Diabetologists are well aware that metformin therapy tends to promote a modest degree of weight loss – whereas therapy with sulfonylureas or insulin tends to promote weight gain, often exacerbating the underlying problem.¹²⁶ Of the newer diabetes therapies, GLP-1 agonists also tend to lower body weight; it is not likely to be accidental that they also activate hepatic AMPK.^{126, 127} Metformin also promotes weight loss in the context of polycystic ovary syndrome, and has also been employed with some success to prevent weight gain in patients treated with certain anti-psychotic agents.¹²⁸⁻¹³⁰ Hence, there is no reason to suspect that weight control benefits of AMPK activation will be confined to diabetics.¹³¹

Increased hepatic fatty acid oxidation may be a key mediator of metformin's impact on weight control. Efficient hepatic fatty acid oxidation sends a satiety signal to the brain via the vagus nerve; this mechanism likely contributes to the satiety associated with prolonged fasting.¹³²⁻¹³⁴ Moreover, it is intriguing to note that expression of a constitutively active form of AMPK α 2 in mouse liver induces uncoupling protein-2 (UCP-2) – possibly owing to increased function of Foxo3a and PGC-1alpha, which drive its transcription in endothelial cells.¹³⁵⁻¹³⁷ Arguably, this effect could be exploited to transform the liver into a thermogenic organ – in which uncoupled oxidation of fatty acids yields CO₂ and heat – when used in conjunction with other strategies that boost hepatic fatty acid oxidation.^{138, 139}

AMPK also suppresses hepatic de novo lipogenesis, via inhibition of the transcription and post-translational processing of the key transcription factor for lipogenic enzymes, SREBP-1c.^{49, 140, 141} While this effect helps to explain the utility of AMPK activators for preventing obesity in rodents, it is less

likely to be of importance in humans, in whom hepatic de novo lipogenesis tends to be of minor significance.¹⁴²

A recent Japanese study has found that regular vinegar ingestion can promote modest weight loss in obese subjects.¹⁴³ This is pertinent in light of the fact – as discussed below – that vinegar (acetic acid) has the potential to activate hepatic AMPK.¹⁴⁴ Vinegar also induced UCP-2 in mouse liver – an effect which was blocked by inhibition of AMPK.¹⁴⁴

Preventing and Controlling Cancer

A number of recent epidemiological studies have found that diabetics using metformin, as opposed to alternative therapies, are at lower risk for cancer, including specifically breast, prostate and colorectal cancer.¹⁴⁵⁻¹⁵³ It is likely that AMPK's inhibitory impact on mTORC1 activity plays a major role in this effect. Other circumstances which likewise decrease mTORC1 activity – elevated adiponectin levels, rapamycin therapy, and plant-based diets or superior insulin sensitivity (associated with low serum levels of insulin and free IGF-I) – have also been linked to lower cancer risk in epidemiological analyses.¹⁵⁴ This likely is because mTORC1 works in various ways to promote cellular proliferation and inhibit apoptosis - effects which could be expected to promote the accumulation of mutations in, and aid the survival of, pre-cancerous stem cells.

mTORC1 phosphorylates 4EBP-1, thereby freeing the translation initiating factor eIF4E from inhibitory binding.¹⁵⁵ eIF4E functions to expedite the translation of a number of “weak” mRNAs which otherwise would be translated to a minimal extent. Some of these mRNAs are handicapped by complex hairpin structures in their 5' UTRs; others are characterized by a specific nucleotide structure in their 3' UTRs, and require binding to eIF4E to achieve extranuclear transport. Among these weak mRNAs are some which code for proteins that promote proliferation – cyclin D1, c-myc, ODC – and others that block apoptosis – survivin, Bcl-2, Bcl-xL, Mcl-2, dad1.^{154, 156-159} Hence, mTORC1, acting via eIF4E, up-regulates the expression of a number of proteins which are conducive to cancerous transformation. Intriguingly, overexpression of eIF4E in fibroblasts and other cell lines has been shown to promote transformation in vitro.¹⁶⁰ The weak mRNAs whose translation is expedited by mRNAs also code for a number of proteins that render transformed cells more aggressive in their behavior, and more chemoresistant; not surprisingly, constitutive activation of mTORC1, and/or overexpression of eIF4E, is observed in a high proportion of advanced malignancies.¹⁵⁸

Additionally, there is recent evidence that mTORC1 very rapidly up-regulates the activity of Gq-coupled receptors that can promote proliferation in some cell types by somehow activating the Raf-MEK-Erk1/2 pathway.¹⁶¹ Hence, insulin, IGF-I and various other growth factors have the potential to boost the mitogenic response to various hormones and cytokines that activate such receptors. Metformin, via AMPK, has been shown to antagonize this effect.^{162, 163} Since this effect of mTORC1 is rapid in onset, modulation of protein translation is clearly not involved; the direct target of mTORC1's activity in this regard has not yet been defined.

The contribution of the mTORC1 signaling to cancer aggressiveness suggests that AMPK activation may be a rational strategy for cancer management, and indeed there is burgeoning interest in metformin as an adjunctive agent in cancer therapy.¹⁶⁴⁻¹⁶⁶ Much of this interest was prompted by several epidemiological studies which have concluded that diabetic cancer patients receiving metformin therapy tend to have

better prognoses than those receiving alternative diabetes therapies,¹⁶⁷⁻¹⁶⁹ this effect may contribute to a reduction in total cancer mortality in diabetics using metformin.¹⁷⁰ While metformin has obvious potential for slowing cancer growth, there is also new evidence that metformin may specifically target cancer stem cells that are thought to be largely responsible for cancer chemoresistance and recurrence; metformin may change the differentiation state of these cells, and/or make them more susceptible to cytotoxic drugs.¹⁷¹⁻¹⁷⁴ Hence, it is hoped that metformin, employed as an adjuvant to chemotherapy regimens, may increase chances for a cure, or at least notably enhance the contribution of such regimens to survival time.

While Western oncologists are focusing on metformin as a cancer therapy adjuvant, quite a number of Chinese studies have demonstrated cancer-retardant or cancer-preventive activity for berberine, *in vitro* and in mice.¹⁷⁵⁻¹⁷⁷ AMPK activation, rather than metformin *per se*, is emerging as a key strategy for prevention and control of cancer.

The modest reduction in serum insulin and free IGF-I achievable with AMPK activation can also be expected to down-regulate mTORC1 activity, complementing the more direct impact of AMPK in this regard.³⁶ Moreover, low insulin and free IGF-I should decrease Akt activity in many tissues; Akt works in a number of ways to suppress apoptosis and thereby raise cancer risk.¹⁷⁸

AMPK may have a more specific role to play in the prevention of post-menopausal breast cancer. It is now appreciated that the predominant source of the estrogen that drives cancer induction post-menopausally is the aromatase activity of breast stroma.¹⁷⁹ AMPK activation has recently been shown to inhibit the transcription of aromatase in breast stroma. In these stromal cells, aromatase transcription is driven by a CREB/CRTC2 complex; AMPK phosphorylates the coactivator CRTC2 in a way that promotes its eviction from the nucleus, thereby preventing transcription of the aromatase gene.^{180, 181} In this formulation, the elevated serum levels of estrogen in obese post-menopausal women are seen as a red herring; rather, the nexus between obesity, estrogen and elevated breast cancer risk is driven by leptin, which suppresses AMPK activity in breast stroma by down-regulating LKB1.^{180, 182}

Preventing Neurodegenerative Disorders

The ability of AMPK to promote macroautophagy suggests that AMPK activation may have potential for preventing or controlling neurodegenerative disorders characterized by intraneuronal or extracellular accumulation of toxic protein aggregates. It stands to reason that autophagy, the “house cleaning” strategy of our cells, should be of particular importance to maintaining the healthful structure and function of long-lived, difficult-to-replace cells such as neurons.¹⁸³⁻¹⁸⁶ Indeed, transgenic mice with severely impaired capacity for neuronal autophagy develop neurodegeneration at an early age, and dysfunctional neuronal autophagy is often observed in neurodegenerative disorders.^{183, 187} On the other hand, overwhelmingly intense activation of autophagy can contribute to neuronal death when not properly balanced by biosynthesis.¹⁸⁸ These considerations have led some researchers to suggest that AMPK-mediated up-regulation of autophagy may have greater net utility in the early stages of neurodegenerative disorders, than in their advanced stages.¹⁸⁹ In other words, AMPK activation is best viewed as a preventive rather than therapeutic strategy for addressing neurodegeneration.

With respect to Alzheimers risk, AMPK activators have been shown to decrease extracellular amyloid-beta accumulation, owing to increased autophagic degradation of this protein.^{190, 191} How AMPK

influences amyloid-beta production per se has been the subject of conflicting reports,¹⁹²⁻¹⁹⁴ but in any case up-regulation of autophagy can suppress deposition of amyloid-beta aggregates. Indeed, in transgenic APP/PS1 mice prone to amyloid deposition, resveratrol administration reduced amyloid deposition in the brain globally; this effect was significant in the cortex though not the hippocampus.¹⁹⁰

With respect to tau phosphorylation, a key factor in the formation of the neurofibrillary tangles characteristic of Alzheimers, AMPK appears to have equivocal effects. Sirt1 activity opposes such phosphorylation, so AMPK has the potential to block tau phosphorylation via Sirt1 activation.^{189, 195, 196} On the other hand, AMPK also has the potential to phosphorylate tau directly.¹⁹⁷ It is not yet clear which of these mechanisms is predominant during the elevation of Alzheimers.

The ability of AMPK to boost eNOS activity in the cerebral microvasculature may also aid Alzheimers prevention, in light of recent evidence that NO of vascular origin acts to impede the synthesis of amyloid-beta by suppressing expression of the BACE1 protease required for its production.¹⁹⁸ This intriguing finding may help to rationalize the numerous studies demonstrating that people with elevated cardiovascular risk factors – risk factors that would be expected to compromise vascular NO bioactivity – are at increased risk for Alzheimers as they age.¹⁹⁹ Hence, the favorable impact of AMPK activation on vascular health may tend to reduce risk for dementia, both by limiting stroke risk, and by helping to control cerebral amyloid-beta production.

Activated microglia, via production of peroxynitrite and various pro-inflammatory cytokines and prostanoids, are suspected to contribute to the pathogenesis of many neurodegenerative disorders.²⁰⁰ In light of the anti-inflammatory potential of AMPK as noted above, it is reasonable to suspect that AMPK activation might be protective in this regard, and indeed several studies demonstrate that AMPK - activators can exert anti-inflammatory effects on cultured microglia. The ability of berberine-activated AMPK to suppress induction of iNOS and Cox-2 in LPS or interferon- γ -exposed microglia might at least partially reflect the fact that AMPK phosphorylates and triggers the intranuclear transport of HuR, a protein which up-regulates the translation of iNOS and Cox-2 mRNAs by binding to AU-rich regions in their 3' UTRs, thereby enhancing the half-life of these mRNAs; loss of cytoplasmic HuR therefore decreases the protein expression of iNOS and Cox-2.²⁰¹⁻²⁰⁶

(Parenthetically, it should be noted that the ability of AMPK to down-regulate iNOS translation suggests a role for AMPK activators in the prevention of septic shock. Indeed, berberine administration has been reported to enhance survival in a murine model of endotoxemia.²⁰⁷ A further corollary is that the suppressive impact of AMPK activators on Cox-2 induction may contribute to the cancer-preventive potential of these agents; the marked cancer prevention associated with daily low-dose aspirin use is likely attributable to partial inhibition of Cox-2.²⁰⁸)

Although risk for dementia and cognitive dysfunction are increased in diabetics, so far little epidemiology has focused on the impact of specific diabetic therapies on dementia risk. However, one recent study has found that, as compared to diabetic not receiving drug therapy, those receiving metformin were about 25% less likely to develop dementia.²⁰⁹ Dementia risk also trended slightly lower in patients receiving sulfonylureas, so it is not clear whether the apparent protection associated with metformin reflected a specific impact on the brain, or simply superior glycemic control. The impact of metformin therapy on risk for Parkinson's disease has not yet been examined.

Preserving Cartilage and Bone

Diabetics treated with metformin have been found to be at decidedly lower risk for fractures than those treated with thiazolidinediones.^{210, 211} In part, this reflects an adverse effect of the latter – but there is growing reason to suspect that metformin and AMPK exert a favorable effect on bone density and structure. Metformin exposure *in vitro* promotes osteoblastic differentiation and behavior – increasing alkaline phosphatase activity and type 1 collagen production.²¹²⁻²¹⁴ These effects appear to be contingent on the AMPK-mediated induction of the short heterodimer partner orphan nuclear receptor cited above. Concurrently, metformin antagonizes osteoclast development by reducing osteoblast production of RANKL, while increasing production of osteoprotegerin (a decoy receptor for RANKL).²¹⁵ Suppression of RANKL expression and osteoclasts development has also been seen with berberine and other AMPK activators.^{216, 217} Moreover, both metformin and berberine have demonstrated favorable effects on bone density and structure in ovariectomized rats, and in other rodent models of bone loss.^{215, 218, 219} In light of the fact that the bone protective effects of estrogen are thought to be mediated largely by induction of eNOS in osteoblasts, it is pertinent to recall that AMPK can activate this enzyme directly.^{220, 221}

AMPK may also aid the preservation of cartilage in the context of osteoarthritis. Cartilage loss in osteoarthritis is believed to reflect a catabolic impact of cytokines, most notably IL-1, on chondrocytes, associated with increased production of collagenolytic metalloproteinases and down-regulation of the tissue inhibitor of metalloproteinases.²²² Oxidative stress, NF-kappaB activation, and iNOS induction are key mediators of these effects.²²³⁻²²⁷ It is reasonable to suspect that AMPK might act to oppose these effects, and indeed, a recent study has found that AMPK activators notably suppress the catabolic response of chondrocytes to IL-1 or TNF- α exposure; notably, chondrocyte production of MMP-3, MMP-13, and NO was suppressed.²²⁸ Conversely, knockout of AMPK α with small interfering RNA exacerbated the catabolic response of chondrocytes to IL-1 and TNF- α . Chinese researchers have reported a very analogous anti-catabolic response when IL-1 treated chondrocytes are exposed to berberine; in addition, they report that intra-articular administration of berberine provides protection from cartilage damage in rats concurrently given intra-articular injections of IL-1.²²⁹ So far, no epidemiological studies have addressed the impact of metformin therapy on cartilage status in diabetics.

If indeed AMPK activators can protect the healthful structure and function of the vasculature, bones, and cartilage, they may be viewed more generally as beneficial for the health of connective tissues. Such a view would be highly consistent with the thesis that AMPK activation has “anti-aging” potential.

Controlling Autoimmunity

AMPK activation may have potential for controlling autoimmune disorders driven by autoreactive Th1 or Th17 lymphocytes. In the murine model of multiple sclerosis, experimental autoimmune encephalomyelitis (EAE), metformin, berberine, and AICAR have been shown to be highly protective.²³⁰⁻²³³ Some of this protection appears to stem from inhibition of NF-kappaB activation and costimulatory protein expression in antigen-presenting cells, which lessens the ability of these cells to activate autoreactive Th1 and Th17 helper lymphocytes.²³¹ Indeed, when EAE was induced in mice concurrently treated with berberine, adoptive transfer of Th17 cells from these mice to untreated mice completely failed to induce EAE in the recipients; transfer of Th1 cells induced only an attenuated syndrome.²³¹ Importantly, berberine treatment did not influence the relative number of CD+4FoxP3+ regulatory T

cells. The likelihood that AMPK plays a physiological role in controlling autoimmunity is suggested by the fact that EAE is an especially aggressive syndrome in AMPK α 1-null mice.²³⁴

It seems unlikely that EAE will be the only autoimmune disorder in which AMPK activation is protective. Indeed, berberine and AICAR have been shown to ameliorate murine models of colitis.²³⁵⁻²³⁸ Much further research addressing the immunomodulatory potential of AMPK activation is warranted.

Practical Strategies for Implementing AMPK Activation

The hypothesis that AMPK activators may, at least in some measure, confer health benefits comparable to those seen with sustained calorie restriction, prompts an inquiry into the range of health effects that might flow from chronic AMPK activation. The data cited above suggest that such activation may indeed reduce risk for atherosclerosis, heart attack, and stroke; reduce risk for ventricular hypertrophy while aiding control of congestive failure; ameliorate the severity of metabolic syndrome, reduce diabetes risk, and improve diabetic control; help to prevent weight gain; decrease risk for a number of types of cancer, and serve as a worthwhile adjunct to cancer chemotherapy; reduce risk for dementia and possibly other neurodegenerative disorders; help to preserve the proper structure and function of bone and cartilage; and possibly help to prevent or control certain autoimmune disorders. The scope of these potential benefits is so vast that it does indeed lend credence to the notion that AMPK activation has “anti-aging” activity that may promote a notable augmentation of healthspan.

Metformin is the world’s most widely used diabetic drug, typically administered in a dose of 500-850 mg twice daily. A minority of patients who use it experience some GI upset, which tends to become less significant over time. Otherwise, it appears to be safe and well tolerated; the risk for lactic acidosis that prompted the banning of the related biguanide drug phenformin, appears to be almost non-existent with prescribed doses of metformin.²³⁹ For non-diabetics who seek to employ AMPK activation as a longterm health promotion strategy, metformin has the disadvantage that it is available only by prescription.

Berberine, on the other hand, is a natural compound, found in a number of medicinal herbs, that is currently available in its pure form as a nutraceutical in the U.S. It is widely used in China for diabetes management. Typical dose regimens are 500 mg 2-3 times daily, or 300 mg 3 times daily; these are reported to achieve an improvement in glycemic control roughly comparable to that seen with metformin – accompanied by a greater reduction in LDL cholesterol.^{85, 111} A small minority of patients experience significant constipation, but otherwise it is well tolerated. Berberine is not efficiently absorbed, and innovations in delivery – microemulsification or cyclodextrin inclusion complexation – might enable lower doses to provide worthwhile benefit.^{240, 241} Berberine may have tremendous potential as a “life extension” nutraceutical.

Resveratrol, a polyphenol found in red wine and many other foods, has recently been widely promoted in the U.S. as an aging-retardant nutraceutical, owing to reports that it activates Sirt1 in cell cultures and improves survival in obese mice. Unfortunately, it is conjugated rapidly and completely following absorption; despite reasonably efficient absorption, orally administered resveratrol do not appear to achieve sustained serum levels of free resveratrol sufficient to inhibit mitochondrial ATP synthase and thereby activate AMPK.^{21, 22} However, oral resveratrol might have the potential to transiently activate AMPK in the intestinal mucosa. Resveratrol, as well as the AMPK activators metformin, berberine, and AICAR, and acetic acid (see below), have been shown to boost intestinal production of the incretin

hormone glucagon-like peptide-1 (GLP-1), which in turn stimulates hepatic AMPK activity,²⁴²⁻²⁴⁶ moreover, GLP-1 may exert protective effects on other tissues as well.²⁴⁷⁻²⁴⁹ Perhaps this rationalizes the modestly favorable results of resveratrol supplementation (150 mg daily) reported in a recent clinical study.²⁵⁰

Remarkably, vinegar- dilute acetic acid – can also activate AMPK in some tissues. This likely reflects the fact that the initial step of acetate metabolism, in which acetate is phosphorylated, generates AMP in the process. Vinegar-mediated activation of AMPK has indeed been demonstrated in the liver and of vinegar-fed rats, and in human endothelial cells in vitro.^{144, 251, 252} This effect will presumably be more transitory than that of AMPK-activating drugs, as acetate is rapidly oxidized following ingestion. Nonetheless, oral vinegar administration has exerted some intriguing effects, both in rodents and in clinical studies, that arguably are attributable to AMPK activation. In light of the favorable impact of metformin on weight control, it is interesting to note that vinegar administration suppresses weight gain in rats fed a high-fat diet, and aids glycemic control in diabetic mice,^{144, 251} moreover, in a placebo-controlled clinical trial, obese subjects ingesting 15-30 ml vinegar daily achieved modest but statistically significant weight loss compared to those receiving placebo vinegar.¹⁴³ Moreover, in post-menopausal women, vinegar administration boosts flow-mediated vasodilation, an effect likely attributable to AMPK-mediated phosphorylation of eNOS.²⁵² Regular vinegar use may modestly improve glycemic control in human diabetics, and inclusion of vinegar in meals acutely lowers the postprandial glucose response, apparently by slowing the absorption of starch or polysaccharides.²⁵³⁻²⁵⁵ These observations are intriguing in light of the common folkloric belief that apple cider vinegar can confer wide-ranging health protective benefits. And it is reasonable to speculate that some of the protective health benefits associated with moderate regular ingestion of alcohol may in fact be mediated by the acetic acid evolved by ethanol metabolism – and hence by AMPK!²⁵⁶ (Unfortunately, these are often counterbalanced by adverse effects of acetaldehyde and ethanol per se.) Moreover, other short-chain fatty acids generate AMP when metabolized, and hence can activate AMPK.²⁵⁷ It recently has been credibly proposed that AMPK activation mediated by the short-chain fatty acids stemming from colonic metabolism of dietary fiber may be largely responsible for the favorable impact of high-fiber diets on control of metabolic syndrome;²⁵⁸ conceivably, this phenomenon might also impact risk for colorectal cancer.²⁵⁹ Regular use of vinegar, fiber-rich diets, and moderate alcohol consumption, may represent wholly nutritional strategies for evoking, to a modest degree, some of the health protection afforded by AMPK activation.

References

- (1) Xiao B, Sanders MJ, Underwood E, Heath R, Mayer FV, Carmena D, Jing C, Walker PA, Eccleston JF, Haire LF, Saiu P, Howell SA, Aasland R, Martin SR, Carling D, Gamblin SJ. Structure of mammalian AMPK and its regulation by ADP. *Nature* 2011 April 14;472(7342):230-3.
- (2) Carling D, Mayer FV, Sanders MJ, Gamblin SJ. AMP-activated protein kinase: nature's energy sensor. *Nat Chem Biol* 2011;7(8):512-8.

- (3) Hawley SA, Ross FA, Chevtzoff C, Green KA, Evans A, Fogarty S, Towler MC, Brown LJ, Ogunbayo OA, Evans AM, Hardie DG. Use of cells expressing gamma subunit variants to identify diverse mechanisms of AMPK activation. *Cell Metab* 2010 June 9;11(6):554-65.
- (4) Wu Y, Song P, Xu J, Zhang M, Zou MH. Activation of protein phosphatase 2A by palmitate inhibits AMP-activated protein kinase. *J Biol Chem* 2007 March 30;282(13):9777-88.
- (5) Kimura T, Tomura H, Sato K, Ito M, Matsuoka I, Im DS, Kuwabara A, Mogi C, Itoh H, Kurose H, Murakami M, Okajima F. Mechanism and role of high density lipoprotein-induced activation of AMP-activated protein kinase in endothelial cells. *J Biol Chem* 2010 February 12;285(7):4387-97.
- (6) Cool B, Zinker B, Chiou W, Kifle L, Cao N, Perham M, Dickinson R, Adler A, Gagne G, Iyengar R, Zhao G, Marsh K, Kym P, Jung P, Camp HS, Frevert E. Identification and characterization of a small molecule AMPK activator that treats key components of type 2 diabetes and the metabolic syndrome. *Cell Metab* 2006 June;3(6):403-16.
- (7) Holland WL, Miller RA, Wang ZV, Sun K, Barth BM, Bui HH, Davis KE, Bikman BT, Halberg N, Rutkowski JM, Wade MR, Tenorio VM, Kuo MS, Brozinick JT, Zhang BB, Birnbaum MJ, Summers SA, Scherer PE. Receptor-mediated activation of ceramidase activity initiates the pleiotropic actions of adiponectin. *Nat Med* 2011 January;17(1):55-63.
- (8) Smith AR, Visioli F, Frei B, Hagen TM. Lipoic acid significantly restores, in rats, the age-related decline in vasomotion. *Br J Pharmacol* 2008 April;153(8):1615-22.
- (9) Lee WJ, Song KH, Koh EH, Won JC, Kim HS, Park HS, Kim MS, Kim SW, Lee KU, Park JY. Alpha-lipoic acid increases insulin sensitivity by activating AMPK in skeletal muscle. *Biochem Biophys Res Commun* 2005 July 8;332(3):885-91.
- (10) Hardie DG. AMP-activated/SNF1 protein kinases: conserved guardians of cellular energy. *Nat Rev Mol Cell Biol* 2007 October;8(10):774-85.
- (11) Kukidome D, Nishikawa T, Sonoda K, Imoto K, Fujisawa K, Yano M, Motoshima H, Taguchi T, Matsumura T, Araki E. Activation of AMP-activated protein kinase reduces hyperglycemia-induced mitochondrial reactive oxygen species production and promotes mitochondrial biogenesis in human umbilical vein endothelial cells. *Diabetes* 2006 January;55(1):120-7.
- (12) Colombo SL, Moncada S. AMPKalpha1 regulates the antioxidant status of vascular endothelial cells. *Biochem J* 2009 July 15;421(2):163-9.
- (13) Chen ZP, Mitchelhill KI, Michell BJ, Stapleton D, Rodriguez-Crespo I, Witters LA, Power DA, Ortiz de Montellano PR, Kemp BE. AMP-activated protein kinase phosphorylation of endothelial NO synthase. *FEBS Lett* 1999 January 29;443(3):285-9.
- (14) Chen Z, Peng IC, Sun W, Su MI, Hsu PH, Fu Y, Zhu Y, DeFea K, Pan S, Tsai MD, Shyy JY. AMP-activated protein kinase functionally phosphorylates endothelial nitric oxide synthase Ser633. *Circ Res* 2009 February 27;104(4):496-505.
- (15) Canto C, Auwerx J. Calorie Restriction: Is AMPK a Key Sensor and Effector? *Physiology (Bethesda)* 2011 August;26(4):214-24.

- (16) Dolinsky VW, Morton JS, Oka T, Robillard-Frayne I, Bagdan M, Lopaschuk GD, Des RC, Walsh K, Davidge ST, Dyck JR. Calorie restriction prevents hypertension and cardiac hypertrophy in the spontaneously hypertensive rat. *Hypertension* 2010 September;56(3):412-21.
- (17) Fulco M, Cen Y, Zhao P, Hoffman EP, McBurney MW, Sauve AA, Sartorelli V. Glucose restriction inhibits skeletal myoblast differentiation by activating SIRT1 through AMPK-mediated regulation of Nampt. *Dev Cell* 2008 May;14(5):661-73.
- (18) Canto C, Gerhart-Hines Z, Feige JN, Lagouge M, Noriega L, Milne JC, Elliott PJ, Puigserver P, Auwerx J. AMPK regulates energy expenditure by modulating NAD⁺ metabolism and SIRT1 activity. *Nature* 2009 April 23;458(7241):1056-60.
- (19) Um JH, Park SJ, Kang H, Yang S, Foretz M, McBurney MW, Kim MK, Viollet B, Chung JH. AMP-activated protein kinase-deficient mice are resistant to the metabolic effects of resveratrol. *Diabetes* 2010 March;59(3):554-63.
- (20) Canto C, Jiang LQ, Deshmukh AS, Matakki C, Coste A, Lagouge M, Zierath JR, Auwerx J. Interdependence of AMPK and SIRT1 for metabolic adaptation to fasting and exercise in skeletal muscle. *Cell Metab* 2010 March 3;11(3):213-9.
- (21) Walle T, Hsieh F, DeLegge MH, Oatis JE, Jr., Walle UK. High absorption but very low bioavailability of oral resveratrol in humans. *Drug Metab Dispos* 2004 December;32(12):1377-82.
- (22) Boocock DJ, Faust GE, Patel KR, Schinas AM, Brown VA, Ducharme MP, Booth TD, Crowell JA, Perloff M, Gescher AJ, Steward WP, Brenner DE. Phase I dose escalation pharmacokinetic study in healthy volunteers of resveratrol, a potential cancer chemopreventive agent. *Cancer Epidemiol Biomarkers Prev* 2007 June;16(6):1246-52.
- (23) Shaw RJ. LKB1 and AMP-activated protein kinase control of mTOR signalling and growth. *Acta Physiol (Oxf)* 2009 May;196(1):65-80.
- (24) Chong ZZ, Shang YC, Zhang L, Wang S, Maiese K. Mammalian target of rapamycin: hitting the bull's-eye for neurological disorders. *Oxid Med Cell Longev* 2010 November;3(6):374-91.
- (25) Blagosklonny MV. Calorie restriction: decelerating mTOR-driven aging from cells to organisms (including humans). *Cell Cycle* 2010 February 15;9(4):683-8.
- (26) Greer EL, Oskoui PR, Banko MR, Maniar JM, Gygi MP, Gygi SP, Brunet A. The energy sensor AMP-activated protein kinase directly regulates the mammalian FOXO3 transcription factor. *J Biol Chem* 2007 October 12;282(41):30107-19.
- (27) Olmos Y, Valle I, Borniquel S, Tierrez A, Soria E, Lamas S, Monsalve M. Mutual dependence of Foxo3a and PGC-1alpha in the induction of oxidative stress genes. *J Biol Chem* 2009 May 22;284(21):14476-84.
- (28) Egan D, Kim J, Shaw RJ, Guan KL. The autophagy initiating kinase ULK1 is regulated via opposing phosphorylation by AMPK and mTOR. *Autophagy* 2011 June;7(6):643-4.

- (29) Hariharan N, Maejima Y, Nakae J, Paik J, DePinho RA, Sadoshima J. Deacetylation of FoxO by Sirt1 Plays an Essential Role in Mediating Starvation-Induced Autophagy in Cardiac Myocytes. *Circ Res* 2010 December 10;107(12):1470-82.
- (30) Canto C, Jiang LQ, Deshmukh AS, Matakci C, Coste A, Lagouge M, Zierath JR, Auwerx J. Interdependence of AMPK and SIRT1 for metabolic adaptation to fasting and exercise in skeletal muscle. *Cell Metab* 2010 March 3;11(3):213-9.
- (31) Bergamini E, Cavallini G, Cecchi L, Donati A, Dolfi C, Gori Z, Innocenti B, Maccheroni M, Marino M, Masini M, Paradiso C, Pollera M, Trentalance A. A proposed mechanism of the antiaging action of diet restriction. *Aging (Milano)* 1998 April;10(2):174-5.
- (32) Cuervo AM. Calorie restriction and aging: the ultimate "cleansing diet". *J Gerontol A Biol Sci Med Sci* 2008 June;63(6):547-9.
- (33) Dhahbi JM, Mote PL, Fahy GM, Spindler SR. Identification of potential caloric restriction mimetics by microarray profiling. *Physiol Genomics* 2005 November 17;23(3):343-50.
- (34) De L, V, La MA, Orvieto R, Morgante G. Effect of metformin on insulin-like growth factor (IGF) I and IGF-binding protein I in polycystic ovary syndrome. *J Clin Endocrinol Metab* 2000 April;85(4):1598-600.
- (35) Jakubowicz DJ, Seppala M, Jakubowicz S, Rodriguez-Armas O, Rivas-Santiago A, Koistinen H, Koistinen R, Nestler JE. Insulin reduction with metformin increases luteal phase serum glycodelin and insulin-like growth factor-binding protein 1 concentrations and enhances uterine vascularity and blood flow in the polycystic ovary syndrome. *J Clin Endocrinol Metab* 2001 March;86(3):1126-33.
- (36) McCarty MF. Chronic activation of AMP-activated kinase as a strategy for slowing aging. *Med Hypotheses* 2004;63(2):334-9.
- (37) Bartke A. Minireview: role of the growth hormone/insulin-like growth factor system in mammalian aging. *Endocrinology* 2005 September;146(9):3718-23.
- (38) Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, Nadon NL, Wilkinson JE, Frenkel K, Carter CS, Pahor M, Javors MA, Fernandez E, Miller RA. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* 2009 July 16;460(7253):392-5.
- (39) Selman C, Tullet JM, Wieser D, Irvine E, Lingard SJ, Choudhury AI, Claret M, Al-Qassab H, Carmignac D, Ramadani F, Woods A, Robinson IC, Schuster E, Batterham RL, Kozma SC, Thomas G, Carling D, Okkenhaug K, Thornton JM, Partridge L, Gems D, Withers DJ. Ribosomal protein S6 kinase 1 signaling regulates mammalian life span. *Science* 2009 October 2;326(5949):140-4.
- (40) Anisimov VN. Metformin for aging and cancer prevention. *Aging (Albany NY)* 2010 November;2(11):760-74.
- (41) Anisimov VN, Berstein LM, Popovich IG, Zabezhinski MA, Egorin PA, Piskunova TS, Semenchenko AV, Tyndyk ML, Yurova MN, Kovalenko IG, Poroshina TE. If started early in life, metformin treatment increases life span and postpones tumors in female SHR mice. *Aging (Albany NY)* 2011 February;3(2):148-57.

- (42) Baur JA, Pearson KJ, Price NL, Jamieson HA, Lerin C, Kalra A, Prabhu VV, Allard JS, Lopez-Lluch G, Lewis K, Pistell PJ, Poosala S, Becker KG, Boss O, Gwinn D, Wang M, Ramaswamy S, Fishbein KW, Spencer RG, Lakatta EG, Le CD, Shaw RJ, Navas P, Puigserver P, Ingram DK, de CR, Sinclair DA. Resveratrol improves health and survival of mice on a high-calorie diet. *Nature* 2006 November 16;444(7117):337-42.
- (43) Smith DL, Jr., Elam CF, Jr., Mattison JA, Lane MA, Roth GS, Ingram DK, Allison DB. Metformin supplementation and life span in Fischer-344 rats. *J Gerontol A Biol Sci Med Sci* 2010 May;65(5):468-74.
- (44) Pearson KJ, Baur JA, Lewis KN, Peshkin L, Price NL, Labinskyy N, Swindell WR, Kamara D, Minor RK, Perez E, Jamieson HA, Zhang Y, Dunn SR, Sharma K, Pleshko N, Woollett LA, Csiszar A, Ikeno Y, Le CD, Elliott PJ, Becker KG, Navas P, Ingram DK, Wolf NS, Ungvari Z, Sinclair DA, de CR. Resveratrol delays age-related deterioration and mimics transcriptional aspects of dietary restriction without extending life span. *Cell Metab* 2008 August;8(2):157-68.
- (45) Varady KA, Hellerstein MK. Alternate-day fasting and chronic disease prevention: a review of human and animal trials. *Am J Clin Nutr* 2007 July;86(1):7-13.
- (46) Johnson JB, Laub DR, John S. The effect on health of alternate day calorie restriction: eating less and more than needed on alternate days prolongs life. *Med Hypotheses* 2006;67(2):209-11.
- (47) Johnson JB, Summer W, Cutler RG, Martin B, Hyun DH, Dixit VD, Pearson M, Nassar M, Telljohann R, Maudsley S, Carlson O, John S, Laub DR, Mattson MP. Alternate day calorie restriction improves clinical findings and reduces markers of oxidative stress and inflammation in overweight adults with moderate asthma. *Free Radic Biol Med* 2007 March 1;42(5):665-74.
- (48) McCarty MF. Could carbohydrate-concentrated diets mimic calorie restriction in slowing the aging process? *Medical Hypotheses* 2011;submitted for publication.
- (49) Zhou G, Myers R, Li Y, Chen Y, Shen X, Fenyk-Melody J, Wu M, Ventre J, Doebber T, Fujii N, Musi N, Hirshman MF, Goodyear LJ, Moller DE. Role of AMP-activated protein kinase in mechanism of metformin action. *J Clin Invest* 2001 October;108(8):1167-74.
- (50) Musi N, Hirshman MF, Nygren J, Svanfeldt M, Bavenholm P, Rooyackers O, Zhou G, Williamson JM, Ljunqvist O, Efendic S, Moller DE, Thorell A, Goodyear LJ. Metformin increases AMP-activated protein kinase activity in skeletal muscle of subjects with type 2 diabetes. *Diabetes* 2002 July;51(7):2074-81.
- (51) Lee YS, Kim WS, Kim KH, Yoon MJ, Cho HJ, Shen Y, Ye JM, Lee CH, Oh WK, Kim CT, Hohnen-Behrens C, Gosby A, Kraegen EW, James DE, Kim JB. Berberine, a natural plant product, activates AMP-activated protein kinase with beneficial metabolic effects in diabetic and insulin-resistant states. *Diabetes* 2006 August;55(8):2256-64.
- (52) Turner N, Li JY, Gosby A, To SW, Cheng Z, Miyoshi H, Taketo MM, Cooney GJ, Kraegen EW, James DE, Hu LH, Li J, Ye JM. Berberine and its more biologically available derivative, dihydroberberine, inhibit mitochondrial respiratory complex I: a mechanism for the action of berberine to activate AMP-activated protein kinase and improve insulin action. *Diabetes* 2008 May;57(5):1414-8.

- (53) Effect of intensive blood-glucose control with metformin on complications in overweight patients with type 2 diabetes (UKPDS 34). UK Prospective Diabetes Study (UKPDS) Group. *Lancet* 1998 September 12;352(9131):854-65.
- (54) Scarpello JH. Improving survival with metformin: the evidence base today. *Diabetes Metab* 2003 September;29(4 Pt 2):6S36-43.
- (55) Holman RR, Paul SK, Bethel MA, Matthews DR, Neil HA. 10-year follow-up of intensive glucose control in type 2 diabetes. *N Engl J Med* 2008 October 9;359(15):1577-89.
- (56) Tzoulaki I, Molokhia M, Curcin V, Little MP, Millett CJ, Ng A, Hughes RI, Khunti K, Wilkins MR, Majeed A, Elliott P. Risk of cardiovascular disease and all cause mortality among patients with type 2 diabetes prescribed oral antidiabetes drugs: retrospective cohort study using UK general practice research database. *BMJ* 2009;339:b4731.
- (57) Masoudi FA, Inzucchi SE, Wang Y, Havranek EP, Foody JM, Krumholz HM. Thiazolidinediones, metformin, and outcomes in older patients with diabetes and heart failure: an observational study. *Circulation* 2005 February 8;111(5):583-90.
- (58) Roussel R, Travert F, Pasquet B, Wilson PW, Smith SC, Jr., Goto S, Ravaud P, Marre M, Porath A, Bhatt DL, Steg PG. Metformin use and mortality among patients with diabetes and atherothrombosis. *Arch Intern Med* 2010 November 22;170(21):1892-9.
- (59) Schramm TK, Gislason GH, Vaag A, Rasmussen JN, Folke F, Hansen ML, Fosbol EL, Kober L, Norgaard ML, Madsen M, Hansen PR, Torp-Pedersen C. Mortality and cardiovascular risk associated with different insulin secretagogues compared with metformin in type 2 diabetes, with or without a previous myocardial infarction: a nationwide study. *Eur Heart J* 2011 August;32(15):1900-8.
- (60) Cooke JP. The pivotal role of nitric oxide for vascular health. *Can J Cardiol* 2004 August;20 Suppl B:7B-15B.
- (61) Massion PB, Balligand JL. Relevance of nitric oxide for myocardial remodeling. *Curr Heart Fail Rep* 2007 March;4(1):18-25.
- (62) Drew BG, Fidge NH, Gallon-Beaumier G, Kemp BE, Kingwell BA. High-density lipoprotein and apolipoprotein AI increase endothelial NO synthase activity by protein association and multisite phosphorylation. *Proc Natl Acad Sci U S A* 2004 May 4;101(18):6999-7004.
- (63) Deng G, Long Y, Yu YR, Li MR. Adiponectin directly improves endothelial dysfunction in obese rats through the AMPK-eNOS Pathway. *Int J Obes (Lond)* 2010 January;34(1):165-71.
- (64) Zhang Y, Lee TS, Kolb EM, Sun K, Lu X, Sladek FM, Kassab GS, Garland T, Jr., Shyy JY. AMP-activated protein kinase is involved in endothelial NO synthase activation in response to shear stress. *Arterioscler Thromb Vasc Biol* 2006 June;26(6):1281-7.
- (65) Young A, Wu W, Sun W, Benjamin LH, Wang N, Li YS, Shyy JY, Chien S, Garcia-Cardena G. Flow activation of AMP-activated protein kinase in vascular endothelium leads to Kruppel-like factor 2 expression. *Arterioscler Thromb Vasc Biol* 2009 November;29(11):1902-8.

- (66) Salminen A, Hyttinen JM, Kaarniranta K. AMP-activated protein kinase inhibits NF-kappaB signaling and inflammation: impact on healthspan and lifespan. *J Mol Med (Berl)* 2011 July;89(7):667-76.
- (67) Yeung F, Hoberg JE, Ramsey CS, Keller MD, Jones DR, Frye RA, Mayo MW. Modulation of NF-kappaB-dependent transcription and cell survival by the SIRT1 deacetylase. *EMBO J* 2004 June 16;23(12):2369-80.
- (68) Wang S, Zhang M, Liang B, Xu J, Xie Z, Liu C, Viollet B, Yan D, Zou MH. AMPKalpha2 deletion causes aberrant expression and activation of NAD(P)H oxidase and consequent endothelial dysfunction in vivo: role of 26S proteasomes. *Circ Res* 2010 April 2;106(6):1117-28.
- (69) Viana R, Aguado C, Esteban I, Moreno D, Viollet B, Knecht E, Sanz P. Role of AMP-activated protein kinase in autophagy and proteasome function. *Biochem Biophys Res Commun* 2008 May 9;369(3):964-8.
- (70) Alba G, El BR, varez-Maqueda M, Chacon P, Vega A, Monteseirin J, Santa MC, Pintado E, Bedoya FJ, Bartrons R, Sobrino F. Stimulators of AMP-activated protein kinase inhibit the respiratory burst in human neutrophils. *FEBS Lett* 2004 August 27;573(1-3):219-25.
- (71) Ceolotto G, Gallo A, Papparella I, Franco L, Murphy E, Iori E, Pagnin E, Fadini GP, Albiero M, Semplicini A, Avogaro A. Rosiglitazone reduces glucose-induced oxidative stress mediated by NAD(P)H oxidase via AMPK-dependent mechanism. *Arterioscler Thromb Vasc Biol* 2007 December;27(12):2627-33.
- (72) McCarty MF, Barroso-Aranda J, Contreras F. AMP-activated kinase may suppress NADPH oxidase activation in vascular tissues. *Med Hypotheses* 2009 April;72(4):468-70.
- (73) Piwkowska A, Rogacka D, Jankowski M, Dominiczak MH, Stepinski JK, Angielski S. Metformin induces suppression of NAD(P)H oxidase activity in podocytes. *Biochem Biophys Res Commun* 2010 March 5;393(2):268-73.
- (74) Bess E, Fisslthaler B, Fromel T, Fleming I. Nitric oxide-induced activation of the AMP-activated protein kinase alpha2 subunit attenuates IkappaB kinase activity and inflammatory responses in endothelial cells. *PLoS ONE* 2011;6(6):e20848.
- (75) Wang S, Xu J, Song P, Viollet B, Zou MH. In vivo activation of AMP-activated protein kinase attenuates diabetes-enhanced degradation of GTP cyclohydrolase I. *Diabetes* 2009 August;58(8):1893-901.
- (76) Li K, Yao W, Zheng X, Liao K. Berberine promotes the development of atherosclerosis and foam cell formation by inducing scavenger receptor A expression in macrophage. *Cell Res* 2009 August;19(8):1006-17.
- (77) Li D, Wang D, Wang Y, Ling W, Feng X, Xia M. Adenosine monophosphate-activated protein kinase induces cholesterol efflux from macrophage-derived foam cells and alleviates atherosclerosis in apolipoprotein E-deficient mice. *J Biol Chem* 2010 October 22;285(43):33499-509.

- (78) Lee TS, Pan CC, Peng CC, Kou YR, Chen CY, Ching LC, Tsai TH, Chen SF, Lyu PC, Shyue SK. Anti-atherogenic effect of berberine on LXRA α -ABCA1-dependent cholesterol efflux in macrophages. *J Cell Biochem* 2010 September 1;111(1):104-10.
- (79) Guan S, Wang B, Li W, Guan J, Fang X. Effects of berberine on expression of LOX-1 and SR-BI in human macrophage-derived foam cells induced by ox-LDL. *Am J Chin Med* 2010;38(6):1161-9.
- (80) Ishii N, Matsumura T, Kinoshita H, Motoshima H, Kojima K, Tsutsumi A, Kawasaki S, Yano M, Senokuchi T, Asano T, Nishikawa T, Araki E. Activation of AMP-activated protein kinase suppresses oxidized low-density lipoprotein-induced macrophage proliferation. *J Biol Chem* 2009 December 11;284(50):34561-9.
- (81) Ching YP, Davies SP, Hardie DG. Analysis of the specificity of the AMP-activated protein kinase by site-directed mutagenesis of bacterially expressed 3-hydroxy 3-methylglutaryl-CoA reductase, using a single primer variant of the unique-site-elimination method. *Eur J Biochem* 1996 May 1;237(3):800-8.
- (82) Kong W, Wei J, Abidi P, Lin M, Inaba S, Li C, Wang Y, Wang Z, Si S, Pan H, Wang S, Wu J, Wang Y, Li Z, Liu J, Jiang JD. Berberine is a novel cholesterol-lowering drug working through a unique mechanism distinct from statins. *Nat Med* 2004 December;10(12):1344-51.
- (83) Abidi P, Zhou Y, Jiang JD, Liu J. Extracellular signal-regulated kinase-dependent stabilization of hepatic low-density lipoprotein receptor mRNA by herbal medicine berberine. *Arterioscler Thromb Vasc Biol* 2005 October;25(10):2170-6.
- (84) Cicero AF, Rovati LC, Setnikar I. Eulipidemic effects of berberine administered alone or in combination with other natural cholesterol-lowering agents. A single-blind clinical investigation. *Arzneimittelforschung* 2007;57(1):26-30.
- (85) Zhang Y, Li X, Zou D, Liu W, Yang J, Zhu N, Huo L, Wang M, Hong J, Wu P, Ren G, Ning G. Treatment of type 2 diabetes and dyslipidemia with the natural plant alkaloid berberine. *J Clin Endocrinol Metab* 2008 July;93(7):2559-65.
- (86) Kong WJ, Wei J, Zuo ZY, Wang YM, Song DQ, You XF, Zhao LX, Pan HN, Jiang JD. Combination of simvastatin with berberine improves the lipid-lowering efficacy. *Metabolism* 2008 August;57(8):1029-37.
- (87) Fu YN, Xiao H, Ma XW, Jiang SY, Xu M, Zhang YY. Metformin attenuates pressure overload-induced cardiac hypertrophy via AMPK activation. *Acta Pharmacol Sin* 2011 July;32(7):879-87.
- (88) Beauloye C, Bertrand L, Horman S, Hue L. AMPK activation, a preventive therapeutic target in the transition from cardiac injury to heart failure. *Cardiovasc Res* 2011 May 1;90(2):224-33.
- (89) Benes J, Kazdova L, Drahota Z, Houstek J, Medrikova D, Kopecky J, Kovarova N, Vrbacky M, Sedmera D, Strnad H, Kolar M, Petrak J, Benada O, Skaroupkova P, Cervenka L, Melenovsky V. Effect of metformin therapy on cardiac function and survival in a volume-overload model of heart failure in rats. *Clin Sci (Lond)* 2011 July 1;121(1):29-41.

- (90) Zhang CX, Pan SN, Meng RS, Peng CQ, Xiong ZJ, Chen BL, Chen GQ, Yao FJ, Chen YL, Ma YD, Dong YG. Metformin attenuates ventricular hypertrophy by activating the AMP-activated protein kinase-endothelial nitric oxide synthase pathway in rats. *Clin Exp Pharmacol Physiol* 2011 January;38(1):55-62.
- (91) Zhang P, Hu X, Xu X, Fassett J, Zhu G, Viollet B, Xu W, Wiczer B, Bernlohr DA, Bache RJ, Chen Y. AMP activated protein kinase- α 2 deficiency exacerbates pressure-overload-induced left ventricular hypertrophy and dysfunction in mice. *Hypertension* 2008 November;52(5):918-24.
- (92) Li HL, Yin R, Chen D, Liu D, Wang D, Yang Q, Dong YG. Long-term activation of adenosine monophosphate-activated protein kinase attenuates pressure-overload-induced cardiac hypertrophy. *J Cell Biochem* 2007 April 1;100(5):1086-99.
- (93) Hong Y, Hui SC, Chan TY, Hou JY. Effect of berberine on regression of pressure-overload induced cardiac hypertrophy in rats. *Am J Chin Med* 2002;30(4):589-99.
- (94) Hong Y, Hui SS, Chan BT, Hou J. Effect of berberine on catecholamine levels in rats with experimental cardiac hypertrophy. *Life Sci* 2003 April 18;72(22):2499-507.
- (95) Zeng XH, Zeng XJ, Li YY. Efficacy and safety of berberine for congestive heart failure secondary to ischemic or idiopathic dilated cardiomyopathy. *Am J Cardiol* 2003 July 15;92(2):173-6.
- (96) Phillips LK, Prins JB. The link between abdominal obesity and the metabolic syndrome. *Curr Hypertens Rep* 2008 April;10(2):156-64.
- (97) Unger RH, Clark GO, Scherer PE, Orci L. Lipid homeostasis, lipotoxicity and the metabolic syndrome. *Biochim Biophys Acta* 2010 March;1801(3):209-14.
- (98) Rasouli N, Molavi B, Elbein SC, Kern PA. Ectopic fat accumulation and metabolic syndrome. *Diabetes Obes Metab* 2007 January;9(1):1-10.
- (99) Julius U. Influence of plasma free fatty acids on lipoprotein synthesis and diabetic dyslipidemia. *Exp Clin Endocrinol Diabetes* 2003 August;111(5):246-50.
- (100) Bulcao C, Ribeiro-Filho FF, Sanudo A, Roberta Ferreira SG. Effects of simvastatin and metformin on inflammation and insulin resistance in individuals with mild metabolic syndrome. *Am J Cardiovasc Drugs* 2007;7(3):219-24.
- (101) Luque-Ramirez M, Escobar-Morreale HF. Treatment of polycystic ovary syndrome (PCOS) with metformin ameliorates insulin resistance in parallel with the decrease of serum interleukin-6 concentrations. *Horm Metab Res* 2010 October;42(11):815-20.
- (102) Fidan E, Onder EH, Yilmaz M, Yilmaz H, Kocak M, Karahan C, Erem C. The effects of rosiglitazone and metformin on inflammation and endothelial dysfunction in patients with type 2 diabetes mellitus. *Acta Diabetol* 2011 March 23.
- (103) Phillips SA, Ciaraldi TP, Kong AP, Bandukwala R, Aroda V, Carter L, Baxi S, Mudaliar SR, Henry RR. Modulation of circulating and adipose tissue adiponectin levels by antidiabetic therapy. *Diabetes* 2003 March;52(3):667-74.

- (104) Sharma PK, Bhansali A, Sialy R, Malhotra S, Pandhi P. Effects of pioglitazone and metformin on plasma adiponectin in newly detected type 2 diabetes mellitus. *Clin Endocrinol (Oxf)* 2006 December;65(6):722-8.
- (105) Tarkun I, Dikmen E, Cetinarslan B, Canturk Z. Impact of treatment with metformin on adipokines in patients with polycystic ovary syndrome. *Eur Cytokine Netw* 2010 December 1;21(4):272-7.
- (106) Abbasi F, Kamath V, Rizvi AA, Carantoni M, Chen YD, Reaven GM. Results of a placebo-controlled study of the metabolic effects of the addition of metformin to sulfonylurea-treated patients. Evidence for a central role of adipose tissue. *Diabetes Care* 1997 December;20(12):1863-9.
- (107) Abbasi F, Carantoni M, Chen YD, Reaven GM. Further evidence for a central role of adipose tissue in the antihyperglycemic effect of metformin. *Diabetes Care* 1998 August;21(8):1301-5.
- (108) Wijkander J, Landstrom TR, Manganiello V, Belfrage P, Degerman E. Insulin-induced phosphorylation and activation of phosphodiesterase 3B in rat adipocytes: possible role for protein kinase B but not mitogen-activated protein kinase or p70 S6 kinase. *Endocrinology* 1998 January;139(1):219-27.
- (109) Garton AJ, Yeaman SJ. Identification and role of the basal phosphorylation site on hormone-sensitive lipase. *Eur J Biochem* 1990 July 20;191(1):245-50.
- (110) Watt MJ, Holmes AG, Pinnamaneni SK, Garnham AP, Steinberg GR, Kemp BE, Febbraio MA. Regulation of HSL serine phosphorylation in skeletal muscle and adipose tissue. *Am J Physiol Endocrinol Metab* 2006 March;290(3):E500-E508.
- (111) Gu Y, Zhang Y, Shi X, Li X, Hong J, Chen J, Gu W, Lu X, Xu G, Ning G. Effect of traditional Chinese medicine berberine on type 2 diabetes based on comprehensive metabonomics. *Talanta* 2010 May 15;81(3):766-72.
- (112) Park H, Kaushik VK, Constant S, Prentki M, Przybytkowski E, Ruderman NB, Saha AK. Coordinate regulation of malonyl-CoA decarboxylase, sn-glycerol-3-phosphate acyltransferase, and acetyl-CoA carboxylase by AMP-activated protein kinase in rat tissues in response to exercise. *J Biol Chem* 2002 September 6;277(36):32571-7.
- (113) Dagher Z, Ruderman N, Tornheim K, Ido Y. Acute regulation of fatty acid oxidation and amp-activated protein kinase in human umbilical vein endothelial cells. *Circ Res* 2001 June 22;88(12):1276-82.
- (114) McGarry JD, Leatherman GF, Foster DW. Carnitine palmitoyltransferase I. The site of inhibition of hepatic fatty acid oxidation by malonyl-CoA. *J Biol Chem* 1978 June 25;253(12):4128-36.
- (115) Muoio DM, Seefeld K, Witters LA, Coleman RA. AMP-activated kinase reciprocally regulates triacylglycerol synthesis and fatty acid oxidation in liver and muscle: evidence that sn-glycerol-3-phosphate acyltransferase is a novel target. *Biochem J* 1999 March 15;338 (Pt 3):783-91.

- (116) Knowler WC, Barrett-Connor E, Fowler SE, Hamman RF, Lachin JM, Walker EA, Nathan DM. Reduction in the incidence of type 2 diabetes with lifestyle intervention or metformin. *N Engl J Med* 2002 February 7;346(6):393-403.
- (117) El-Assaad W, Buteau J, Peyot ML, Nolan C, Roduit R, Hardy S, Joly E, Dbaibo G, Rosenberg L, Prentki M. Saturated fatty acids synergize with elevated glucose to cause pancreatic beta-cell death. *Endocrinology* 2003 September;144(9):4154-63.
- (118) Zhou J, Zhou S, Tang J, Zhang K, Guang L, Huang Y, Xu Y, Ying Y, Zhang L, Li D. Protective effect of berberine on beta cells in streptozotocin- and high-carbohydrate/high-fat diet-induced diabetic rats. *Eur J Pharmacol* 2009 March 15;606(1-3):262-8.
- (119) Gao N, Zhao TY, Li XJ. The protective effect of berberine on beta-cell lipoapoptosis. *J Endocrinol Invest* 2011 February;34(2):124-30.
- (120) Kirpichnikov D, McFarlane SI, Sowers JR. Metformin: an update. *Ann Intern Med* 2002 July 2;137(1):25-33.
- (121) He L, Sabet A, Djedjos S, Miller R, Sun X, Hussain MA, Radovick S, Wondisford FE. Metformin and insulin suppress hepatic gluconeogenesis through phosphorylation of CREB binding protein. *Cell* 2009 May 15;137(4):635-46.
- (122) Koo SH, Flechner L, Qi L, Zhang X, Sreaton RA, Jeffries S, Hedrick S, Xu W, Boussouar F, Brindle P, Takemori H, Montminy M. The CREB coactivator TORC2 is a key regulator of fasting glucose metabolism. *Nature* 2005 October 20;437(7062):1109-11.
- (123) Kim YD, Park KG, Lee YS, Park YY, Kim DK, Nedumaran B, Jang WG, Cho WJ, Ha J, Lee IK, Lee CH, Choi HS. Metformin inhibits hepatic gluconeogenesis through AMP-activated protein kinase-dependent regulation of the orphan nuclear receptor SHP. *Diabetes* 2008 February;57(2):306-14.
- (124) Lee JM, Seo WY, Song KH, Chanda D, Kim YD, Kim DK, Lee MW, Ryu D, Kim YH, Noh JR, Lee CH, Chiang JY, Koo SH, Choi HS. AMPK-dependent repression of hepatic gluconeogenesis via disruption of CREB-CRTC2 complex by orphan nuclear receptor small heterodimer partner. *J Biol Chem* 2010 October 15;285(42):32182-91.
- (125) Xia X, Yan J, Shen Y, Tang K, Yin J, Zhang Y, Yang D, Liang H, Ye J, Weng J. Berberine improves glucose metabolism in diabetic rats by inhibition of hepatic gluconeogenesis. *PLoS ONE* 2011;6(2):e16556.
- (126) Meneghini LF, Orozco-Beltran D, Khunti K, Caputo S, Damci T, Liebl A, Ross SA. Weight Beneficial Treatments for Type 2 Diabetes. *J Clin Endocrinol Metab* 2011 September 7.
- (127) Ben-Shlomo S, Zvibel I, Shnell M, Shlomain A, Chepurko E, Halpern Z, Barzilai N, Oren R, Fishman S. Glucagon-like peptide-1 reduces hepatic lipogenesis via activation of AMP-activated protein kinase. *J Hepatol* 2011 June;54(6):1214-23.
- (128) Harborne LR, Sattar N, Norman JE, Fleming R. Metformin and weight loss in obese women with polycystic ovary syndrome: comparison of doses. *J Clin Endocrinol Metab* 2005 August;90(8):4593-8.

- (129) Nieuwenhuis-Ruifrok AE, Kuchenbecker WK, Hoek A, Middleton P, Norman RJ. Insulin sensitizing drugs for weight loss in women of reproductive age who are overweight or obese: systematic review and meta-analysis. *Hum Reprod Update* 2009 January;15(1):57-68.
- (130) Praharaj SK, Jana AK, Goyal N, Sinha VK. Metformin for olanzapine-induced weight gain: a systematic review and meta-analysis. *Br J Clin Pharmacol* 2011 March;71(3):377-82.
- (131) Desilets AR, Dhakal-Karki S, Dunican KC. Role of metformin for weight management in patients without type 2 diabetes. *Ann Pharmacother* 2008 June;42(6):817-26.
- (132) Scharrer E. Control of food intake by fatty acid oxidation and ketogenesis. *Nutrition* 1999 September;15(9):704-14.
- (133) Leonhardt M, Langhans W. Fatty acid oxidation and control of food intake. *Physiol Behav* 2004 December 30;83(4):645-51.
- (134) St-Onge MP, Jones PJ. Physiological effects of medium-chain triglycerides: potential agents in the prevention of obesity. *J Nutr* 2002 March;132(3):329-32.
- (135) Foretz M, Ancellin N, Andreelli F, Saintillan Y, Grondin P, Kahn A, Thorens B, Vaulont S, Viollet B. Short-term overexpression of a constitutively active form of AMP-activated protein kinase in the liver leads to mild hypoglycemia and fatty liver. *Diabetes* 2005 May;54(5):1331-9.
- (136) Olmos Y, Valle I, Borniquel S, Tierrez A, Soria E, Lamas S, Monsalve M. Mutual dependence of Foxo3a and PGC-1alpha in the induction of oxidative stress genes. *J Biol Chem* 2009 May 22;284(21):14476-84.
- (137) Xie Z, Zhang J, Wu J, Viollet B, Zou MH. Upregulation of mitochondrial uncoupling protein-2 by the AMP-activated protein kinase in endothelial cells attenuates oxidative stress in diabetes. *Diabetes* 2008 December;57(12):3222-30.
- (138) McCarty MF. Hepatothermic therapy of obesity: rationale and an inventory of resources. *Med Hypotheses* 2001 September;57(3):324-36.
- (139) McCarty MF. High mitochondrial redox potential may promote induction and activation of UCP2 in hepatocytes during hepatothermic therapy. *Med Hypotheses* 2005;64(6):1216-9.
- (140) Yap F, Craddock L, Yang J. Mechanism of AMPK suppression of LXR-dependent Srebp-1c transcription. *Int J Biol Sci* 2011;7(5):645-50.
- (141) Li Y, Xu S, Mihaylova MM, Zheng B, Hou X, Jiang B, Park O, Luo Z, Lefai E, Shyy JY, Gao B, Wierzbicki M, Verbeuren TJ, Shaw RJ, Cohen RA, Zang M. AMPK phosphorylates and inhibits SREBP activity to attenuate hepatic steatosis and atherosclerosis in diet-induced insulin-resistant mice. *Cell Metab* 2011 April 6;13(4):376-88.
- (142) Hellerstein MK. De novo lipogenesis in humans: metabolic and regulatory aspects. *Eur J Clin Nutr* 1999 April;53 Suppl 1:S53-S65.

- (143) Kondo T, Kishi M, Fushimi T, Ugajin S, Kaga T. Vinegar intake reduces body weight, body fat mass, and serum triglyceride levels in obese Japanese subjects. *Biosci Biotechnol Biochem* 2009 August;73(8):1837-43.
- (144) Kondo T, Kishi M, Fushimi T, Kaga T. Acetic acid upregulates the expression of genes for fatty acid oxidation enzymes in liver to suppress body fat accumulation. *J Agric Food Chem* 2009 July 8;57(13):5982-6.
- (145) Evans JM, Donnelly LA, Emslie-Smith AM, Alessi DR, Morris AD. Metformin and reduced risk of cancer in diabetic patients. *BMJ* 2005 June 4;330(7503):1304-5.
- (146) Libby G, Donnelly LA, Donnan PT, Alessi DR, Morris AD, Evans JM. New users of metformin are at low risk of incident cancer: a cohort study among people with type 2 diabetes. *Diabetes Care* 2009 September;32(9):1620-5.
- (147) Currie CJ, Poole CD, Gale EA. The influence of glucose-lowering therapies on cancer risk in type 2 diabetes. *Diabetologia* 2009 September;52(9):1766-77.
- (148) Wright JL, Stanford JL. Metformin use and prostate cancer in Caucasian men: results from a population-based case-control study. *Cancer Causes Control* 2009 November;20(9):1617-22.
- (149) Landman GW, Kleefstra N, van Hateren KJ, Groenier KH, Gans RO, Bilo HJ. Metformin associated with lower cancer mortality in type 2 diabetes: ZODIAC-16. *Diabetes Care* 2010 February;33(2):322-6.
- (150) Bodmer M, Meier C, Krahenbuhl S, Jick SS, Meier CR, Meier CR. Long-term metformin use is associated with decreased risk of breast cancer. *Diabetes Care* 2010 March 18.
- (151) Bowker SL, Yasui Y, Veugelers P, Johnson JA. Glucose-lowering agents and cancer mortality rates in type 2 diabetes: assessing effects of time-varying exposure. *Diabetologia* 2010 April 21.
- (152) Zhang ZJ, Zheng ZJ, Kan H, Song Y, Cui W, Zhao G, Kip KE. Reduced Risk of Colorectal Cancer With Metformin Therapy in Patients With Type 2 Diabetes: A meta-analysis. *Diabetes Care* 2011 October;34(10):2323-8.
- (153) Decensi A, Puntoni M, Goodwin P, Cazzaniga M, Gennari A, Bonanni B, Gandini S. Metformin and cancer risk in diabetic patients: a systematic review and meta-analysis. *Cancer Prev Res (Phila)* 2010 November;3(11):1451-61.
- (154) McCarty MF. mTORC1 activity as a determinant of cancer risk - Rationalizing the cancer-preventive effects of adiponectin, metformin, rapamycin, and low-protein vegan diets. *Med Hypotheses* 2011 October;77(4):642-8.
- (155) Gingras AC, Raught B, Gygi SP, Niedzwiecka A, Miron M, Burley SK, Polakiewicz RD, Wyslouch-Cieszynska A, Aebersold R, Sonenberg N. Hierarchical phosphorylation of the translation inhibitor 4E-BP1. *Genes Dev* 2001 November 1;15(21):2852-64.
- (156) Culjkovic B, Topisirovic I, Borden KL. Controlling gene expression through RNA regulons: the role of the eukaryotic translation initiation factor eIF4E. *Cell Cycle* 2007 January 1;6(1):65-9.

- (157) Mamane Y, Petroulakis E, Rong L, Yoshida K, Ler LW, Sonenberg N. eIF4E--from translation to transformation. *Oncogene* 2004 April 19;23(18):3172-9.
- (158) De BA, Graff JR. eIF-4E expression and its role in malignancies and metastases. *Oncogene* 2004 April 19;23(18):3189-99.
- (159) Graff JR, Konicek BW, Carter JH, Marcusson EG. Targeting the eukaryotic translation initiation factor 4E for cancer therapy. *Cancer Res* 2008 February 1;68(3):631-4.
- (160) Smith MR, Jaramillo M, Liu YL, Dever TE, Merrick WC, Kung HF, Sonenberg N. Translation initiation factors induce DNA synthesis and transform NIH 3T3 cells. *New Biol* 1990 July;2(7):648-54.
- (161) Kisfalvi K, Rey O, Young SH, Sinnott-Smith J, Rozengurt E. Insulin potentiates Ca²⁺ signaling and phosphatidylinositol 4,5-bisphosphate hydrolysis induced by Gq protein-coupled receptor agonists through an mTOR-dependent pathway. *Endocrinology* 2007 July;148(7):3246-57.
- (162) Rozengurt E, Sinnott-Smith J, Kisfalvi K. Crosstalk between insulin/insulin-like growth factor-1 receptors and G protein-coupled receptor signaling systems: a novel target for the antidiabetic drug metformin in pancreatic cancer. *Clin Cancer Res* 2010 May 1;16(9):2505-11.
- (163) Kisfalvi K, Eibl G, Sinnott-Smith J, Rozengurt E. Metformin disrupts crosstalk between G protein-coupled receptor and insulin receptor signaling systems and inhibits pancreatic cancer growth. *Cancer Res* 2009 August 15;69(16):6539-45.
- (164) Jalving M, Gietema JA, Lefrandt JD, de JS, Reyners AK, Gans RO, de Vries EG. Metformin: taking away the candy for cancer? *Eur J Cancer* 2010 September;46(13):2369-80.
- (165) Micic D, Cvijovic G, Trajkovic V, Duntas LH, Polovina S. Metformin: its emerging role in oncology. *Hormones (Athens)* 2011 January;10(1):5-15.
- (166) Dowling RJ, Goodwin PJ, Stambolic V. Understanding the benefit of metformin use in cancer treatment. *BMC Med* 2011;9:33.
- (167) Jiralerspong S, Palla SL, Giordano SH, Meric-Bernstam F, Liedtke C, Barnett CM, Hsu L, Hung MC, Hortobagyi GN, Gonzalez-Angulo AM. Metformin and pathologic complete responses to neoadjuvant chemotherapy in diabetic patients with breast cancer. *J Clin Oncol* 2009 July 10;27(20):3297-302.
- (168) Tan BX, Yao WX, Ge J, Peng XC, Du XB, Zhang R, Yao B, Xie K, Li LH, Dong H, Gao F, Zhao F, Hou JM, Su JM, Liu JY. Prognostic influence of metformin as first-line chemotherapy for advanced nonsmall cell lung cancer in patients with type 2 diabetes. *Cancer* 2011 April 26.
- (169) Lee JH, Kim TI, Jeon SM, Hong SP, Cheon JH, Kim WH. The effects of metformin on the survival of colorectal cancer patients with diabetes mellitus. *Int J Cancer* 2011 September 12.
- (170) Bo S, Ciccone G, Rosato R, Villosio P, Appendino G, Ghigo E, Grassi G. Cancer mortality reduction and metformin. A retrospective cohort study in type 2 diabetic patients. *Diabetes Obes Metab* 2011 August 3.

- (171) Hirsch HA, Iliopoulos D, Tsiichlis PN, Struhl K. Metformin selectively targets cancer stem cells, and acts together with chemotherapy to block tumor growth and prolong remission. *Cancer Res* 2009 October 1;69(19):7507-11.
- (172) Iliopoulos D, Hirsch HA, Struhl K. Metformin decreases the dose of chemotherapy for prolonging tumor remission in mouse xenografts involving multiple cancer cell types. *Cancer Res* 2011 May 1;71(9):3196-201.
- (173) Vazquez-Martin A, Oliveras-Ferraro C, Del BS, Martin-Castillo B, Menendez JA. The anti-diabetic drug metformin suppresses self-renewal and proliferation of trastuzumab-resistant tumor-initiating breast cancer stem cells. *Breast Cancer Res Treat* 2011 April;126(2):355-64.
- (174) Vazquez-Martin A, Oliveras-Ferraro C, Cufi S, Del BS, Martin-Castillo B, Menendez JA. Metformin regulates breast cancer stem cell ontogeny by transcriptional regulation of the epithelial-mesenchymal transition (EMT) status. *Cell Cycle* 2010 September 15;9(18):3807-14.
- (175) Sun Y, Xun K, Wang Y, Chen X. A systematic review of the anticancer properties of berberine, a natural product from Chinese herbs. *Anticancer Drugs* 2009 October;20(9):757-69.
- (176) Tang J, Feng Y, Tsao S, Wang N, Curtain R, Wang Y. Berberine and Coptidis rhizoma as novel antineoplastic agents: a review of traditional use and biomedical investigations. *J Ethnopharmacol* 2009 October 29;126(1):5-17.
- (177) Diogo CV, Machado NG, Barbosa IA, Serafim TL, Burgeiro A, Oliveira PJ. Berberine as a promising safe anti-cancer agent - is there a role for mitochondria? *Curr Drug Targets* 2011 June;12(6):850-9.
- (178) Song G, Ouyang G, Bao S. The activation of Akt/PKB signaling pathway and cell survival. *J Cell Mol Med* 2005 January;9(1):59-71.
- (179) Simpson ER, Misso M, Hewitt KN, Hill RA, Boon WC, Jones ME, Kovacic A, Zhou J, Clyne CD. Estrogen--the good, the bad, and the unexpected. *Endocr Rev* 2005 May;26(3):322-30.
- (180) Brown KA, McInnes KJ, Hunger NI, Oakhill JS, Steinberg GR, Simpson ER. Subcellular localization of cyclic AMP-responsive element binding protein-regulated transcription coactivator 2 provides a link between obesity and breast cancer in postmenopausal women. *Cancer Res* 2009 July 1;69(13):5392-9.
- (181) Brown KA, Hunger NI, Docanto M, Simpson ER. Metformin inhibits aromatase expression in human breast adipose stromal cells via stimulation of AMP-activated protein kinase. *Breast Cancer Res Treat* 2010 September;123(2):591-6.
- (182) Geisler J, Haynes B, Ekse D, Dowsett M, Lonning PE. Total body aromatization in postmenopausal breast cancer patients is strongly correlated to plasma leptin levels. *J Steroid Biochem Mol Biol* 2007 April;104(1-2):27-34.
- (183) Banerjee R, Beal MF, Thomas B. Autophagy in neurodegenerative disorders: pathogenic roles and therapeutic implications. *Trends Neurosci* 2010 December;33(12):541-9.
- (184) Xilouri M, Stefanis L. Autophagy in the central nervous system: implications for neurodegenerative disorders. *CNS Neurol Disord Drug Targets* 2010 December;9(6):701-19.

- (185) Puyal J, Ginet V, Grishchuk Y, Truttmann AC, Clarke PG. Neuronal Autophagy as a Mediator of Life and Death: Contrasting Roles in Chronic Neurodegenerative and Acute Neural Disorders. *Neuroscientist* 2011 April 27.
- (186) Jimenez-Sanchez M, Thompson F, Zavodsky E, Rubinsztein DC. Autophagy and polyglutamine diseases. *Prog Neurobiol* 2011 September 10.
- (187) Hara T, Nakamura K, Matsui M, Yamamoto A, Nakahara Y, Suzuki-Migishima R, Yokoyama M, Mishima K, Saito I, Okano H, Mizushima N. Suppression of basal autophagy in neural cells causes neurodegenerative disease in mice. *Nature* 2006 June 15;441(7095):885-9.
- (188) Li J, Benashski SE, Venna VR, McCullough LD. Effects of metformin in experimental stroke. *Stroke* 2010 November;41(11):2645-52.
- (189) Salminen A, Kaarniranta K, Haapasalo A, Soininen H, Hiltunen M. AMP-activated protein kinase: a potential player in Alzheimer's disease. *J Neurochem* 2011 August;118(4):460-74.
- (190) Vingtdeux V, Giliberto L, Zhao H, Chandakkar P, Wu Q, Simon JE, Janle EM, Lobo J, Ferruzzi MG, Davies P, Marambaud P. AMP-activated protein kinase signaling activation by resveratrol modulates amyloid-beta peptide metabolism. *J Biol Chem* 2010 March 19;285(12):9100-13.
- (191) Vingtdeux V, Chandakkar P, Zhao H, d'Abramo C, Davies P, Marambaud P. Novel synthetic small-molecule activators of AMPK as enhancers of autophagy and amyloid-beta peptide degradation. *FASEB J* 2011 January;25(1):219-31.
- (192) Marambaud P, Zhao H, Davies P. Resveratrol promotes clearance of Alzheimer's disease amyloid-beta peptides. *J Biol Chem* 2005 November 11;280(45):37377-82.
- (193) Chen Y, Zhou K, Wang R, Liu Y, Kwak YD, Ma T, Thompson RC, Zhao Y, Smith L, Gasparini L, Luo Z, Xu H, Liao FF. Antidiabetic drug metformin (GlucophageR) increases biogenesis of Alzheimer's amyloid peptides via up-regulating BACE1 transcription. *Proc Natl Acad Sci U S A* 2009 March 10;106(10):3907-12.
- (194) Won JS, Im YB, Kim J, Singh AK, Singh I. Involvement of AMP-activated-protein-kinase (AMPK) in neuronal amyloidogenesis. *Biochem Biophys Res Commun* 2010 September 3;399(4):487-91.
- (195) Greco SJ, Sarkar S, Johnston JM, Tezapsidis N. Leptin regulates tau phosphorylation and amyloid through AMPK in neuronal cells. *Biochem Biophys Res Commun* 2009 February 27;380(1):98-104.
- (196) Min SW, Cho SH, Zhou Y, Schroeder S, Haroutunian V, Seeley WW, Huang EJ, Shen Y, Masliah E, Mukherjee C, Meyers D, Cole PA, Ott M, Gan L. Acetylation of tau inhibits its degradation and contributes to tauopathy. *Neuron* 2010 September 23;67(6):953-66.
- (197) Thornton C, Bright NJ, Sastre M, Muckett PJ, Carling D. AMP-activated protein kinase (AMPK) is a tau kinase, activated in response to amyloid beta-peptide exposure. *Biochem J* 2011 March 15;434(3):503-12.

- (198) Austin SA, Santhanam AV, Katusic ZS. Endothelial Nitric Oxide Modulates Expression and Processing of Amyloid Precursor Protein. *Circ Res* 2010 December 2.
- (199) Monsuez JJ, Gesquiere-Dando A, Rivera S. Cardiovascular prevention of cognitive decline. *Cardiol Res Pract* 2011;2011:250970.
- (200) Brown GC. Mechanisms of inflammatory neurodegeneration: iNOS and NADPH oxidase. *Biochem Soc Trans* 2007 November;35(Pt 5):1119-21.
- (201) Lu DY, Tang CH, Chen YH, Wei IH. Berberine suppresses neuroinflammatory responses through AMP-activated protein kinase activation in BV-2 microglia. *J Cell Biochem* 2010 June 1;110(3):697-705.
- (202) Wang W, Fan J, Yang X, Furer-Galban S, Lopez dS, I, von KC, Guo J, Georas SN, Fougelle F, Hardie DG, Carling D, Gorospe M. AMP-activated kinase regulates cytoplasmic HuR. *Mol Cell Biol* 2002 May;22(10):3425-36.
- (203) Wang W, Yang X, Kawai T, Lopez dS, I, Mazan-Mamczarz K, Chen P, Chook YM, Quensel C, Kohler M, Gorospe M. AMP-activated protein kinase-regulated phosphorylation and acetylation of importin alpha1: involvement in the nuclear import of RNA-binding protein HuR. *J Biol Chem* 2004 November 12;279(46):48376-88.
- (204) Di MS, Mazroui R, Dallaire P, Chittur S, Tenenbaum SA, Radzioch D, Marette A, Gallouzi IE. NF-kappa B-mediated MyoD decay during muscle wasting requires nitric oxide synthase mRNA stabilization, HuR protein, and nitric oxide release. *Mol Cell Biol* 2005 August;25(15):6533-45.
- (205) Linker K, Pautz A, Fechir M, Hubrich T, Greeve J, Kleinert H. Involvement of KSRP in the post-transcriptional regulation of human iNOS expression-complex interplay of KSRP with TTP and HuR. *Nucleic Acids Res* 2005;33(15):4813-27.
- (206) Sengupta S, Jang BC, Wu MT, Paik JH, Furneaux H, Hla T. The RNA-binding protein HuR regulates the expression of cyclooxygenase-2. *J Biol Chem* 2003 July 4;278(27):25227-33.
- (207) Li F, Wang HD, Lu DX, Wang YP, Qi RB, Fu YM, Li CJ. Neutral sulfate berberine modulates cytokine secretion and increases survival in endotoxemic mice. *Acta Pharmacol Sin* 2006 September;27(9):1199-205.
- (208) Rothwell PM, Fowkes FG, Belch JF, Ogawa H, Warlow CP, Meade TW. Effect of daily aspirin on long-term risk of death due to cancer: analysis of individual patient data from randomised trials. *Lancet* 2010 December 6.
- (209) Hsu CC, Wahlqvist ML, Lee MS, Tsai HN. Incidence of dementia is increased in type 2 diabetes and reduced by the use of sulfonylureas and metformin. *J Alzheimers Dis* 2011;24(3):485-93.
- (210) Vestergaard P, Rejnmark L, Mosekilde L. Relative fracture risk in patients with diabetes mellitus, and the impact of insulin and oral antidiabetic medication on relative fracture risk. *Diabetologia* 2005 July;48(7):1292-9.

- (211) Rejnmark L. Bone effects of glitazones and other anti-diabetic drugs. *Curr Drug Saf* 2008 September;3(3):194-8.
- (212) Jang WG, Kim EJ, Bae IH, Lee KN, Kim YD, Kim DK, Kim SH, Lee CH, Franceschi RT, Choi HS, Koh JT. Metformin induces osteoblast differentiation via orphan nuclear receptor SHP-mediated transactivation of Runx2. *Bone* 2011 April 1;48(4):885-93.
- (213) Cortizo AM, Sedlinsky C, McCarthy AD, Blanco A, Schurman L. Osteogenic actions of the anti-diabetic drug metformin on osteoblasts in culture. *Eur J Pharmacol* 2006 April 24;536(1-2):38-46.
- (214) Kanazawa I, Yamaguchi T, Yano S, Yamauchi M, Sugimoto T. Metformin enhances the differentiation and mineralization of osteoblastic MC3T3-E1 cells via AMP kinase activation as well as eNOS and BMP-2 expression. *Biochem Biophys Res Commun* 2008 October 24;375(3):414-9.
- (215) Mai QG, Zhang ZM, Xu S, Lu M, Zhou RP, Zhao L, Jia CH, Wen ZH, Jin DD, Bai XC. Metformin stimulates osteoprotegerin and reduces RANKL expression in osteoblasts and ovariectomized rats. *J Cell Biochem* 2011 October;112(10):2902-9.
- (216) Hu JP, Nishishita K, Sakai E, Yoshida H, Kato Y, Tsukuba T, Okamoto K. Berberine inhibits RANKL-induced osteoclast formation and survival through suppressing the NF-kappaB and Akt pathways. *Eur J Pharmacol* 2008 February 2;580(1-2):70-9.
- (217) Lee YS, Kim YS, Lee SY, Kim GH, Kim BJ, Lee SH, Lee KU, Kim GS, Kim SW, Koh JM. AMP kinase acts as a negative regulator of RANKL in the differentiation of osteoclasts. *Bone* 2010 November;47(5):926-37.
- (218) Gao Y, Li Y, Xue J, Jia Y, Hu J. Effect of the anti-diabetic drug metformin on bone mass in ovariectomized rats. *Eur J Pharmacol* 2010 June 10;635(1-3):231-6.
- (219) Li H, Miyahara T, Tezuka Y, Namba T, Suzuki T, Dowaki R, Watanabe M, Nemoto N, Tonami S, Seto H, Kadota S. The effect of kampo formulae on bone resorption in vitro and in vivo. II. Detailed study of berberine. *Biol Pharm Bull* 1999 April;22(4):391-6.
- (220) Armour KE, Armour KJ, Gallagher ME, Godecke A, Helfrich MH, Reid DM, Ralston SH. Defective bone formation and anabolic response to exogenous estrogen in mice with targeted disruption of endothelial nitric oxide synthase. *Endocrinology* 2001 February;142(2):760-6.
- (221) Samuels A, Perry MJ, Gibson RL, Colley S, Tobias JH. Role of endothelial nitric oxide synthase in estrogen-induced osteogenesis. *Bone* 2001 July;29(1):24-9.
- (222) Pelletier JP, DiBattista JA, Roughley P, McCollum R, Martel-Pelletier J. Cytokines and inflammation in cartilage degradation. *Rheum Dis Clin North Am* 1993 August;19(3):545-68.
- (223) Lo YY, Conquer JA, Grinstein S, Cruz TF. Interleukin-1 beta induction of c-fos and collagenase expression in articular chondrocytes: involvement of reactive oxygen species. *J Cell Biochem* 1998 April 1;69(1):19-29.

- (224) Grange L, Nguyen MV, Lardy B, Derouazi M, Campion Y, Trocme C, Paelet MH, Gaudin P, Morel F. NAD(P)H oxidase activity of Nox4 in chondrocytes is both inducible and involved in collagenase expression. *Antioxid Redox Signal* 2006 September;8(9-10):1485-96.
- (225) Ahmad R, Sylvester J, Ahmad M, Zafarullah M. Involvement of H-Ras and reactive oxygen species in proinflammatory cytokine-induced matrix metalloproteinase-13 expression in human articular chondrocytes. *Arch Biochem Biophys* 2011 March 15;507(2):350-5.
- (226) Pelletier JP, Jovanovic D, Fernandes JC, Manning P, Connor JR, Currie MG, Di Battista JA, Martel-Pelletier J. Reduced progression of experimental osteoarthritis in vivo by selective inhibition of inducible nitric oxide synthase. *Arthritis Rheum* 1998 July;41(7):1275-86.
- (227) Pelletier JP, Lascau-Coman V, Jovanovic D, Fernandes JC, Manning P, Connor JR, Currie MG, Martel-Pelletier J. Selective inhibition of inducible nitric oxide synthase in experimental osteoarthritis is associated with reduction in tissue levels of catabolic factors. *J Rheumatol* 1999 September;26(9):2002-14.
- (228) Terkeltaub R, Yang B, Lotz M, Liu-Bryan R. Chondrocyte AMP-activated protein kinase activity suppresses matrix degradation responses to proinflammatory cytokines interleukin-1beta and tumor necrosis factor alpha. *Arthritis Rheum* 2011 July;63(7):1928-37.
- (229) Hu PF, Chen WP, Tang JL, Bao JP, Wu LD. Protective effects of berberine in an experimental rat osteoarthritis model. *Phytother Res* 2011 June;25(6):878-85.
- (230) Nath N, Khan M, Paintlia MK, Singh I, Hoda MN, Giri S. Metformin attenuated the autoimmune disease of the central nervous system in animal models of multiple sclerosis. *J Immunol* 2009 June 15;182(12):8005-14.
- (231) Qin X, Guo BT, Wan B, Fang L, Lu L, Wu L, Zang YQ, Zhang JZ. Regulation of Th1 and Th17 cell differentiation and amelioration of experimental autoimmune encephalomyelitis by natural product compound berberine. *J Immunol* 2010 August 1;185(3):1855-63.
- (232) Ma X, Jiang Y, Wu A, Chen X, Pi R, Liu M, Liu Y. Berberine attenuates experimental autoimmune encephalomyelitis in C57 BL/6 mice. *PLoS ONE* 2010;5(10):e13489.
- (233) Nath N, Giri S, Prasad R, Salem ML, Singh AK, Singh I. 5-aminoimidazole-4-carboxamide ribonucleoside: a novel immunomodulator with therapeutic efficacy in experimental autoimmune encephalomyelitis. *J Immunol* 2005 July 1;175(1):566-74.
- (234) Nath N, Khan M, Rattan R, Mangalam A, Makkar RS, de MC, Bertrand L, Singh I, Chen Y, Viollet B, Giri S. Loss of AMPK exacerbates experimental autoimmune encephalomyelitis disease severity. *Biochem Biophys Res Commun* 2009 August 14;386(1):16-20.
- (235) Zhou H, Mineshita S. The effect of berberine chloride on experimental colitis in rats in vivo and in vitro. *J Pharmacol Exp Ther* 2000 September;294(3):822-9.
- (236) Lee IA, Hyun YJ, Kim DH. Berberine ameliorates TNBS-induced colitis by inhibiting lipid peroxidation, enterobacterial growth and NF-kappaB activation. *Eur J Pharmacol* 2010 December 1;648(1-3):162-70.

- (237) Bai A, Yong M, Ma AG, Ma Y, Weiss CR, Guan Q, Bernstein CN, Peng Z. Novel anti-inflammatory action of 5-aminoimidazole-4-carboxamide ribonucleoside with protective effect in dextran sulfate sodium-induced acute and chronic colitis. *J Pharmacol Exp Ther* 2010 June;333(3):717-25.
- (238) Bai A, Ma AG, Yong M, Weiss CR, Ma Y, Guan Q, Bernstein CN, Peng Z. AMPK agonist downregulates innate and adaptive immune responses in TNBS-induced murine acute and relapsing colitis. *Biochem Pharmacol* 2010 December 1;80(11):1708-17.
- (239) Salpeter SR, Greyber E, Pasternak GA, Salpeter EE. Risk of fatal and nonfatal lactic acidosis with metformin use in type 2 diabetes mellitus. *Cochrane Database Syst Rev* 2010;(4):CD002967.
- (240) Gui SY, Wu L, Peng DY, Liu QY, Yin BP, Shen JZ. Preparation and evaluation of a microemulsion for oral delivery of berberine. *Pharmazie* 2008 July;63(7):516-9.
- (241) Battu SK, Repka MA, Maddineni S, Chittiboyina AG, Avery MA, Majumdar S. Physicochemical characterization of berberine chloride: a perspective in the development of a solution dosage form for oral delivery. *AAPS PharmSciTech* 2010 September;11(3):1466-75.
- (242) Dao TM, Waget A, Klopp P, Serino M, Vachoux C, Pechere L, Drucker DJ, Champion S, Barthelemy S, Barra Y, Burcelin R, Seree E. Resveratrol increases glucose induced GLP-1 secretion in mice: a mechanism which contributes to the glycemic control. *PLoS ONE* 2011;6(6):e20700.
- (243) Mulherin AJ, Oh AH, Kim H, Grieco A, Lauffer LM, Brubaker PL. Mechanisms Underlying Metformin-Induced Secretion of Glucagon-Like Peptide-1 from the Intestinal L Cell. *Endocrinology* 2011 October 4.
- (244) Yu Y, Liu L, Wang X, Liu X, Liu X, Xie L, Wang G. Modulation of glucagon-like peptide-1 release by berberine: in vivo and in vitro studies. *Biochem Pharmacol* 2010 April 1;79(7):1000-6.
- (245) Freeland KR, Wolever TM. Acute effects of intravenous and rectal acetate on glucagon-like peptide-1, peptide YY, ghrelin, adiponectin and tumour necrosis factor-alpha. *Br J Nutr* 2010 February;103(3):460-6.
- (246) Ben-Shlomo S, Zvibel I, Shnell M, Shlomain A, Chepurko E, Halpern Z, Barzilai N, Oren R, Fishman S. Glucagon-like peptide-1 reduces hepatic lipogenesis via activation of AMP-activated protein kinase. *J Hepatol* 2011 June;54(6):1214-23.
- (247) Huisamen B, Genade S, Lochner A. Signalling pathways activated by glucagon-like peptide-1 (7-36) amide in the rat heart and their role in protection against ischaemia. *Cardiovasc J Afr* 2008 March;19(2):77-83.
- (248) Hattori Y, Jojima T, Tomizawa A, Satoh H, Hattori S, Kasai K, Hayashi T. A glucagon-like peptide-1 (GLP-1) analogue, liraglutide, upregulates nitric oxide production and exerts anti-inflammatory action in endothelial cells. *Diabetologia* 2010 October;53(10):2256-63.
- (249) Villanueva-Penacarrillo ML, Martin-Duce A, Ramos-Alvarez I, Gutierrez-Rojas I, Moreno P, Nuche-Berenguer B, Acitores A, Sancho V, Valverde I, Gonzalez N. Characteristic of GLP-1

effects on glucose metabolism in human skeletal muscle from obese patients. *Regul Pept* 2011 June 7;168(1-3):39-44.

- (250) Timmers S, Konings E, Bilet L, Houtcooper RH, van de Weijer T, Goosens GH, Hoeks J, van der Krieken S, Ryu D, Kersten S, Moonen-Kornips E, Hesselink MK, Kunz I, Schrauwen-Henderling VB, Blaak EE, Auwerx J, Schrauwen P. Calorie restriction-like effects of 30 days of resveratrol supplementation on energy metabolism and metabolic profile in obese humans. *Cell Metab* 2011;14:612-22.
- (251) Sakakibara S, Yamauchi T, Oshima Y, Tsukamoto Y, Kadowaki T. Acetic acid activates hepatic AMPK and reduces hyperglycemia in diabetic KK-A(y) mice. *Biochem Biophys Res Commun* 2006 June 2;344(2):597-604.
- (252) Sakakibara S, Murakami R, Takahashi M, Fushimi T, Murohara T, Kishi M, Kajimoto Y, Kitakaze M, Kaga T. Vinegar intake enhances flow-mediated vasodilatation via upregulation of endothelial nitric oxide synthase activity. *Biosci Biotechnol Biochem* 2010;74(5):1055-61.
- (253) Johnston CS, White AM, Kent SM. Preliminary evidence that regular vinegar ingestion favorably influences hemoglobin A1c values in individuals with type 2 diabetes mellitus. *Diabetes Res Clin Pract* 2009 May;84(2):e15-e17.
- (254) Ostman E, Granfeldt Y, Persson L, Bjorck I. Vinegar supplementation lowers glucose and insulin responses and increases satiety after a bread meal in healthy subjects. *Eur J Clin Nutr* 2005 September;59(9):983-8.
- (255) Johnston CS, Steplewska I, Long CA, Harris LN, Ryals RH. Examination of the antiglycemic properties of vinegar in healthy adults. *Ann Nutr Metab* 2010;56(1):74-9.
- (256) McCarty MF. Does regular ethanol consumption promote insulin sensitivity and leanness by stimulating AMP-activated protein kinase? *Med Hypotheses* 2001 September;57(3):405-7.
- (257) Peng L, Li ZR, Green RS, Holzman IR, Lin J. Butyrate enhances the intestinal barrier by facilitating tight junction assembly via activation of AMP-activated protein kinase in Caco-2 cell monolayers. *J Nutr* 2009 September;139(9):1619-25.
- (258) Hu GX, Chen GR, Xu H, Ge RS, Lin J. Activation of the AMP activated protein kinase by short-chain fatty acids is the main mechanism underlying the beneficial effect of a high fiber diet on the metabolic syndrome. *Med Hypotheses* 2010 January;74(1):123-6.
- (259) Tang Y, Chen Y, Jiang H, Nie D. The role of short-chain fatty acids in orchestrating two types of programmed cell death in colon cancer. *Autophagy* 2011 February;7(2):235-7.